1	From microplastics to pixels: Testing the robustness of two machine
2	learning approaches for automated, Nile red-based marine microplastic
3	identification
4	
5	Authors
6	Nelle Meyers ^{1,2,3,*} , Bavo De Witte ² , Natascha Schmidt ^{4,5} , Dorte Herzke ^{4,6} , Jean-Luc Fuda ⁵ , David Vanavermaete ² ,
7	Colin R. Janssen ^{3,7} , Gert Everaert ¹
8	*Corresponding author. E-mail address: <u>nelle.meyers@vliz.be</u>
9 10	Affiliations
11	¹ Flanders Marine Institute (VLIZ), InnovOcean Campus, Jacobsenstraat 1, 8400 Ostend, Belgium.
12	² Flanders Research Institute for Agriculture, Fisheries and Food (ILVO), Animal Sciences Unit - Aquatic Environment
13	and Quality, InnovOcean Campus, Jacobsenstraat 1, 8400 Ostend, Belgium.
14	³ Department of Animal Sciences and Aquatic Ecology, Blue Growth Research Lab, Ghent University, 9000 Gent,
15	Belgium.
16	⁴ NILU, The FRAM Centre, P.O. Box 6606, 9296 Tromsø, Norway.
17	⁵ Aix Marseille University, Toulon University, CNRS, IRD, Mediterranean Institute of Oceanography (MIO) UM 110,
18	Marseille, France.
19	⁶ Norwegian Institute for Public Health (NIPH), P.O.Box 222 Skøyen, 0213 Oslo, Norway.
20 21	⁷ Blue Growth Research Lab, Ghent University, Bluebridge, Wetenschapspark 1, 8400 Ostend, Belgium.
22	Keywords
23	Microplastics; Machine learning; Automation; Fluorescence; Nile red; Weathered plastics; Monitoring;
24	Marine pollution
25	
26	Abstract
27	
28	Despite the urgent need for accurate and robust observations of microplastics in the marine environment
29	to assess current and future environmental risks, existing procedures remain labour-intensive, especially
30	for smaller-sized microplastics. In addition to this, microplastic analysis faces challenges due to
31	environmental weathering, impacting the reliability of research relying on pristine plastics. This study
32	addresses these knowledge gaps by testing the robustness of two automated analysis techniques which

33 combine machine learning algorithms with fluorescent colouration of Nile red (NR)-stained particles. 34 Heterogeneously shaped uncoloured MPs of various polymers—polyethylene (PE), polyethylene 35 terephthalate (PET), polypropylene (PP), polystyrene (PS), and polyvinyl chloride (PVC)—ranging from 100 36 to 1000 µm in size and weathered under semi-controlled surface and deep-sea conditions, were stained 37 with NR and imaged using fluorescence stereomicroscopy. This study assessed and compared the 38 accuracy of decision tree (DT) and random forest (RF) models in detecting and identifying these weathered 39 plastics. Additionally, their analysis time and model complexity were evaluated, as well as the lower size 40 limit $(2 - 4 \mu m)$ and the interoperability of the approach. Decision tree and RF models were comparably 41 accurate in detecting and identifying pristine plastic polymers (both > 90%). For the detection of 42 weathered microplastics, both yielded sufficiently high accuracies (> 77%), although only RF models were 43 reliable for polymer identification (> 70%), except for PET particles. The RF models showed an accuracy > 44 90% for particle predictions based on 12-30 pixels, which translated to microplastics sized < 10 μ m. 45 Although the RF classifier did not produce consistent results across different labs, the inherent flexibility 46 of the method allows for its swift adaptation and optimisation, ensuring the possibility to fine-tune the 47 method to specific research goals through customised datasets, thereby strengthening its robustness. The 48 developed method is particularly relevant due to its ability to accurately analyse MPs weathered under 49 various marine conditions, as well as ecotoxicologically relevant MP sizes, making it highly applicable to 50 real-world environmental samples.

51

52

1. Introduction

53 54

55 Microplastics (MPs), plastic particles ranging from 1 μ m to 5 mm (Hartmann et al., 2019; Arthur et al., 56 2009), are a widespread and persistent pollutant, detected even in remote areas far from human activities 57 (Peeken et al., 2018; Bergmann et al., 2019; Ross et al. 2021, Van Cauwenberghe et al., 2013, Peng et al., 58 2018). Accurate and robust observations are essential for evaluating the current and future environmental 59 risks posed by MPs, however, MP analysis continues to be labour-intensive to this day (Primpke et al., 60 2020a). Indeed, analytical identification of MPs in a wide range of marine environmental matrices is a 61 critical yet challenging part of the research. Consequently, an extensive spectrum of analytical methods 62 has been developed in the last decade to meet a variety of research and monitoring purposes, based on 63 the requirements associated with research and monitoring observations of MPs. However, many of these

techniques have considerable limitations in terms of resolution, minimum particle size, human bias
interference, labour intensity and analysis time, and consequently, analysis cost (Primpke et al., 2020a).

66

67 Fluorescence microscopy, combined with fluorescent dyes, allows to effectively visualise MPs. The 68 fluorescent dye Nile red (NR) has been widely used in MP studies because of its high adsorption for 69 plastics, its affinity for a wide range of polymers, its short incubation time (10-30 min), and its 70 effectiveness to detect MPs down to a few µm (Maes et al., 2017; Shruti et al., 2022). Nile red fluorescence 71 of MPs is often imaged using fluorescence microscopes equipped with a camera, for subsequent visual or 72 automated image analysis. A major flaw of the NR approach is the co-staining of lipid-rich organic material, 73 which interferes with MP analysis (e.g. Maxwell et al., 2020; Shruti et al., 2022). However, false positives 74 resulting from this issue can be mitigated using a multiple-filter approach (Meyers et al., 2022).

75

76 The integration of automation into various MP detection and identification techniques has recently led to 77 a remarkable transformation in MP research. The application of Artificial Intelligence (AI) in the field of 78 research has revolutionised the way researchers approach MP analysis, with machine learning (ML) 79 gaining particular popularity as it enables a faster, more cost-effective, and less biased particle 80 identification (Guo et al., 2024). Numerous prediction models based on ML have been developed for the 81 automated detection and identification of MPs, where the main principles often rely on vibrational 82 spectroscopy (Lin et al., 2022). Frequently used algorithms to do so include decision trees (DTs), random 83 forests (RFs), support vector machines, K-nearest neighbours, and neural networks (Yan et al., 2022; Lin 84 et al., 2022). Automation of MP analysis methods has however introduced both opportunities and 85 challenges. Selecting the best model is data-dependent, where finding a trade-off between model 86 performance, model complexity and computational speed is crucial (Maxwell et al., 2018). Machine 87 learning programs can process large amounts of data with a high accuracy but can take up a lot of time to 88 do so, especially when limited computing power is available, making it costly. For example, Focal Plane 89 Array (FPA) array-based μ -FTIR images combined with a spectroscopic analysis pipeline allows for the 90 automated comparison of MP polymers and their unique infrared spectra with spectral libraries using 91 pattern recognition algorithms, but overall analysis time is still relatively high (Primpke et al., 2017). For 92 instance, it may take 4 h to scan 14 x 14 mm with a pixel resolution around 11 μ m (64 x 64 FPA detector 93 elements) (Bergmann et al., 2019; Primpke et al., 2020a), while an additional 4 – 48 h is needed for spectral 94 analysis by spectral correlation (Peeken et al., 2018; Primpke et al., 2020b; Primpke et al., 2020a). A 95 compromise between model complexity and model performance also exists: although they often perform

better, more complex ML algorithms such as RFs lack transparency as they are much harder to interpret,
especially for non-experts in the field of AI (Breiman et al., 1993; Breiman et al., 2001; Witten et al., 2002).
Simpler models like DTs are more intuitive and easier to interpret due to their transparency. However,
they are less effective for complex or noisy datasets and are more prone to overfitting, which occurs when
a ML model performs significantly better on training data than it does on new datasets (Breiman et al.,
1993; Breiman et al., 1996; Witten et al., 2002).

102

103 Microplastics in the marine environment undergo weathering, e.g. due to mechanical forces, UV radiation, 104 microbial colonisation, and hydrostatic pressure, leading to alterations in their physical and chemical 105 structure (Fotopoulou et al., 2019; Shah et al., 2008; Fauvelle et al., 2021). These changes can complicate 106 their detection and analysis (Dong et al., 2020; Liu et al., 2020), underscoring the need to consider 107 weathered MPs in method development, effect studies, and leaching experiments, along with their 108 pristine representatives. Standard Raman and IR spectra of MPs were shown to be significantly impacted 109 by environmental weathering processes through shifts in their spectra (Dong et al., 2020) which in turn 110 can interfere with the spectral matching process. Shifts in crystallinity and polarity (Maes et al., 2017) can 111 also impact the fluorescence of NR-stained plastics due to the solvatochromism of the dye, i.e. its 112 fluorescence changes based on the polarity of its environment, in this way affecting the accuracy of the 113 approach.

114

115 At present, the bulk of method development-focused research and effect studies still relies on pristine 116 MPs (Waldman and Rillig, 2020; Alimi et al., 2022), yet recent studies suggest that aged plastics behave 117 differently (Arp et al., 2021; Bhagat et al., 2022). This underlines the importance of implementing 118 environmentally relevant MPs into future MP research for more accurate findings. To tackle this problem, 119 MP structural and chemical transformations brought on by weathering processes are nowadays being 120 studied during laboratory simulations. However, most research focuses only on a few ageing processes 121 when mimicking environmentally relevant conditions (Alimi et al., 2022), while the degradation of MPs in 122 the marine environment is influenced by fluctuations in weathering processes and abiotic factors like 123 seawater temperature, salinity and hydrostatic pressure. In addition, degradation is affected by the 124 polymer type, size, structure, shape, and density of MPs. The complex interplay between these variables 125 makes it difficult to realistically simulate MP degradation in a laboratory setting.

127 There is still much to uncover regarding the abundance, behaviour, and potential effects of smaller-sized 128 MPs (< 100 μ m). While methods capable of reaching this MP size threshold exist, the time and costs 129 involved in analysis hinder routine assessments, emphasising the need for cost-efficient and high-130 throughput analysis methods. Moreover, smaller-sized MPs are an emerging concern in the field of 131 ecotoxicology (Beiras et al., 2020). A smaller size translates into an increased surface-to-volume ratio of 132 MPs, which renders them more bioavailable (Mattsson et al., 2015, Wagner and Reemtsma, 2019). Their 133 size can also accelerate physicochemical and biochemical reactions at their surface (Wayman et al., 2021). 134 This disparity between current analysis methodologies and ecotoxicologically relevant MP sizes stresses 135 the existing gap in our understanding of the true extent and impact of MP contamination in the marine 136 environment. Addressing this issue is therefore imperative for a comprehensive assessment of MP 137 pollution.

138

A last significant challenge MP research faces is the interoperability of analysis methods. Ensuring a constant performance of a method across different laboratories, regardless of the diverse laboratory conditions, is of paramount importance in scientific research. Consistency across various laboratories strengthens the credibility of obtained findings using that method, creates a sense of confidence in the scientific community, and hence encourages the widespread adoption and application of a method. This in turn contributes to a more comprehensive understanding and management of MP pollution in marine ecosystems.

146

147 Despite the urgent need for accurate and robust observations of MPs to assess environmental risks, many 148 existing methods are labour-intensive and costly and may not always guarantee consistent performance 149 across different laboratories. Moreover, current research often relies on pristine plastics, overlooking the 150 complexities introduced by environmental weathering, which can alter the physical and chemical 151 properties of MPs and impact analytical reliability. Additionally, there is a significant gap in cost-efficient 152 methods for analysing smaller, ecotoxicologically relevant MPs. In response to the current shortcomings, 153 this research focused on testing the robustness of two recently developed, automated, Nile-red (NR) 154 based MP analysis techniques, created using two different ML algorithms. To do so, their ability to 155 accurately analyse MPs weathered under semi-controlled surface water and deep-sea water conditions 156 was assessed and compared, along with their analysis time and model complexity. Additionally, the size 157 limit of the overall best performing technique was determined, i.e. the minimum particle size for which 158 the model algorithm produces sufficiently accurate predictions in terms of plastic identity and polymer

type. Finally, we tested whether the knowledge rules generated by this classifier produce consistent results across different labs. The novelty of the developed method lies in its broad applicability, as it covers a diverse range of plastic polymers, weathering conditions, and instrumentation types, providing a comprehensive tool for advancing MP research. By verifying the robustness of the models, we assess their reliability for widespread adoption and application, which serves as a cornerstone for a comprehensive understanding and effective management of MP pollution within a marine context.

- 165
- 166

2. Materials and Methods

167

168 This work used an open-source approach that combines NR-stained particle fluorescence with machine 169 learning models, following a comprehensive six-step protocol. First, two sets of Red, Green, and Blue 170 (RGB) colour datasets were created using two types of microscopes: a fluorescence stereomicroscope 171 (FSM) and a fluorescence microscope (FM). Each set had two datasets for training a 'Plastic Detection 172 Model' (PDM) to classify particles as plastic or non-plastic, and a 'Polymer Identification Model' (PIM) to 173 classify plastic particles by polymer type (Fig. 1 - step 1). This was done as per Meyers et al 2022 and 174 Meyers et al., 2024a, where RGB-colour values extracted from pixels of particles photographed with a 175 fluorescence microscope were used to generate RGB statistics that make up the datasets, which in turn 176 were used to train models and make predictions of a particle's identity based on its RGB statistics. The 177 first set of RGB datasets, constructed at the laboratory of the Flanders Research Institute for Agriculture, 178 Fisheries and Food (ILVO set with ILVO datasets 1 and 2) were used to generate and validate a total of five 179 ILVO PDMs and five ILVO PIMs, and this using two types of ML algorithms, i.e. a decision tree (DT) and a 180 random forest (RF) classifier (Fig. 1 - step 1). To do so, the datasets were split into 80% training data and 181 20% test data.

182

183 Secondly, the average number of correctly classified instances (CCI) + standard deviation (SD) (%) for a 184 subset of particles unknown to the models (test datasets) was calculated for each of the models, and 185 compared for both classifiers: plastic/non-plastics for the PDMs, and polymer type for the PIMs. Cohen's 186 Kappa statistic, used to compare observed accuracies with expected accuracies, was also calculated (Fig. 187 1 - step 2). Running five simulations per model (PDM/PIM) for each classifier enhanced the robustness 188 and reliability of the evaluation process by mitigating the influence of random variations or chance 189 occurrences on model performance. Thirdly, after also comparing their analysis time and model 190 complexity (cfr. '2.2 Model construction and classifier comparison') for the overall best scoring model algorithm, the average accuracy of the ILVO models based on the best performing ML algorithm was
tested for the detection and identification of MPs weathered at sea under semi-controlled, surface water
and deep-sea water conditions for a duration of 12 months using five simulations per model type (Fig. 1 step 3).

195

196 As a fourth step, a second set of models was constructed and validated, using the second set of RGB 197 datasets constructed at the laboratory of the Flanders Marine Institute (VLIZ set with VLIZ datasets 1 and 198 2), which comprised RGB data from images acquired at a higher magnification (VLIZ PDM and VLIZ PIM) 199 (Fig. 1 - step 4) To do so, the existing RGB datasets from Meyers et al., 2022 were expanded. In the fifth 200 step, the lower size limit of the developed approach was determined (cfr. '2.3 Lower size limit') using the 201 VLIZ models with the highest magnification (Fig. 1 - step 5). In a sixth and last step, the interoperability of 202 the approach was tested by evaluating the average performance of the ILVO models for the VLIZ datasets 203 1 and 2, and the performance of the VLIZ models for the ILVO datasets 1 and 2, based on five simulations 204 per model type (Fig. 1 - step 6).

205



- Fig. 1. Schematic overview. A schematic overview of the comprehensive, six-step approach employed in this work, where the
 robustness of a microplastic analysis method based on machine learning models combined with fluorescent colouration of NR stained particles was thoroughly tested.
- 210
- 211 <u>2.1 Construction of datasets</u>
- 212 ILVO datasets 1 and 2 were based on images series acquired with a fluorescence stereomicroscope (Leica
- 213 M205 FA Fluorescence stereomicroscope LAS X software), at a magnification of 1 x 10, while images

214 series for the two VLIZ datasets were acquired with a fluorescence microscope (LEICA DM 1000 - Leica 215 Application Suite version 4.13.0), at a 10 x 10 magnification (Fig. 1) (Meyers et al., 2024b). To generate 216 the RGB datasets for the ILVO PIM and the VLIZ PIM (as described in detail in Meyers et al., 2022), five 217 and seven of the most abundantly produced MP polymers globally, respectively, as well as organic 218 materials with high prevalence in the marine environment were selected. First, uncoloured and pristine 219 polyethylene (PE), polyethylene terephthalate (PET), polypropylene (PP), polystyrene (PS), and polyvinyl 220 chloride (PVC) were obtained and cryomilled into heterogeneously shaped particles (50 - $1,200 \mu m$) by 221 specialized companies (Centexbel - Zwijnaarde, Belgium, and CARAT GmbH - Bocholt, Germany). The 222 organic materials cotton, flax (raw and bleached), hemp (raw and bleached), silk, and wool (alpaca and 223 sheep) were also obtained from Centexbel, while chitin, wood, and gull feathers were collected from the 224 beach and cut into similarly sized particles. In the next step, less than 0.5 mg of each material (the different 225 plastic polymers/organic material in general) was added to separate glass beakers containing Milli-Q 226 water. To test the ability of the models to correctly classify unknown polymers as plastics, for the VLIZ 227 PIM, two additional polymers were used, i.e. nylon and PUR particles. As PE and PP are similar with respect 228 to their fluorescence as a consequence of their similar chemical structure (Meyers et al., 2022), it was 229 decided to group them into one class to construct the ILVO PIM ('PE/PP'). All MPs used were pristine, 230 heterogeneously shaped uncoloured fragments, with varying densities (Gago et al, 2019). Next, the 231 content of each beaker was filtered over a PTFE-filter (47 mm diameter, 10 µm pore size, Millipore Ltd.) 232 using a MilliPore manifold system (3 + 3 workstations) (Merck Millipore) and stained with 1 mL of the 233 fluorescent dye Nile red dissolved in acetone (10 µg mL⁻¹) using a glass pipette. After 15 min, filters were 234 rinsed with Milli-Q water and left to dry in a dark environment for 24 h.

235

236 Particles were photographed with both types of microscopes using fixed settings, capturing a series of 3 237 images per particle under a blue, green, and UV microscope filter (cfr. Table S1 for details). Red, Green, 238 and Blue colour data from 135 (ILVO PIM) and 200 (VLIZ PIM) particles per plastic polymer type, or per 239 group of plastics (PE/PP), were analysed to construct ILVO dataset 2 and VLIZ dataset 2 (n = 540 and n = 240 1400). A random subselection of MP particles, evenly spread across all polymers, was used to construct 241 ILVO dataset 1 and VLIZ dataset 1, complemented with RGB data from 420 (ILVO PDM) and 500 (VLIZ 242 PDM) particles of natural origin (n = 840 (2×420) and n = 1000 (2×500)). For the VLIZ datasets, 15 - 50 243 images for each of the material types (i.e. all individual polymers, and a mix of organic materials) were 244 used to build the datasets. Three PTFE filters per material type were used, acquired over three days. For 245 the ILVO datasets, 6 images per material type, capturing an entire PTFE filter, were used. Six PTFE filters

per material were employed, acquired over three days. All dataset sizes allowed for an appropriate statistical power for a small effect size ($\alpha = 0.05$, power = 0.8) (Serdar et al., 2021). Required training dataset sizes of all four models for which prediction error was below 5% were also simulated using the randomForest package in R (Fig. S1) (Liaw & Wiener, 2002).

250

251 Once the images were acquired, extraction of the RGB values from particle pixels was done through 252 automated image analysis in the open-source image processing program ImageJ (Abramoff et al., 2004) 253 according to the updated procedure described in Meyers et al., 2024a. For each particle above a set size 254 limit within each image series, a CSV file was generated with extracted RGB values of all pixels lying on 255 the maximum Feret diameter of that particle, and this for the image acquired under the blue, green and 256 UV filter. Next, per particle, for each microscope filter, statistics were calculated (10th, 50th and 90th 257 percentile as well as the mean value) for each of the three colour values (R, G, and B) in R (R Core Team, 258 2020). All 36 RGB statistics per particle were then automatically compiled into the above-mentioned 259 datasets (cfr. Meyers et al., 2022 for method details).

260

261 <u>2.2 Model construction and classifier comparison</u>

262 For both classifiers (DT and RF), the generated RGB statistics were used as input variables, while the 263 output was particle identity (plastic/non-plastic for the PDM and polymer type for the PIM). Decision tree 264 classifiers create flowchart-like trees, where each internal node represents a test of a feature, the 265 branches represent outcomes of the tests, and leaf nodes signify the output of the algorithm. The datasets 266 were recursively split into smaller datasets based on the values of the features, i.e. the RGB statistics. The 267 CART (Classification and Regression Tree) supervised ML algorithm (Therneau et al., 2015) was used to 268 generate the DT classifiers (Meyers et al., 2022), (five simulations per model type) where, for the PDM 269 and PIM, respectively, a minimum of 8 records had to be present in a node before a split was attempted, 270 as well as a minimum of 3 records in an end node, and splits that did not enhance the model fit by 0.02 271 were eliminated. To avoid overfitting, the model was allowed to grow until a depth where the validation 272 metrics used either stagnated or declined.

273

274 RF classifiers, on the other hand, average the output of multiple DTs fit on random subsamples (63.2%) of 275 the dataset, and this to obtain a single result (bootstrap aggregating or bagging). Breiman's RF algorithm 276 (Breiman, 2001; Liaw & Wiener, 2002) was used to construct the RF classifiers here, where a fitting 277 number of variables (cfr. Fig. S2 "optimal number of parameters") were chosen each time at random out of all predictor variables, and the best split on these variables was used to divide the node. For each tree,
the leftover data (36.8%) was then used to calculate the misclassification rate, i.e. the out of bag (OOB)
error rate. The aggregated error, which determines the overall OOB rate of the classifier, was plotted for
each model as a validation. To fine-tune the RF models, the number of trees to achieve the most accurate
results was also determined (Fig. S3). The final, optimised models all had 500 prediction trees (ntree),
each using max. 12 variables (mtry).

284

285 To visualise classification performance of all four models, non-metric multidimensional scaling (NMDS) 286 plots using the Bray-Curtis similarity index and default settings were generated in R (Fig. 2, Fig. S4) based 287 on one of the five simulations for each model type. For both classifiers, the best features to split the data 288 were based on the Gini impurity metric (Daniya et al., 2020). The accuracy of all models using both 289 classifiers was assessed based on their CCI rate, their Cohen's kappa statistic (McHugh, 2012), which is a 290 measure of interrater reliability, and their confusion matrices. For the DTs, this was done by randomly 291 selecting 4/5 of each dataset to train each of the five models per model type and keeping the remaining 292 1/5 as a validation dataset based on the Pareto principle ratio (Dunford et al., 2014). For the RFs, five-fold 293 cross-validations were performed to estimate the skill of the models on unseen data by re-training the 294 models on the same design. Here, each dataset was split into five randomly chosen equal folds (each 295 containing 1/5 of the dataset). Validation iterated over these folds, using each as a unique test dataset 296 while training the model on the remaining four. Additionally, model training duration, analysis time, and 297 complexity were assessed and compared for both classifier types. Furthermore, when the average overall 298 model accuracy was < 70%, the model was considered unfit for MP analysis.

299

300 RGB datasets derived from pristine MPs were used to construct all models. While the conventional 301 approach may be to train models on datasets developed using weathered plastics, logistical constraints 302 underscore the challenge associated with this approach (discussed under '4.4 Interoperability'). While 303 conducting an extensive weathering of various polymers under environmentally relevant conditions as 304 done here is relevant and feasible (cfr. '2.3 Natural weathering of microplastics'), it implies that models 305 could only be built once weathering is finalised. Therefore, the initial strategy focused on assessing the 306 performance of RF models trained on pristine materials when applied to weathered MPs, providing a 307 pragmatic foundation for subsequent investigations.





Fig. 2: Non-metric multidimensional scaling plots of the training datasets. The non-metric multidimensional scaling (NMDS)
 plots highlight the differences and similarities in fluorescent colouration of all Nile red-stained particles used to train the four
 models: from top to bottom, left to right: ILVO Plastic Detection Model (PDM), VLIZ PDM, ILVO Polymer Identification Model
 (PIM), VLIZ PIM.

316 <u>2.3 Natural weathering of microplastics</u>

317 To test the robustness of the classifiers, their ability to correctly classify naturally weathered plastics was 318 tested. A mix of MP fragments (PE, PET, PP, PS, and PVC) sized 500 - 1000 µm (CARAT GmbH) was filled 319 into stainless steel tubes (316 mesh) (Inoxia Ltd, 45.7 Dunsfold Park, Stovolds Hill, Cranleigh, GU6 8TB, 320 United Kingdom), which were placed in stainless steel containers (Fig. 3). The containers were deployed 321 in subsurface, coastal waters in the Norwegian Sea (Tromsø, Norway; 69.642730, 18.950389) in February 322 2021 and in deep-sea waters (2380 m depth) of the Mediterranean Sea (off the coast of Marseille, France; 323 42.807683, 6.043867) (Fig.3 and Fig. S5) in April 2021, respectively. During the exposure at sea, the 324 stainless-steel containers deployed in subsurface waters were manually cleaned at regular intervals to 325 remove bivalves and other attached organisms. The samples remained submerged in the sea for 12 326 months before being retrieved. Afterward, the stainless-steel tubes were left under a fume hood to dry. 327 Next, the microplastic particles were transferred into burnt (450 °C, 6 h) glass vials and brought to the 328 laboratory.

329

330 Following this, for each type of weathering, polymer particles were stained under the previously described 331 conditions and photographed under the fluorescence stereomicroscope using the same settings as for 332 pristine particles (Table S1). Hereafter, the average predictive performance of the FSM PDM and FSM PIM 333 was tested using a random selection of 30 particles per polymer (PE/PP, PET, PS and PVC), for each type 334 of weathering, totaling 240 particles. Additionally, ATR-FTIR spectra of each studied polymer were 335 acquired in its pristine, surface water-weathered, and deep-sea water-weathered form (Perkin Elmer 336 Frontier FTIR with UATR top plate with diamond/ZnSE crystal, Zaventem, Belgium). For clarity, deep-sea 337 water-weathered (DSW) MPs and surface water-weathered (SFW) MPs will be referred to as DSW MPs 338 and SFW MPs, respectively.



Fig. 3a, 3b and 3c. Controlled natural weathering of microplastics. Stainless steel tubes (3b) were filled with a microplastic
 mixture and placed inside stainless steel containers. These containers were then submerged in subsurface coastal waters of the
 Norwegian Sea near Tromsø, Norway (3a), or in the deep sea off the coast of Marseille, France, in the Mediterranean Sea, for a
 duration of 12 months. For the deep-sea deployment, the containers were mounted on a carousel (3c), which was retrieved after
 weathering using an acoustic release system (Fig. S5).

346

347 2.4 Lower size limit

348 To determine the lower size limit of the two best performing models, for PE, PS, PET and PVC, a small 349 amount of MPs (<0.5 mg) sized < 5 μ m - 300 μ m was added to a glass beaker with Milli-Q water, whose 350 content was then filtered and NR-stained (cfr. '2.1 Construction of datasets'). One filter per polymer was 351 obtained and photographed using the fluorescence microscope, which offers the largest magnification of 352 both microscope types (i.e. 10 x 10). Next, all MPs present were analysed using the VLIZ PDM and PIM, 353 and the number of pixels per MP used to predict the particle identity was assessed. A total of 2010 PET 354 particles, 3939 PE/PP particles, 1596 PS particles and 2533 PVC particles were analysed and subdivided 355 into 10 different size groups (Table S2), representing all particles present on the PTFE filter per polymer. 356 Following this, a graph was constructed to plot predictive accuracy per size class for the PDM and PIM. 357 The lower size limit for each polymer type was determined as the smallest number of pixels and 358 corresponding size class for which both models achieved an accuracy > 70 %.

- 359
- 360
- 361

362 <u>2.5 Interoperability</u>

To test the interoperability of the analysis method, MPs imaged with the Leica M205 FA Fluorescence stereomicroscope were analysed using models trained on images from the LEICA DM 1000, and vice versa. In this process, all particles of datasets 1 and 2 from the VLIZ laboratory were analysed using models trained on datasets 1 and 2 from the ILVO laboratory, respectively, and vice versa.

367

368 <u>2.6. QA/QC</u>

As background control measures to prevent MP contamination, the use of plastic materials was avoided prior to the image acquisition, tools and glassware used were pre-cleaned with Milli-Q water and soap before use, sample processing was conducted in a laminar flow hood, and a 100% cotton lab coat and trousers were worn. Additionally, a subset of pristine MPs to construct the models as well as the weathered MPs were FTIR validated (cfr. Methods Supplement).

374

375 **3. Results**

376

377 <u>3.1 Classifier comparison for pristine materials</u>

378 DT and RF classifiers both had high accuracies (> 90% for all models) based on the obtained CCI rates and 379 Cohen's kappa statistics (Table 1). Fluorescent colouration differed between plastic and organic particles, 380 as well as between most polymers, except for PP and PE (VLIZ PIM), as is apparent from the NMDS plots 381 (Fig. 2, Fig. 4, Fig. S4, Fig. S6). The PDMs based on both classifiers misclassified only a negligible number 382 of organic and plastic materials. The PIM built using the DT approach misidentified PS particles as PE (4.4 383 \pm 9.9%), PET (2.2 \pm 2%), and PVC (2.2 \pm 3.3%). Additionally, it misclassified PE as PS (3 \pm 3.1%), PET particles 384 as PVC (1.5 \pm 2%), and PVC particles as PET (1.5 \pm 2%) and PS (1.5 \pm 2%). When using the RF approach, 385 classification errors primarily involved PE/PP particles being classified as PS (0.7 ± 0%), PET as PS (0.7 ± 386 0%), PS as PVC (1 \pm 0.4%), and PVC as PS (0.7 \pm 0%). For both classifiers, SDs were low and relatively 387 constant, indicating minimal variability among the five obtained accuracies for each of the models. Based 388 on model performances in Table 1, the accuracy of DT and RF classifiers was relatively similar for pristine 389 materials.

390

391

Table 1. Predictive accuracy of decision tree (DT) vs. random forest (RF) classifiers for pristine microplastics. Predictive
 accuracy of the Plastic Detection Model (PDM) and the Polymer Identification Model (PIM) for pristine MPs and organic
 material, based on DT and RF classifiers, using images acquired with a fluorescence stereomicroscope (FSM) (ILVO datasets 1
 and 2).

397		DECISION TREE CLASSIFIER		RANDOM FOREST CLASSIFIER	
398	PRISTINE PLASTICS				
399	FSM (ILVO)	PDM	PIM	PDM	PIM
400	CCI %	99.3 ± 1.1%	95.9 ± 2.3%	100 ± 0%	99 ± 0.5%
401	Cohen's к	1	0.98	1	1
402	PE/PP		97 ± 3.1%		99.26 ± 0%
403	PET		98.5 ± 2%		99.26 ± 0%
404	PS		91.1 ± 8.1%		97.93 ± 1%
405	PVC		94 ± 1.7%		99.26 ± 0%
406	Organic material	99.8 ± 0.5%		100 ± 0%	
407					

408

409 <u>3.2 Classifier comparison for weathered MPs</u>

410 When considering the average predictive reliability of the ILVO models for weathered MPs, model 411 performance was very similar for DT and RF classifiers across different types of weathering (77.2 ± 38% 412 vs. $78.8 \pm 36\%$ for SFW particles, and $86.8 \pm 20.2\%$ vs. $87.8 \pm 17.1\%$ for DSW particles, respectively) (Table 413 2). For SFW MPs, misclassifications were primarily PET particles classified as organic material (CCI rate of 414 20.7 ± 8.3% for DTs and 25.3 ± 1.8% for RFs). For DSW MPs, the main misclassifications involved PE/PP 415 particles identified as organic material (CCI of 57.3% ± 8.3% and 63.3 ± 0%). Average accuracies were 416 substantially higher for SFW particles when PET particles were excluded from the analysis, for both DT 417 and RF classifiers (96 ± 6.9% vs. 96.7 ± 5.8%, respectively). In contrast, for DSW particles, excluding PET 418 had a negligible effect on accuracy ($85.8 \pm 24.6\%$ vs. 87.6 ± 21 , respectively).

419

420 While the predictive accuracy of the PIM remained high for polymers such as PE/PP (SFW: 95.3 \pm 3% and 421 DSW: 98 \pm 1.8%) and PS (only SFW: 89.3 \pm 23.9%) using the DT classifier, PET and PVC showed substantial 422 misclassification rates. For SFW and DSW MPs, respectively, PET had a CCI rate of 11.3 \pm 7.7% and 36.7 \pm 423 22.9%, while PVC had a CCI rate of 58.7 \pm 14.3% and 57.3 \pm 27.6%. Moreover, the predictive accuracies 424 for PS showed a relatively large variability (SD of 23.9%), indicating inconsistent performance of the DT 425 classifier for this polymer type.

427 When assessing the overall PIM performance, the RF classifiers outperformed the DTs. The average 428 accuracies for RFs were 70 \pm 39.2% for SFW MPs and 80.3 \pm 14.8% for DSW MPs, compared to 63.7 \pm 429 38.4% and 60.8 ± 26.3% for DTs, respectively. Similar to the PDM, the average accuracy for SFW particles 430 substantially increased, and the SD decreased when PET particles were excluded (88.9 ± 12.8%). For DSW 431 particles, the accuracy remained relatively consistent (84 ± 15.7%). For the DT classifiers, only the average 432 predictive accuracy of the PDM exceeded the 70% threshold. In contrast, both the PDM and PIM exceeded 433 this threshold when using the RF classifiers. The RF models were hence considered most reliable for the 434 accurate identification of MP polymers, except for the analysis of SFW PET particles (accuracy of 25.3 ± 435 1.8%).

436

The fluorescent colouration and intensity of pristine and SFW PET particles differed considerably compared to other polymers, particularly under the blue filter, which is crucial for some model parameters (Fig. 4 and Fig. S7). Attenuated Total Reflectance (ATR) spectra of each polymer, acquired in their pristine, surface water-weathered, and deep-sea water-weathered forms, are freely available in the open-access repository Marine Data Archive (Meyers et al., 2024c), and are visualised in Fig. S8 - S12.

442

The ILVO PDMs and PIMs using DT classifiers consisted of five single trees, each based on different training datasets. The PDMs were pruned to a depth of 1, and had a total of 2 leaf nodes, while the PIMs were pruned to a depth of 5 to prevent overfitting, with a total of 9 leaf nodes, including multiple PE, PET, and PS and nodes. In contrast to these simpler models, the RF-based ILVO PDMs and PIMs required 100 trees each to produce accurate results, respectively (Fig. S3), with tree depths ranging from 1 - 5. Although model complexity increased, the computational time for generating predictions was < 10 s for both the DT and RF approaches.

- 450
- 451
- 452
- 453
- 454
- 455
- 456

Table 2A (upper) and 2B (lower): Predictive accuracy of decision tree (DT) (2A) vs. random forest (RF) (2B) classifiers for
 weathered microplastics. Predictive accuracy of the Plastic Detection Model (PDM) and the Polymer Identification Model (PIM)
 for surface water-weathered MPs and deep-sea water-weathered MPs using DT and RF classifiers.

	SURFACE WATE	LK	DEEP-SEA WATER	
	WEATHERED MPS		WEATHERED MPS	
DECISION TREE				
CLASSIFIER	PDM	PIM	PDM	PIM
FSM (ILVO)				
CCI %	99.3 ± 1.1%	95.9 ± 2.3%	99.3 ± 1.1%	95.9 ± 2.3%
Cohen's κ	1	0.98	1	1
Total number of				
tested MPs	//.2 ± 38.1%	63.7±38.4%	86.8 ± 20.2%	60.8 ± 26.3%
PE/PP	88 ± 3%	95.3 ± 3%	57.3 ± 8.3%	98 ± 1.8%
PET	20.7 ± 8.3%	11.3 ± 7.7%	90 ± 4.1%	36.7 ± 22.9%
PS	100 ± 0%	89.3 ± 23.9%	100 ± 0%	51.3 ± 25.3%
PVC	100 ± 0%	58.7 ± 14.3%	100 ± 0%	57.3 ± 27.6%
	SURFACE WAT	FER	DEEP-SEA WATER	
	SURFACE WAT WEATHERED N	rer MPS	DEEP-SEA WATER WEATHERED MPS	
RANDOM FOREST	SURFACE WAT	TER MPS	DEEP-SEA WATER WEATHERED MPS	
RANDOM FOREST CLASSIFIER	SURFACE WAT WEATHERED N	rer MPS PIM	DEEP-SEA WATER WEATHERED MPS PDM	РІМ
RANDOM FOREST CLASSIFIER FSM (ILVO)	SURFACE WAT WEATHERED N	rer Mps PIM	DEEP-SEA WATER WEATHERED MPS PDM	PIM
RANDOM FOREST CLASSIFIER FSM (ILVO)	SURFACE WAT WEATHERED N PDM 100 ± 0%	rer MPS PIM 99 ± 0.5%	DEEP-SEA WATER WEATHERED MPS PDM 100 ± 0%	PIM 99 ± 0.5%
RANDOM FOREST CLASSIFIER FSM (ILVO) CCI % Cohen's к	SURFACE WAT WEATHERED M PDM 100 ± 0% 1	FER MPS PIM 99 ± 0.5% 0.98	DEEP-SEA WATER WEATHERED MPS PDM 100 ± 0% 1	PIM 99 ± 0.5% 1
RANDOM FOREST CLASSIFIER FSM (ILVO) CCI % Cohen's κ	SURFACE WAT WEATHERED N PDM 100 ± 0% 1 78.8 ± 36.0%	FER MPS PIM 99 ± 0.5% 0.98 70.0 ± 39.2%	DEEP-SEA WATER WEATHERED MPS PDM 100 ± 0% 1	PIM 99 ± 0.5% 1 80.3 ± 14.8%
RANDOM FOREST CLASSIFIER FSM (ILVO) CCI % Cohen's κ Total number of tested MPs	SURFACE WAT WEATHERED N PDM 100 ± 0% 1 78.8 ± 36.0% (96.7 ± 5.8%	FER MPS PIM 99 ± 0.5% 0.98 70.0 ± 39.2% (88.9 ± 12.8%	DEEP-SEA WATER WEATHERED MPS PDM $100 \pm 0\%$ 1 $87.8 \pm 17.1\% (87.6 \pm 21)$	PIM 99±0.5% 1 80.3±14.89 (84±15.7
RANDOMFORESTCLASSIFIERFSM (ILVO)CCI %Cohen's κTotal number oftested MPs	SURFACE WAT WEATHERED N PDM 100 ± 0% 1 78.8 ± 36.0% (96.7 ± 5.8% without PET)	FER MPS PIM 99 ± 0.5% 0.98 70.0 ± 39.2% (88.9 ± 12.8% without PET)	DEEP-SEA WATER WEATHERED MPS PDM 100 ± 0% 1 87.8 ± 17.1% (87.6 ± 21 without PET)	PIM 99 ± 0.5% 1 80.3 ± 14.8% (84 ± 15.7 without PET
RANDOMFORESTCLASSIFIERFSM (ILVO)CCI %Cohen's κTotal number oftested MPsPE/PP	SURFACE WAT WEATHERED N PDM 100 ± 0% 1 78.8 ± 36.0% (96.7 ± 5.8% without PET) 90 ± 0%	FER MPS PIM 99 ± 0.5% 0.98 70.0 ± 39.2% (88.9 ± 12.8% without PET) 99.3 ± 1.5%	DEEP-SEA WATER WEATHERED MPS PDM 100 ± 0% 1 87.8 ± 17.1% (87.6 ± 21 without PET) 63.3 ± 0%	PIM 99±0.5% 1 80.3±14.8% (84±15.7 without PET 100±0%
RANDOMFORESTCLASSIFIERFSM (ILVO)CCI %Cohen's κTotal number oftested MPsPE/PPPET	SURFACE WAT WEATHERED N PDM 100 ± 0% 1 78.8 ± 36.0% (96.7 ± 5.8% without PET) 90 ± 0% 25.3 ± 0.5%	FER MPS PIM 99 ± 0.5% 0.98 70.0 ± 39.2% (88.9 ± 12.8% without PET) 99.3 ± 1.5% 86.7 ± 0%	DEEP-SEA WATER WEATHERED MPS PDM $100 \pm 0\%$ 1 $87.8 \pm 17.1\% (87.6 \pm 21 without PET)$ $63.3 \pm 0\%$ $88.7 \pm 0.5\%$	PIM 99 ± 0.5% 1 80.3 ± 14.8% (84 ± 15.7 without PET 100 ± 0% 69.3 ± 1.5%
RANDOM CLASSIFIERFORESTFSM (ILVO)CCI %Cohen's κTotal number of tested MPsPE/PPPET PS	SURFACE WAT WEATHERED N PDM $100 \pm 0\%$ 1 $78.8 \pm 36.0\%$ (96.7 ± 5.8% without PET) 90 ± 0% 25.3 ± 0.5% 100 ± 0%	FER MPS PIM 99 ± 0.5% 0.98 70.0 ± 39.2% (88.9 ± 12.8% without PET) 99.3 ± 1.5% 86.7 ± 0% 92.7 ± 1.5%	DEEP-SEA WATER WEATHERED MPS PDM $100 \pm 0\%$ 1 $87.8 \pm 17.1\% (87.6 \pm 21 without PET)$ $63.3 \pm 0\%$ $88.7 \pm 0.5\%$ $100 \pm 0\%$	PIM 99±0.5% 1 80.3±14.8% (84±15.7 without PET 100±0% 69.3±1.5% 83.3±7.8%



495

Fig. 4. Stereomicroscopic images of pristine and weathered microplastics. Images of four Nile red-stained, commonly produced
 microplastic polymers, acquired with a fluorescence stereomicroscope, under a blue, green and UV filter, and used by the models
 to predict a particle's plastic identity, and to identify its polymer type based on fluorescence colouration and intensity. In this
 figure, pristine microplastics per polymer type are shown, as well as surface water-weathered microplastics and deep-sea water weathered microplastics.

502 <u>3.3 Lower size limit</u>

503 The VLIZ PDM and PIM both achieved sufficient accuracy (> 70%) for the lowest size class tested across all 504 polymers (Fig. S13, Table S2). This class corresponds to particles with a maximum Feret diameter of 6 to 505 30 pixels, meaning that predictions were based on the same number of RGB statistics. Under the 506 magnification used, this corresponds to particles < 10 μm. After conversion, the smallest accurately identified particles were 2 μm (6 pixels) for PET, 4 μm (12 pixels) for both PE/PP and PS, and 4 μm (12
pixels) for PVC. The lower size limit was determined by the smallest particle present on the respective
PTFE filters (Table S2).

510

511 <u>3.4 Interoperability</u>

512 To assess the interoperability of the models, the ILVO models were tested on VLIZ datasets 1 and 2, and 513 vice versa. Both models designed for plastic detection demonstrated a robust ability to identify nearly all 514 plastic particles accurately when using the optimised in-lab method, achieving CCI rates of $100 \pm 0\%$ (VLIZ 515 models) and 96.9 ± 0.1% (ILVO models). However, the ILVO PDM misclassified certain plastics such as 516 nylon (CCI rate of 98.6 \pm 0%) and PUR (80 \pm 0.8%), which were unknown to the model (Table 3). Despite 517 the high accuracy in plastic detection, the models showed a significant risk of overprediction. The ILVO 518 models correctly identified only $58.7 \pm 0.2\%$ of organic materials photographed in the VLIZ laboratory, 519 while the VLIZ models achieved a mere $0.8 \pm 0.4\%$ accuracy for organic materials photographed in the 520 ILVO laboratory.

521

522 While the models performed well for polymers like PE/PP (with accuracies ranging from 66.2 \pm 4.6% to 523 100 \pm 0%), they were less accurate for other polymers such as PET, PS, and PVC when used with different 524 instrumentation, with CCI rates ranging from 6.8 \pm 0.3% to 55.9 \pm 3%. These disparities indicate that the 525 models in their current form are unsuitable for generating accurate predictions of particles based on 526 images obtained using different microscope types or magnifications.

- 527
- 528
- 529

530 531

532 533 534

535

Table 3: Interoperability of the RF models. The interoperability of the Plastic Detection Model (PDM) and the Polymer Identification Model (PIM) was tested by analysing MPs in the ILVO datasets 1 and 2, acquired with a fluorescence stereomicroscope (FSM) in the ILVO laboratory, using models based on the VLIZ datasets 1 and 2, respectively, acquired with a fluorescence microscope (FM) in the VLIZ laboratory, and vice versa.

542

ILVO MODELS BASED ON FSM

VLIZ MODELS BASED ON FM

ANALYSED PARTICLES BASED ON FM (VLIZ)

ANALYSED PARTICLES BASED ON FSM (ILVO)

RANDOM FOREST		DIM		DINA	
CLASSIFIER	PDIVI	FIN	PDIVI	PIIVI	
PE/PP	100 ± 0%	66.2 ± 4.6%	100 ± 0%	100 ± 0%	
PET	100 ± 0%	55.9 ± 3%	100 ± 0%	93.2 ± 0.3%	
PS	100 ± 0%	78.4 ± 27.4%	100 ± 0%	6.8 ± 0.3%	
PVC	100 ± 0%	100 ± 0%	100 ± 0%	18.5 ± 0%	
All plastics	96.9 ± 0.1%	73.3 ± 12.6%	100 ± 0%	35.4 ± 48.8%	
Organic	58.7 ± 0.2%		0.8 ± 0.4%		

543

544 **4. Discussion**

545

546 <u>4.1 Choice of machine learning algorithm</u>

547 In this research, both DT and RF models demonstrated similarly high accuracies in detecting plastics and 548 identifying the polymer composition of pristine materials. When applied to weathered plastic particles, 549 both models maintained high detection accuracies (SFW: 77.2 ± 38% (DT) vs. 78.8 ± 36% (RF); DSW: 86.8 550 \pm 20% (DT) vs. 87.8 \pm 17 (RF)) (Tables 1 and 2). However, for identifying the specific polymers of weathered 551 plastics, only the RF models achieved sufficiently high accuracies (SFW: 63.7 ± 38.4% (DT) vs. 70 ± 39.2% 552 (RF); DSW: 60.8 ± 26.3% (DT) vs. 80.3 ± 14.8 (RF)). Decision tree and RF algorithms represent two 553 prominent and popular ML approaches within MP research (Li et al., 2023; Wu et al., 2023; Yao et al., 554 2023), each offering advantages and challenges. Decision tree models are more prone to overfitting, 555 which can reduce accuracy on new data. A maximum depth as well as a minimum sample size per node 556 should be specified to prevent the tree from growing endlessly, ensuring the model captures only the 557 most general and crucial aspects of the dataset. Despite these challenges, DTs are valued for their 558 transparency and interpretability. The simplicity of these white box models allows for an easy 559 visualisation, which makes them particularly useful when model clarity is essential.

561 Random forest models, classified as black box models, are inherently more complex than DTs, which can 562 pose challenges in understanding their internal structure and interpreting the reasoning behind certain 563 predictions. Unlike DTs, RFs do not provide a clear breakdown of weighted scores, but they are generally 564 more robust, effectively handling the instability and overfitting issues that can affect a single DT (Bienefeld 565 et al., 2022). Random forest models can leverage the power of multiple DTs for decision making by 566 creating random subsets of features to build multiple smaller DTs, which are then combined. In qualitative 567 data classification, such as in this study, the predictions of these DTs are aggregated through a majority 568 voting system to produce a final prediction (Svetnik et al., 2003). Although RFs do not require pruning, 569 their performance is sensitive to the number of predictive variables considered. Increasing these variables 570 enhances model strength but also raises intercorrelation, while decreasing them has the opposite effect. 571 Additionally, while increasing the number of trees generally improves predictive accuracy, a threshold 572 exists beyond which there is no significant performance gain anymore. However, with an increasing 573 number of trees to be tested often comes an increased analysis time.

574

575 Our study underlines the robustness of the RF algorithm in managing alterations in polymers due to 576 weathering in surface and deep-sea waters. The collective strength of RFs was essential in maintaining a 577 sufficient predictive accuracy despite potential variations in fluorescence colouration and intensity caused 578 by MP weathering. Moreover, the ability of the RF models to mitigate overfitting while enhancing 579 interoperability contributed to their superior performance in capturing potential spectral changes or 580 changes in fluorescence intensity across different polymers. Despite their complexity, the increase in 581 computational costs associated with RF models compared to the DT models was negligible. Therefore, for 582 analyses where both MP quantification and identification are required, RF emerges as the superior 583 algorithm. However, if polymer identification is not a research goal, both DT and RF classifiers allow for a 584 reliable analysis of MPs in a cost- and time-effective manner.

585

586 The ability to accurately analyse weathered MPs makes the developed models particularly relevant, as it 587 ensures their applicability to real-world environmental samples. Other studies have also leveraged RF 588 classifiers for MP analysis: Vitali et al., 2024 developed a method for analysing MPs in bottled water using 589 NR staining and an RF-based automated image processing workflow, achieving precise quantification and 590 sizing of MPs down to 10 μm. At the same time, Wang et al., 2024 utilised flow cytometry coupled with 591 ML, including RF algorithms, to effectively differentiate MPs from natural particles in aqueous 592 suspensions. Alternative methods for identifying MP polymer types not based on chemical analysis include confocal fluorescence microscopy combined with fluorescence life-time imaging microscopy (FLIM),
which distinguishes polymers based on emission spectra but has limited validation in real-world samples
(Sancataldo et al., 2020). Another approach, using photoluminescence spectroscopy alongside NR
staining, differentiates plastics based on their Stokes shift, although the impact of weathering on spectral
emission still requires further study (Konde et al., 2020).

598

599 The effective application of the new method requires a solid understanding of machine learning 600 techniques, where DTs offer a straightforward and simple interpretation through their transparent inner 601 structure, while the more complex RFs require a specialised knowledge, potentially requiring additional 602 training. Next to this, while the multi-filter approach reduces the number of false positives compared to 603 single-filter methods (Meyers et al., 2022) and enables polymer identification, its increased complexity 604 requires a thorough understanding of the NR staining process, familiarity with the expected fluorescent 605 colouration of reference plastics, and the ability to effectively operate fluorescence microscopes with 606 multiple filters.

607

608 <u>4.2 Environmental weathering of microplastics and its consequences for microplastic analysis</u>

609 The developed models presented in this work leveraged the solvatochromic properties of NR to predict 610 particle identity. The emission spectrum of NR shifts based on the polarity of its environment, which 611 allows for distinguishing MPs into "polar" and "hydrophobic" categories according to their polymer 612 characteristics (Maes et al., 2017). As polymer polarity increases, the maximum emission wavelengths 613 shift towards longer wavelengths, facilitating further classification into specific polymer types through 614 quantification of their fluorescent colouration (Meyers et al., 2022). Notably, the fluorescent colouration 615 of SFW PET particles following NR staining substantially differed from that of pristine PET particles, a 616 change not observed in DSW PET particles (Fig. 4). This suggests susceptibility of this polymer to 617 weathering processes that are more dominant in sea-surface waters, such as UV radiation and 618 microorganism settlement. To improve the accuracy of predictions for polymers like PET, it is 619 recommended to include RGB data from weathered particles in the training datasets. This can be achieved 620 through artificial weathering processes or semi-controlled environmental weathering, as implemented in 621 this study. Incorporating such data would likely enhance the overall predictive accuracy of the models. 622 Alternatively, RGB datasets could be constructed using naturally weathered MPs, although this approach 623 may be more labour- and time-intensive.

625 Plastic waste typically decomposes slowly but weathers and breaks down into MPs when exposed to UV 626 radiation, mechanical abrasion, temperature changes, and biodegradation. These weathering processes 627 rapidly alter the physical and chemical properties of MPs, affecting their environmental behaviour, 628 including increased leaching of additives, changes in molecular weight and surface roughness, and 629 enhanced pollutant absorption due to biofilm formation (Liu et al., 2019; Duan et al., 2021). The absence 630 of a pretreatment step, which generally applies for water samples low in organic content (Gago et al., 631 2019), may have allowed biofilm residues to remain on the MPs, potentially affecting NR staining and 632 fluorescence analysis. A recent validation study from our laboratory that employed digestion methods 633 (Meyers et al., 2024) demonstrated higher accuracy in identifying weathered MPs, with 98.25 ± 3.04% of 634 DSW MPs and 100% of SFW MPs correctly identified by the PDM. The slightly lower accuracy observed in 635 the current study (78.8 \pm 36% for SFW MPs and 87.8 \pm 17% for DSW MPs) may be attributed to the 636 presence and costaining of biofilms on the untreated MPs (Macedo et al., 2005). The polymer 637 identification accuracy for weathered plastics was similar in both studies.

638

639 Environmental ageing processes pose challenges for MP analysis methods, which are often designed and 640 tested using pristine MPs. As became apparent from the obtained results, weathering-induced changes in 641 chemical and physical properties can affect the fluorescent colouration of NR-stained MPs, hindering 642 accurate model classification. Standard Raman and IR spectra of MPs are also affected by aging, leading 643 to shifts that complicate matching with commercial libraries of pristine materials (Fig. S8-S12) (Dong et 644 al., 2020). Developing reference libraries based on naturally weathered MPs could address this issue, but 645 it may increase total analysis time and labour costs. However, adding weathered MP spectra to 646 commercial libraries of pristine MPs has also been shown to improve spectral matching accuracy for 647 environmental samples (De Frond et al., 2021). The ATR spectra generated during this study have been 648 made publicly available (Meyers et al., 2024c). Weathering-induced polymer alterations and potential 649 changes in associated additives may also complicate identification and quantification based on mass 650 spectrometry, another frequently employed analysis (Primpke et al., 2020a). In addition to this, 651 weathering processes may interfere with the step preceding the analysis, where samples are extracted 652 from sample matrices. Environmental degradation has been shown to alter the densities of certain 653 polymers (Kowalski et al., 2016), potentially making it more difficult to efficiently isolate all MPs present 654 in sediment samples.

- 655
- 656

657 <u>4.3 Weathered reference plastics</u>

658 Researchers are increasingly recognising the importance of using reference materials that mimic the 659 properties of weathered MPs when developing and testing new methods, next to the incorporation of a 660 variety of MP polymers, sizes and shapes. This study employed a unique approach by weathering a mix of 661 MPs at sea under semi-controlled surface water and deep-sea water conditions for 12 months, offering a 662 compromise between controlled laboratory simulations and the weathering of MPs under untraceable 663 natural conditions. By adopting this approach, we address inherent challenges associated with both 664 methodologies: unlike conventional laboratory experiments, the weathering process took place under 665 authentic natural conditions. Consequently, the MPs within the containers were exposed to the combined 666 effects of diverse degradation processes and various influencing factors. Periodic cleaning of the surface 667 water containers ensured unobstructed water flow, facilitating natural biodegradation processes. 668 However, the degree of UV weathering in the surface weathering experiment might have been influenced 669 by the experimental setup, as sunlight only partially passed through the container pores. Similarly, the 670 impact of sea wave force may have been partially mitigated by the container. Nevertheless, this setup was 671 imperative for the controlled natural degradation of MPs within the confines of the container. Despite 672 being potentially less labour- and time-intensive compared to the collection of weathered MPs/the 673 collection and cryomilling of weathered macroplastics, the duration of degradation was set to a period of 674 one year in order to be meaningful. As a last point, despite introducing heterogeneity in shape, the process 675 of cryomilling to produce the used MP fragments did not mimic mechanical weathering to the full extent, 676 as various processes such as stretching, tearing, and crushing also contribute to plastic fragmentation in 677 a marine context. Consequently, crucial MP characteristics for risk assessments, such as particle size, do 678 not fully align with the naturally occurring composition. However, in contrast to environmentally sourced 679 MPs with unknown origins and history, the precise deployment location and bathymetric conditions were 680 known and controlled for, along with the duration of weathering. Meteorological conditions, currents, 681 weather patterns, and UV radiation can be retrospectively traced, and can provide a comprehensive 682 understanding if required. Additionally, the study employed a MP composition of the most prevalently 683 produced and encountered plastics, which enhances the global applicability of the results. Lastly, the MPs 684 in this study underwent a weathering duration of 12 months, surpassing the temporal scope of many prior 685 studies (e.g. Naik et al., 2020). Nonetheless, it is acknowledged that certain MPs require multiple years to 686 exhibit substantial weathering effects (Chamas et al., 2020). Alternatively, environmentally relevant 687 weathered MP samples can be produced by cryomilling plastic macrolitter collected from beaches (Kühn 688 et al., 2018).

Future prospects of the approach applied in this work include deployments of weathering containers over prolonged durations to accurately reflect long-term degradation processes, while the generation of MPs through mechanical abrasion processes representative of the marine environment should be pursued. Next to this, refinements in experimental setup are required to ensure unhindered exposure of MPs to sunlight and mechanical forces.

695

696 <u>4.3 Lower size limit</u>

697 The RF models in this study showed an accuracy > 90% for particles with Feret diameters between 12 and 698 30 pixels, corresponding to MPs < 10 μ m at the microscope magnification used. The ability to identify MPs 699 of ecotoxicologically relevant sizes (Beiras et al., 2020) in a cost- and time-effective way enhances the 700 ecological relevance of the method and fills a critical gap. Other frequently used MP analysis methods 701 show similar lower size limits, e.g. 1 μ m for μ -Raman and 10 - 20 μ m for μ -FTIR (Cabernard et al., 2018; 702 Mintenig et al., 2019; Primpke et al., 2020a). Although no MP size limitation exists for GC-MS-based 703 techniques, a significant drawback is their inability to quantify or characterise MPs physically, information 704 that is essential for risk evaluations (Schwarzer et al., 2022, Qiao et al., 2019). In prior research, NR co-705 staining of organic material like residual fat posed challenges for NR-based MP analysis (Prata et al., 2021). 706 Efficient matrix removal is crucial but often not feasible, risking inaccuracies in MP 707 detection/quantification (Shruti et al., 2022). In this study, however, the use of multiple filters (UV, blue, 708 green) and the inclusion of fluorescence data unique to organic materials and distinct from plastic into 709 the RF models helped to differentiate between these materials. Additionally, the risk of persistent false 710 positives due to similar fluorescence could be reduced using customised RGB datasets (cfr. '4.4 711 Interoperability').

712

Other methods have also succeeded in detecting smaller MPs using a NR-based approach. For example, Ko et al., 2024 introduced a system combining fluorescence labelling with a microfluidic device and particle tracking software, enabling automated size measurement and real-time discrimination of MPs sized 100–1000 nm, such as PS and PVC, in small water samples. Similarly, Bianco et al., 2022 developed a method combining NR staining and flow cytometry and was able to quantify plastic particles in the 0.6– 15 µm size range.

- 719
- 720

722 <u>4.4 Interoperability</u>

723 Ensuring consistent method performance across different laboratories is crucial in scientific research and 724 environmental monitoring. When comparing the performance of models built and tested with distinct 725 microscopes, both demonstrated high accuracies for pristine materials. However, during the 726 interoperability assessment, the accuracy of the RF algorithm decreased when models were built and 727 tested using images acquired with different microscope types. Implementing a preprocessing pipeline as 728 a normalisation step to address differences in image acquisition parameters, or leveraging transfer 729 learning, could help tackle this issue. Another option is the development of new datasets. While this 730 outcome presents a challenge, it is important to note that the foundation of the method lies in training 731 datasets which can be easily and rapidly constructed for a specific laboratory (Meyers et al., 2022). For 732 instance, using a fluorescence stereomicroscope (FSM), over 100 reference particles sized < 500 μm can 733 be filtered onto a single PTFE filter and captured in a single image series (blue, green, and UV filters), 734 enabling the construction of an RGB dataset within a day following staining. To enhance predictive 735 robustness, it is however recommended to include RGB data from MPs stained with NR at different points 736 in time. Building a training dataset, constructing a model, and testing its accuracy can be completed in 737 two days. This inherent flexibility of the method allows for its swift adaptation and optimisation, in this 738 way ensuring the possibility to fine-tune the method to specific laboratory conditions and set 739 requirements. In this way, researchers can account for unique variables in their own research that could 740 potentially affect model performance, such as 1) the specific MP polymer composition being targeted, 2) 741 the matrix type from which these MPs are extracted and which may interact with the MPs' NR 742 fluorescence, 3) the specific types of organic material present in that matrix which could interfere with 743 MP detections (e.g. chitin in seawater samples or lipid residues in gastrointestinal tracts (GITs) of fish), 4) 744 the microscope model and magnification used to perform the analyses, and so on. Consequently, the 745 lower accuracy obtained when testing the interoperability of the models does not impede their 746 implementation elsewhere. Instead, it emphasises the adaptability of the developed method. Giving 747 laboratories the ability to efficiently create customised datasets allows them to address specific 748 challenges encountered, thereby strengthening the robustness of the method and its successful 749 deployment in diverse environments.

750

Additionally, this cost-effective approach is particularly beneficial for laboratories that lack the expensive
equipment often associated with MP analysis (Primpke et al., 2020a). In this way, the approach enables a

broader range of laboratories to engage in MPs research, facilitating the advancement of knowledge on
plastic pollution across diverse marine environments worldwide.

755

5. Conclusion

757

756

758 Both DT and RF models demonstrated high accuracy in detecting pristine and weathered MPs. Despite 759 their complexity, RF models are preferred for polymer identification due to their superior performance 760 and minimal increase in computational time. Although the models generally had a high predictive 761 reliability, incorporating RGB data from weathered particles could further enhance accuracy for specific 762 polymers like PET. The models also proved effective in detecting and identifying MPs smaller than 10 μm, 763 underlining their potential in analysing ecotoxicologically relevant MPs in marine environments. Although 764 interlaboratory assessments revealed challenges related to microscope type variations, the adaptability 765 of the RF models allows customisation to specific conditions, ensuring robustness and successful 766 application in diverse settings.

767

The relevance and novelty of the method are underscored by its ability to accurately analyse MPs weathered under various marine conditions, making it highly applicable to real-world environmental samples. Additionally, its capability to detect ecotoxicologically relevant MP sizes in a cost- and timeeffective manner addresses a critical gap in MP research. Moreover, by providing a cost- and timeeffective alternative to traditional methods, the ML-based method enables a wide range of laboratories to engage in MP research.

774

775 Acknowledgments

776

The authors thank the captain and crew of the R/V Antedon II (Flotte Océanographique Française), as well as Deny Malengros, Laure Papillon and Mélanie Ourgaud for their assistance in the deployment and retrieval of the deep-sea samples. We also thank Mattias Bossaer from VLIZ for assisting in the acquisition of ATR spectra of both pristine and weathered microplastics. Lastly, we thank the partners of the JPI Oceans Andromeda project who supported this work, as well as BELSPO, RBINS OD Nature, the Pluxin project, the French Agence Nationale de Recherche (ANR-19-383 JOCE-0002-01) and the Norwegian Research Council (311313/E40) for funding or for financing ship time.

784	
785	Appendix A. Supplementary data
786	
787	Supplementary data to this article can be found online at:
788	
789	Ethical Approval
790	
791	Not applicable
792	
793	Consent to Participate
794	
795	Not applicable
796	
797	Consent to Publish
798	
799	Not applicable
800	
801	Data availability
802	
803	The RGB colour datasets of all models are freely available: https://doi.org/10.14284/665. Other data will
804	be made available on reasonable request.
805	
806	Authors Contributions
807	
808	Nelle Meyers: Conceptualisation, Data curation, Formal analysis, Investigation, Methodology, Project
809	administration, Software, Validation, Visualisation, Writing – original draft, Writing – review & editing.
810	Bavo De Witte: Conceptualisation, Methodology, Project administration, Supervision, Funding
811	acquisition, Writing – review & editing. Natascha Schmidt: Conceiving, designing, and performing of
812	weathering experiments, Writing – review & editing. Dorte Herzke: Conceiving, designing, and performing
813	of weathering experiments, Funding acquisition, Writing – review & editing. Jean-Luc Fuda: Conceiving,

814	designing, and performing of weathering experiments, Writing – review & editing. David Vanavermaete:
815	Methodology, Software, Writing - review & editing. Colin Janssen: Supervision, Funding acquisition,
816	Writing - review & editing. Gert Everaert: Conceptualisation, Methodology, Project administration,
817	Supervision, Funding acquisition, Writing – review & editing.
818	
819	Funding
820	
821	This work was supported by the Andromeda project (JPI Oceans). We thank BELSPO [contract no
822	B2/20E/P1/Andromeda], the Pluxin project, the French Agence Nationale de Recherche (ANR-19-383
823	JOCE-0002-01) and the Norwegian Research Council (311313/E40) for funding.
824	
825	Competing interests
826	
827	The authors declare that they have no known competing financial interests or personal relationships that
828	could have appeared to influence the work reported in this paper.
829	
830	References
831	
832 833	Abràmoff, M.D., Magalhães, P.J. and Ram, S.J., 2004. Image processing with ImageJ. Biophotonics international, 11(7), pp.36-42.
834	Alimi, O.S., Claveau-Mallet, D., Kurusu, R.S., Lapointe, M., Bayen, S. and Tufenkji, N., 2022. Weathering pathways and protocols
835	for environmentally relevant microplastics and nanoplastics: What are we missing?. Journal of Hazardous Materials, 423,
836	p.126955.
838	Arp, H.P.H., Kühnel, D., Rummel, C., MacLeod, M., Potthoff, A., Reichelt, S., Roio-Nieto, F., Schmitt-Jansen, M., Sonnenberg, J.,
839	Toorman, E. and Jahnke, A., 2021. Weathering plastics as a planetary boundary threat: exposure, fate, and hazards.
840	Environmental science & technology, 55(11), pp.7246-7255.
841	
842	Arthur, C., Baker, J.E. and Bamford, H.A., 2009. Proceedings of the International Research Workshop on the Occurrence, Effects,
043 844	and Fate of Microplastic Marine Debris, September 9-11, 2008, University of Washington Tacoma, Tacoma, WA, USA.
845	Beiras, R. and Schönemann, A.M., 2020. Currently monitored microplastics pose negligible ecological risk to the global ocean.
846	Scientific reports, 10(1), p.22281.
847	

- Bergmann, M., Mützel, S., Primpke, S., Tekman, M.B., Trachsel, J. and Gerdts, G., 2019. White and wonderful? Microplastics
 prevail in snow from the Alps to the Arctic. Science advances, 5(8), p.eaax1157.
- 850
- Bhagat, K., Barrios, A.C., Rajwade, K., Kumar, A., Oswald, J., Apul, O. and Perreault, F., 2022. Aging of microplastics increases their
 adsorption affinity towards organic contaminants. Chemosphere, 298, p.134238.
- 853
- Bianco, A., Carena, L., Peitsaro, N., Sordello, F., Vione, D. and Passananti, M., 2023. Rapid detection of nanoplastics and small
 microplastics by Nile-Red staining and flow cytometry. Environmental Chemistry Letters, 21(2), pp.647-653.
- 856
- Bienefeld, C., Kirchner, E., Vogt, A. and Kacmar, M., 2022. On the importance of temporal information for remaining useful life
 prediction of rolling bearings using a random forest regressor. Lubricants, 10(4), p.67.
- 860 Breiman, L., Friedman, J.H., Olshen, R.A., Stone, C.J. Classification and Regression Trees, Chapman & Hall, 1993.
- 861

- 862 Breiman, L., 1996. Bagging predictors. Machine learning, 24, pp.123-140.
- 863

865

- 864 Breiman, L., 2001. Random forests. Machine learning, 45, pp.5-32.
- Cabernard, L., Roscher, L., Lorenz, C., Gerdts, G. and Primpke, S., 2018. Comparison of Raman and Fourier transform infrared
 spectroscopy for the quantification of microplastics in the aquatic environment. Environmental science & technology, 52(22),
 pp.13279-13288.
- 869

872

Catarino, A.I., Kramm, J., Voelker, C., Henry, T.B. and Everaert, G., 2021. Risk posed by microplastics: Scientific evidence and public
 perception. Current Opinion in Green and Sustainable Chemistry, 29, p.100467.

- 873 Chamas, A., Moon, H., Zheng, J., Qiu, Y., Tabassum, T., Jang, J.H., Abu-Omar, M., Scott, S.L. and Suh, S., 2020. Degradation rates
 874 of plastics in the environment. ACS Sustainable Chemistry & Engineering, 8(9), pp.3494-3511.
- 875
 876 Cowger, W., Gray, A., Christiansen, S.H., DeFrond, H., Deshpande, A.D., Hemabessiere, L., Lee, E., Mill, L., Munno, K., Ossmann,
- B.E. and Pittroff, M., 2020. Critical review of processing and classification techniques for images and spectra in microplastic
 research. Applied Spectroscopy, 74(9), pp.989-1010.
- 879
 880 Daniya, T., Geetha, M. and Kumar, K.S., 2020. Classification and regression trees with gini index. Advances in Mathematics:
 881 Scientific Journal, 9(10), pp.8237-8247.
 - 882
 - Bong, M., Zhang, Q., Xing, X., Chen, W., She, Z. and Luo, Z., 2020. Raman spectra and surface changes of microplastics weathered
 under natural environments. Science of The Total Environment, 739, p.139990.
 - 885

886 Duan, J., Bolan, N., Li, Y., Ding, S., Atugoda, T., Vithanage, M., Sarkar, B., Tsang, D.C. and Kirkham, M.B., 2021. Weathering of 887 microplastics and interaction with other coexisting constituents in terrestrial and aquatic environments. Water Research, 196, 888 p.117011. 889 890 Dunford, R., Su, Q. and Tamang, E., 2014. The pareto principle. 891 892 Fauvelle, V., Garel, M., Tamburini, C., Nerini, D., Castro-Jiménez, J., Schmidt, N., Paluselli, A., Fahs, A., Papillon, L., Booth, A.M. 893 and Sempéré, R., 2021. Organic additive release from plastic to seawater is lower under deep-sea conditions. Nature 894 Communications, 12(1), p.4426. 895 896 Fotopoulou, K.N. and Karapanagioti, H.K., 2019. Degradation of various plastics in the environment. Hazardous chemicals 897 associated with plastics in the marine environment, pp.71-92. 898 899 Gago, J., Filgueiras, A., Pedrotti, M.L., Caetano, M. and Frias, J., 2019. Standardised protocol for monitoring microplastics in 900 seawater. Deliverable 4.1. 901 902 903 Guo, P., Wang, Y., Moghaddamfard, P., Meng, W., Wu, S. and Bao, Y., 2024. Artificial intelligence-empowered collection and 904 characterization of microplastics: A review. Journal of Hazardous Materials, p.134405. 905 906 Harris-Birtill, D. and Harris-Birtill, R., 2021. Understanding computation time: a critical discussion of time as a computational 907 performance metric. In Time in Variance (pp. 220-248). Brill. 908 909 Hartmann, N.B., Huffer, T., Thompson, R.C., Hassellov, M., Verschoor, A., Daugaard, A.E., Rist, S., Karlsson, T., Brennholt, N., Cole, 910 M. and Herrling, M.P., 2019. Are we speaking the same language? Recommendations for a definition and categorization 911 framework for plastic debris. 912 913 Kiki, C., Qiu, Y., Wang, Q., Ifon, B.E., Qin, D., Chabi, K., Yu, C.P., Zhu, Y.G. and Sun, Q., 2022. Induced aging, structural change, and 914 adsorption behavior modifications of microplastics by microalgae. Environment International, 166, p.107382. 915 916 Kinigopoulou, V., Pashalidis, I., Kalderis, D. and Anastopoulos, I., 2022. Microplastics as carriers of inorganic and organic 917 contaminants in the environment: A review of recent progress. Journal of Molecular Liquids, 350, p.118580. 918 919 Ko, K. and Chung, H., 2024. Fluorescence microfluidic system for real-time monitoring of PS and PVC sub-micron microplastics 920 under flowing conditions. Science of The Total Environment, 950, p.175016. 921 Kolandhasamy, P., Su, L., Li, J., Qu, X., Jabeen, K. and Shi, H., 2018. Adherence of microplastics to soft tissue of mussels: a novel 922 way to uptake microplastics beyond ingestion. Science of the total environment, 610, pp.635-640. 923

925 combination with Nile Red staining for microplastic detection. Marine Pollution Bulletin, 159, p.111475. 926 927 Kowalski, N., Reichardt, A.M. and Waniek, J.J., 2016. Sinking rates of microplastics and potential implications of their alteration 928 by physical, biological, and chemical factors. Marine pollution bulletin, 109(1), pp.310-319. 929 930 Kühn, S., Van Oyen, A., Booth, A.M., Meijboom, A. and Van Franeker, J.A., 2018. Marine microplastic: Preparation of relevant 931 test materials for laboratory assessment of ecosystem impacts. Chemosphere, 213, pp.103-113. 932 933 Li, W., Li, X., Tong, J., Xiong, W., Zhu, Z., Gao, X., Li, S., Jia, M., Yang, Z. and Liang, J., 2023. Effects of environmental and 934 anthropogenic factors on the distribution and abundance of microplastics in freshwater ecosystems. Science of The Total 935 Environment, 856, p.159030. 936 937 Liaw, A. and Wiener, M., 2002. Classification and regression by randomForest. R news, 2(3), pp.18-22. 938 939 Lin, J.Y., Liu, H.T. and Zhang, J., 2022. Recent advances in the application of machine learning methods to improve identification 940 of the microplastics in environment. Chemosphere, p.136092. 941 942 Liu, P., Qian, L., Wang, H., Zhan, X., Lu, K., Gu, C. and Gao, S., 2019. New insights into the aging behavior of microplastics 943 accelerated by advanced oxidation processes. Environmental science & technology, 53(7), pp.3579-3588. 944 945 Liu, P., Zhan, X., Wu, X., Li, J., Wang, H. and Gao, S., 2020. Effect of weathering on environmental behavior of microplastics: 946 Properties, sorption and potential risks. Chemosphere, 242, p.125193. 947 948 Liu, R., Wang, Y., Yang, Y., Shen, L., Zhang, B., Dong, Z., Gao, C. and Xing, B., 2023. New insights into adsorption mechanism of 949 pristine and weathered polyamide microplastics towards hydrophilic organic compounds. Environmental Pollution, 317, 950 p.120818. 951 952 Maes, T., Jessop, R., Wellner, N., Haupt, K. and Mayes, A.G., 2017. A rapid-screening approach to detect and quantify microplastics 953 based on fluorescent tagging with Nile Red. Scientific reports, 7(1), p.44501. 954 955 Mattsson, K., Jocic, S., Doverbratt, I. and Hansson, L.A., 2018. Nanoplastics in the aquatic environment. Microplastic 956 contamination in aquatic environments, pp.379-399. 957 958 Maxwell, A.E., Warner, T.A. and Fang, F., 2018. Implementation of machine-learning classification in remote sensing: An applied 959 review. International journal of remote sensing, 39(9), pp.2784-2817. 960 961 McHugh, M.L., 2012. Interrater reliability: the kappa statistic. Biochemia medica, 22(3), pp.276-282. 962

Konde, S., Ornik, J., Prume, J.A., Taiber, J. and Koch, M., 2020. Exploring the potential of photoluminescence spectroscopy in

- 963 Meyers, N., Catarino, A.I., Declercq, A.M., Brenan, A., Devriese, L., Vandegehuchte, M., De Witte, B., Janssen, C. and Everaert, G.,
 964 2022. Microplastic detection and identification by Nile red staining: Towards a semi-automated, cost-and time-effective
 965 technique. Science of the Total Environment, 823, p.153441.
- 967 Meyers, N.; De Witte, B.; Catarino, A. I.; & Everaert, G. 2024a. Standardised operating protocol for automated microplastic
 968 analysis using machine learning models. JPI Oceans Andromeda Project.
- 969

- 970 Meyers, N.; De Witte, B.; Janssen, C.; Everaert, G.; Flanders Marine Institute (VLIZ); Flanders Research Institute for Agriculture,
- 971 Fisheries and Food (ILVO); Ghent University Laboratory for Environmental Toxicology (GhEnToxLab): Belgium; 2024b: RGB
 972 datasets for machine learning-based microplastic analysis update. Marine Data Archive. https://doi.org/10.14284/665
- 973
- 974 Meyers, N.; De Witte, B.; Schmidt, N.; Herzke, D.; Fuda, J.; Vanavermaete, D; Bossaer, M.; Janssen, C.; Everaert, G. 2024c. Infrared
 975 spectra of plastic polymers weathered in the marine environment under semi-controlled conditions.
 976 https://doi.org/10.14284/664
- 977
- Meyers, N., Everaert, G., Hostens, K., Schmidt, N., Herzke, D., Fuda, J.L., Janssen, C.R. and De Witte, B., 2024. Towards reliable
 data: Validation of a machine learning-based approach for microplastics analysis in marine organisms using Nile red
 staining. *Marine Pollution Bulletin*, 207, p.116804.
- 981

- 982 Mintenig, S.M., Löder, M.G., Primpke, S. and Gerdts, G., 2019. Low numbers of microplastics detected in drinking water from
 983 ground water sources. Science of the total environment, 648, pp.631-635.
- 985 Naik, R.A., Rowles III, L.S., Hossain, A.I., Yen, M., Aldossary, R.M., Apul, O.G., Conkle, J. and Saleh, N.B., 2020. Microplastic particle
 986 versus fiber generation during photo-transformation in simulated seawater. Science of The Total Environment, 736, p.139690.
- 987
 988 Paradinas, L.M., James, N.A., Quinn, B., Dale, A. and Narayanaswamy, B.E., 2021. A new collection tool-kit to sample microplastics
- 989 from the marine environment (sediment, seawater, and biota) using citizen science. Frontiers in Marine Science, 8, p.657709.990
- 991 Peeken, I., Primpke, S., Beyer, B., Gütermann, J., Katlein, C., Krumpen, T., Bergmann, M., Hehemann, L. and Gerdts, G., 2018.
- Arctic sea ice is an important temporal sink and means of transport for microplastic. Nature communications, 9(1), p.1505.993
- Peng, X., Chen, M., Chen, S., Dasgupta, S., Xu, H., Ta, K., Du, M., Li, J., Guo, Z. and Bai, S., 2018. Microplastics contaminate the
 deepest part of the world's ocean. Geochemical Perspectives Letters, 9(1), pp.1-5.
- 996
- 997 Prata, J.C., Sequeira, I.F., Monteiro, S.S., Silva, A.L.P., da Costa, J.P., Dias-Pereira, P., Fernandes, A.J.S., da Costa, F.M., Duarte,
- 998 A.C., Rocha-Santos, T., 2021. Preparation of biological samples for microplastic identification by Nile Red. Sci. Total Environ.
- **999** 783, 147065.
- 1000

1001 Primpke, S., Christiansen, S.H., Cowger, W., De Frond, H., Deshpande, A., Fischer, M., Holland, E.B., Meyns, M., O'Donnell, B.A., 1002 Ossmann, B.E. and Pittroff, M., 2020a. Critical assessment of analytical methods for the harmonized and cost-efficient analysis 1003 of microplastics. Applied Spectroscopy, 74(9), pp.1012-1047. 1004 1005 Primpke, S., Cross, R.K., Mintenig, S.M., Simon, M., Vianello, A., Gerdts, G. and Vollertsen, J., 2020b. Toward the systematic 1006 identification of microplastics in the environment: evaluation of a new independent software tool (siMPle) for spectroscopic 1007 analysis. 2020b. Applied Spectroscopy, 74(9), pp.1127-1138. 1008 1009 Qiao, R., Deng, Y., Zhang, S., Wolosker, M. B., Zhu, Q., Ren, H., & Zhang, Y. (2019). Accumulation of different shapes of 1010 microplastics initiates intestinal injury and gut microbiota dysbiosis in the gut of zebrafish. Chemosphere, 236, 124334. 1011 1012 Ross, P.S., Chastain, S., Vassilenko, E., Etemadifar, A., Zimmermann, S., Quesnel, S.A., Eert, J., Solomon, E., Patankar, S., Posacka, 1013 A.M. and Williams, B., 2021. Pervasive distribution of polyester fibres in the Arctic Ocean is driven by Atlantic inputs. Nature 1014 communications, 12(1), p.106. 1015 1016 Sancataldo, G., Avellone, G. and Vetri, V., 2020. Nile Red lifetime reveals microplastic identity. Environmental Science: 1017 Processes & Impacts, 22(11), pp.2266-2275. 1018 1019 Serdar, C.C., Cihan, M., Yücel, D. and Serdar, M.A., 2021. Sample size, power and effect size revisited: simplified and practical 1020 approaches in pre-clinical, clinical and laboratory studies. Biochemia medica, 31(1), pp.27-53. 1021 1022 Shah, A.A., Hasan, F., Hameed, A. and Ahmed, S., 2008. Biological degradation of plastics: a comprehensive review. Biotechnology 1023 advances, 26(3), pp.246-265. 1024 1025 Shruti, V.C., Pérez-Guevara, F., Roy, P.D., Kutralam-Muniasamy, G., 2022. Analyzing microplastics with Nile red: emerging trends, 1026 challenges, and prospects. J. Hazard. Mater. 423, 127171. 1027 1028 Svetnik, V., Liaw, A., Tong, C., Culberson, J.C., Sheridan, R.P. and Feuston, B.P., 2003. Random forest: a classification and regression 1029 tool for compound classification and QSAR modeling. Journal of chemical information and computer sciences, 43(6), pp.1947-1030 1958. 1031 1032 Schwarzer, M., Brehm, J., Vollmer, M., Jasinski, J., Xu, C., Zainuddin, S., Fröhlich, T., Schott, M., Greiner, A., Scheibel, T. & Laforsch, 1033 C. (2022). Shape, size, and polymer dependent effects of microplastics on Daphnia magna. Journal of Hazardous Materials, 426, 1034 128136. 1035 1036 Therneau, T., Atkinson, B., Ripley, B. and Ripley, M.B., 2015. Package 'rpart'. Available online: cran. ma. ic. ac. 1037 uk/web/packages/rpart/rpart. pdf (accessed on 20 April 2016). 1038

1039	Van Cauwenberghe, L., Vanreusel, A., Mees, J. and Janssen, C.R., 2013. Microplastic pollution in deep-sea sediments.
1040	Environmental pollution, 182, pp.495-499.
1041	
1042	Vitali, C., Peters, R.J., Janssen, H.G., Undas, A.K., Munniks, S., Ruggeri, F.S. and Nielen, M.W., 2024. Quantitative image analysis
1043	of microplastics in bottled water using artificial intelligence. Talanta, 266, p.124965.
1044	
1045	Yan, X., Cao, Z., Murphy, A. and Qiao, Y., 2022. An ensemble machine learning method for microplastics identification with FTIR
1046	spectrum. Journal of Environmental Chemical Engineering, 10(4), p.108130.
1047	
1048	Wagner, S. and Reemtsma, T., 2019. Things we know and don't know about nanoplastic in the environment. Nature
1049	nanotechnology, 14(4), pp.300-301.
1050	
1051	Waldman, W.R. and Rillig, M.C., 2020. Microplastic research should embrace the complexity of secondary particles.
1052	
1053	Wang, L., Zhang, J., Huang, W. and He, Y., 2023. Laboratory simulated aging methods, mechanisms and characteristic changes of
1054	microplastics: A review. Chemosphere, p.137744.
1055	
1056	Wang, X., Li, Y., Kroll, A. and Mitrano, D.M., 2024. Differentiating Microplastics from Natural Particles in Aqueous Suspensions
1057	Using Flow Cytometry with Machine Learning. Environmental Science & Technology.
1058	
1059	Wayman, C. and Niemann, H., 2021. The fate of plastic in the ocean environment–a minireview. Environmental Science: Processes
1060	& Impacts, 23(2), pp.198-212.
1061	
1062	Weis, J.S. and Palmquist, K.H., 2021. Reality check: experimental studies on microplastics lack realism. Applied Sciences, 11(18),
1063	p.8529.
1064	
1065	Witten, I.H. and Frank, E., 2002. Data mining: practical machine learning tools and techniques with Java implementations. Acm
1066	Sigmod Record, 31(1), pp.76-77.
1067	
1068	World Health Organization, 2022. Dietary and inhalation exposure to nano-and microplastic particles and potential implications
1069	for human health.
1070	
1071	Wu, P., Wang, B., Lu, Y., Cao, G., Xie, P., Wang, W., Chen, D., Huang, G., Jin, H., Yang, Z. and Cai, Z., 2023. Machine Learning-
1072	Assisted Insights into Sources and Fate of Microplastics in Wastewater Treatment Plants. ACS ES&T Water.
1073	
1074	Yao, J., Li, H. and Yang, H.Y., 2023. Predicting adsorption capacity of pharmaceuticals and personal care products on long-term
1075	aged microplastics using machine learning. Journal of Hazardous Materials, 458, p.131963.