

Beatriz Novoa & Antonio Figueras

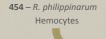


Immunology & Genomics Group Marine Research Institute (IIM-CSIC), Vigo, Spain

Bivalves

Fish

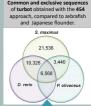
Transcriptomics: High-throughput sequence analysis



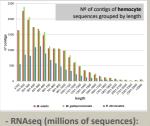








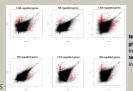
REPROSEED PROJECT:



- 454:

M. galloprovincialis – Mediterranean mussel M. edulis – Blue mussel

R. decussatus – Carpet shell clam



AQUAGENOMICS-CONSOLIDER project:

- 454 sequencing:
- S. maximus Turbot
- D. labrax Sea bass
- S. aurata Sea bream
- •Thousands of sequences included in public databases
- •Characterization and analysis of processes such as:
- feeding, metabolism, growth, behavior, immune response against pathogens...
- •Development of tools such as microarrays



M. galloprovincialis

Microarrays

Applications

- R. philippinarum challenged with V. alginolyticus (Moreira et al. 2013)







Return to

Differences between the transcriptomes of control and challenged clams against bacteria (V. alginolyticus) and parasites (Perkinsus).

- -Turbot challenged with VHSV (Diaz-Rosales et al., 2012): Study of gene expresion differences between resistant and susceptible turbot families against viral challenges.
- Changes in the transcriptome profile after vaccination and after viral infection with /without previous vaccination: which are the changes associated to protection
- Zebrafish as a model for obesity and response to bacterial and viral stimuli

- Identification of bioactive molecules:

•Caspases and apoptotic genes in mussel (Romero *et al.*, 2011): **possible biomarkers for aquatic pollution**

•Pore-forming molecules in mussel (Estévez-Calvar et al., 2011)

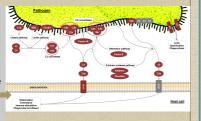
•Immune pathways inferred from the Manila clam 454 results (Moreira *et al.*, 2012): putative

"immune moleecules"

TLR signaling pathway

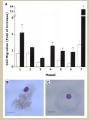
Apoptosis

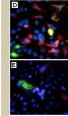
Complement cascade



-The antimicrobial peptides: highly expressed genes in bivalves:

Identification of high variable molecules in mussel: Myticin C: an antimicrobial peptide with chemotactic, antiviral and immunoregulatory properties (Vera et al., 2011;





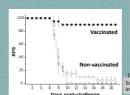
Chemotactic assay (A) of mussel hemocytes. Hemocyte immunocytochemistry after migrating to the chamber with Mytcin C containing plasmid (B) and after migrating to the control chamber (C).

Antiviral properties of Mytcin C: CHSE cells transfected with empty plasmid (D) or Mytcin C containing plasmid (E). Cells expressing mytcins are not infected with the virus.

Blue: DNA staining Green: transfected cells Red: VHSV

- Expression studies: Immunocompetence in mussel larvae

-Identification of bioactive molecules:



•Hepcidin (Pereiro et al., 2012) •WAP65 •NK lysin immunofluorescence assay:

NK lysin expressing cells do not become infected Green: SVCV infected EPC cells ge: Nk lysin expressing cells

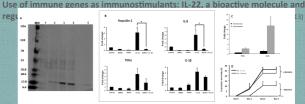


against VHSV in turbot:

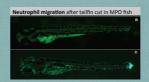
Immune pathways inferred from the turbot 454 results (Pereiro et al.

e genes to produce a higher protection induced by DNA vaccine

Use of immune genes as imp stimulants: IL-22, a bioactive molecule and key



Recombinant <u>turbot</u> IL-22 (A). The IL-22 induced inflamma the blocking of the IL-22 with morpholinos after an *in vivo A* mortality (C, D).



The zebrafish as a model for the study of i 30 reconstruction of the effect of the SVCV infection in zebrafish embryos after 24 hours. The virus induces cell death by pyroptosis and second in the properties of the study of infection in zebrafish embryos after 24 hours.

