



The importance of fine-scale, vertical profiles in characterising nematode community structure

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Abstract

The spatial heterogeneity of the nematode community on an intertidal flat (the Molenplaat) in the Westerschelde estuary (SW Netherlands) has been investigated. The extent to which macroscale (km) variability was more important than microscale (m) variability was tested. In addition, the importance of vertical distribution profile in the sediment in explaining the horizontal macroscale variability was evaluated. Differences in the structure of the community were analysed at a kilometre scale at three sites that differed in chemico-physical features. The differences in geochemical and physical conditions on a horizontal scale were reflected in species composition and trophic structure of the nematode communities, and to a much lesser extent in their total abundance and species diversity.

Detailed investigation of vertical depth profiles showed more pronounced differences between environmentally divergent sites. Sediment granulometry appears to be important in controlling the fauna in the upper sediment layers. At depth, similar faunal assemblages were found irrespective of sediment granulometry, suggesting that other environmental features are more dominant.

Vertically, nematode species showed depth distributions that were indicative of sediment characteristics related to the site-specific hydrodynamic regime. Pronounced vertical segregation of nematode species was observed within sandy sediment under strong hydrodynamic and food-stressed conditions. A surface-dwelling nematode community of large predatory enoplids was separated from a deposit feeding xyalid-microlaimid community in deeper sediment layers (beneath 2 cm). Causal factors for this segregation are thought to be species interactions, feeding strategies and/or physical disturbance. In the finest sediments, with high silt content, almost all nematode species were confined to the upper sediment layers (1.5 cm). A sharp decline in density and diversity with depth was observed. Key factors for this distribution pattern are possibly related to the limited oxygen penetration in surface layers and the occurrence of sulphide in deeper sediment layers. At intermediate hydrodynamic and granulometric conditions, a gradual shifting of nematode community was observed with depth, with dominant nematode species maxima present at specific depth layers. © 2003 Elsevier Ltd. All rights reserved.

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1. Introduction

Knowledge of spatial patterns of benthic organisms and the scale of these patterns contribute to a better understanding of benthic community structure and functioning. Such information is often the best, if not

the only way for assessing interspecific interactions, which to a considerable extent determine community structure. Comparisons between the spatial patterns of consumers and resources provide information on trophic interactions and the spatial scales at which these interactions occur (Pickney & Sandulli, 1990; Sandulli & Pickney, 1999).

On a vertical scale of centimetres, the effect of abiotic characteristics of sediments (e.g. oxygen, water content,

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proximity to surface) on community structure is as important as the other abiotic variables (such as salinity, sedimentological and geomorphological variables), which act on a horizontal scale of hundreds of metres. Moreover, ecological understanding of the functioning of meiobenthic communities is enhanced by knowledge of animal vertical distribution (Soetaert, Vincx, Wittoeck, Tulkens, & Van Gansbeke, 1994). It has been shown that many nematode species exhibit a typical vertical distribution which often relates to a variety of biological, physical and chemical variables (Giere, 1993; Hendelberg & Jensen, 1993; Soetaert et al., 1994; Steyaert, Garner, Van Gansbeke, & Vincx, 1999; Warwick & Gee, 1984; Wetzel, Jensen, & Giere, 1995). It has been argued that the vertical segregation of species will reduce the number of (competitive or predatory) interactions, and this could explain the very high number of species that co-exist in a certain small patch (Joint, Gee, & Warwick, 1982).

The spatial patterns of temperate nematode communities on different horizontal scales have already been investigated extensively in different estuaries. Most of these studies related structural patterns of the nematode assemblages to environmental variables as sedimentary and latitudinal gradients, food resources, salinity, disturbances of different nature (e.g. Guo, Somerfield, Warwick, & Zhang, 2001; Li, Vincx, Herman, & Heip, 1997; Neilson & Boag, 2002; Soetaert, Vincx, Wittoeck, & Tulkens, 1995; Tita, Desrosiers, Vincx, & Clement, 2002; Warwick & Gee, 1984). The spatial patterns of nematode communities are well documented in intertidal and subtidal zones of the Westerschelde. Soetaert et al. (1994) found maximum abundance of the majority of the species in the intertidal zone. Intertidal communities exhibited a well-developed community gradient with sediment depth, whereas the subtidal and channel communities showed distinct and in some cases distorted community patterns associated with large socio-economic pressure by dredging, pollution and consequently oxygen depletion.

This study deals with the spatial heterogeneity of nematode associations on a small intertidal, estuarine flat. Differences in structure of the communities were initially analysed in terms of depth-integrated characteristics for a high number (five) of replicate samples (collected at a metre scale) at three geographically separated (at kilometre scale) and chemico-physically diverging sites. In addition to the comparison of bulk characteristics, spatial differences in community structure are established by microscaled vertical profile analysis of the same community parameters. Whether changing environmental conditions over a small system like the Molenplaat are reflected in the vertical distribution pattern of nematodes species and result in shifting community characteristics with depth in the sediment was examined.

2. Study site

The Molenplaat (51°26'N, 3°57'E) is a small intertidal flat (2–3 km²), located in the turbid, nutrient-rich and heterotrophic Westerschelde estuary (Fig. 1). Salinity in this region of the estuary varies between 20 and 25 (Herman, Middelburg, Widdows, Lucas, & Heip, 2000). The flat is characterised by a high diversity of sediment types over a small distance. The ecology of the tidal flat has been well studied, and detailed background information on pigment distributions (Barranguet et al., 1997; Lucas & Holligan, 1999), microphytobenthos production (Barranguet & Kromkamp, 2000; Barranguet, Kromkamp, & Peene, 1998), photosynthetic activity (Kromkamp et al., 1998), microphytobenthos resuspension (Lucas et al., 2000), nematode feeding behaviour (Hamels, Moens, Muylaert, & Vyverman, 2001; Moens, Herman, Verbeeck, Steyaert, & Vincx, 2000; Moens, Van Gansbeke, & Vincx, 1999), nematode tidal migration (Steyaert, Herman, Moens, & Vincx, 2001) and microbenthic (Hamels et al., 1998) and macrofaunal (Herman et al., 2000) communities is available. A review of results of the project, dealing with benthic community structure and sediment processes on the Molenplaat is given by Herman, Middelburg, and Heip (2001).

The three sites were selected on the basis of their sediment characteristics (Table 1). Site 2 (57°2'N, 26°3'E) has the finest sediment, site 3 (56°8'N, 26°4'E) is more dynamic and sandier and site 1 (57°3'N, 26°15'E) has intermediate characteristics (Herman et al., 2000; Widdows, Brinsley, Salkeld, & Lucas, 2000). Estimates of bottom shear stress (maximal value during a tidal cycle) were produced by a hydrodynamic model. Values were 0.36 Pa for site 2, 0.43 Pa for site 1 and 1.15 Pa for site 3 (Van de Koppel, Herman, Thoolen, & Heip, 2001). The average period of emersion varies between 4.5 h (site 1) and 7 h (sites 2 and 3) per tidal cycle.

Microphytobenthic production (Barranguet et al., 1998), as well as microphytobenthic biomass, as reflected in pigment concentrations, was very high for all three sites in June 1996 (Table 2). The distribution of chlorophyll *a* in the sediment was reported by Hamels et al. (1998). All pigments (chlorophyll *a*, chlorophyll *c*, fucoxanthin) were negatively correlated ($p < 0.05$) with depth in the sediment. Maximum chlorophyll *a* concentrations were recorded at site 1; site 2 had intermediate values and site 3 had the lowest values. For the three sites, the bulk of the algal pigments was present in the top 2 cm of the sediment. For sites 2 and 3 pigment concentrations decreased gradually with depth whereas for site 1 a more distorted depth pattern was recorded (Hamels et al., 1998).

3. Materials and methods

In June 1996, the three sites (at kilometre scale) were sampled during low water at daytime (sediments were

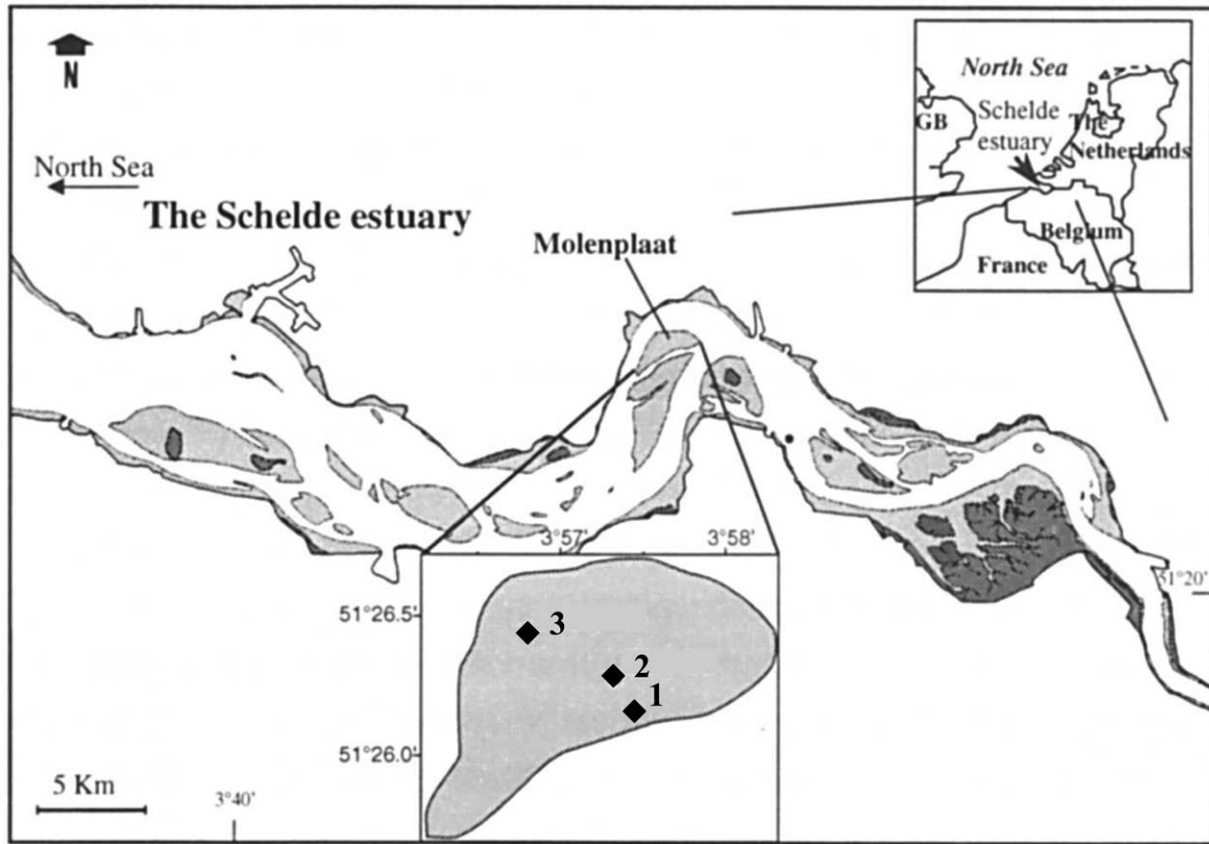


Fig. 1. Location of the sampling sites on the Molenplaat.

Table 1
Sediment characteristics for the three sites

Site depth (cm)	Median grain size (µm)			Fine sand fraction (%)			Medium sand fraction (%)		
	1	2	3	1	2	3	1	2	3
0–1	136.79	76.95	169.58	49.31	24.46	80.71	6.13	0.55	6.97
1–2	147.62	65.61	173.14	57.31	22.78	81.12	7.26	0.53	7.47
2–3	127.63	44.81	173.14	43.77	12.37	79.18	6.87	0.40	8.46
3–4	132.13	68.87	167.24	45.98	15.85	78.98	7.06	1.14	7.39
4–5	132.13	0.01	173.14	46.61	1.57	77.87	6.56	0.04	9.33
5–6	161.54	87.78	176.78	45.83	22.20	78.52	18.23	1.70	10.18
6–7	192.11	95.39	171.94	48.11	27.71	77.18	24.94	0.75	9.17
7–8	196.15	96.72	174.34	49.72	27.80	73.61	25.74	0.88	11.95
Mean	153.26	67.02	172.41	48.33	19.34	78.40	12.85	0.75	8.87

Site depth (cm)	Silt (%)			Very fine sand fraction (%)		
	1	2	3	1	2	3
0–1	24.13	43.32	3.96	19.74	31.67	8.14
1–2	11.73	48.67	4.04	No data	28.01	7.20
2–3	17.05	59.26	3.82	31.44	27.97	8.10
3–4	15.35	45.51	3.88	30.74	37.49	9.42
4–5	14.25	0.42	3.68	31.81	No data	8.47
5–6	9.59	28.34	3.22	26.29	47.58	No data
6–7	7.24	26.94	3.67	19.71	44.56	9.26
7–8	6.71	26.61	4.03	17.82	44.66	9.15
Mean	13.26	34.88	3.79	25.36	37.42	8.53

Data out of the ECOFLAT database (Herman et al., 2001).

Table 2

Photosynthetic pigments for the three sites (mg m^{-2}); chlorophyll *a* data from Hamels et al. (1998)

Site depth (cm)	Chlorophyll <i>a</i>			Chlorophyll <i>c</i>			Fucoxanthin		
	1	2	3	1	2	3	1	2	3
0–1	939.39	175.20	16.48	123.77	22.95	2.96	465.62	140.34	8.55
1–2	79.14	94.88	22.42	15.10	16.33	4.89	48.53	64.05	12.63
2–3	130.06	50.14	10.03	23.53	7.18	2.77	68.46	30.92	7.90
3–4	195.93	25.96	6.29	34.79	2.48	1.12	93.23	14.84	5.61
4–5	51.03	11.42	2.59	8.30	0.98	0.31	23.60	7.16	2.61
5–6	103.12	7.13	1.35	16.61	0.62	No data	44.03	4.30	1.20
6–7	12.89	1.15	0.72	2.23	0.06	No data	9.38	2.37	0.78
7–8	12.36	5.50	1.48	1.66	0.35	No data	10.04	4.66	1.31
Mean	190.49	46.42	7.67	28.25	6.37	2.41	95.36	33.58	5.07

exposed to air). At each of the three sites, five cores (3.6 cm diameter) were taken at 10 m intervals. The samples were divided into 12 horizontal slices (0–0.5, 0.5–1, 1–1.5, 1.5–2, 2–3, 3–4, 4–5, 5–6, 6–8, 8–10, 10–15, 15–20 cm) immediately after sampling and fixed in a hot (70 °C) neutral 4% formaldehyde solution. Meiofaunal organisms retained on a 38 μm sieve were extracted from the sediment by centrifugation with Ludox (density 1.18) (Heip, Vincx, & Vranken, 1985). A 1 mm sieve excluded all macrobenthos. For each slice, all nematodes were counted after staining with Rose Bengal and 120 nematodes were picked out randomly and mounted on Cobb slides for identification to species level. The nematodes were grouped into four feeding guilds, according to the feeding type classification of Wieser (1953).

Samples for the pigment analyses were taken from contiguous cores for meiofauna samples. Particle size distribution was determined by laser diffraction using a Malvern particle sizer. Analytical techniques for determination of the pigment content were described by Hamels et al. (1998), and for the organic carbon content in Herman et al. (2000).

Horizontal and vertical patterns in nematode abundance and community composition were analysed using ordination techniques from the PC-ORD for Windows package (version 4.20, McCune & Mefford, 1999). Through ordination samples are ordered along axes according to their resemblances. A detrended correspondence analysis (DCA) was applied on vertically integrated densities (summation of all depth layers) to test the variability between replicates. Subsequently, another DCA was used to assess total community variability based on non-transformed relative abundances. Species rarer than $F_{\text{max}}/5$ (F_{max} is the frequency of the commonest species) down-weighted in proportion to their frequency. Nematode diversity was expressed as Hill indices N_0 and N_1 (Hill, 1973). In order to test for significant differences in depth-integrated (total sediment column) density and diversity between the three sites, the non-parametric Kruskal–Wallis analysis by rank and pairwise multiple comparison tests were used

(Conover, 1971). If assumptions were met, a univariate two-way analysis of variance (ANOVA) was used to test for significant differences in depth distribution between the three sites. A ‘split-plot’ design was constructed with replicates nested within ‘site’, however, not within ‘depth’, following Steyaert et al. (2001). All data were $\log(x+1)$ transformed prior to analysis. Non-parametric Spearman rank correlation coefficients were calculated to determine relationships between diversity and environmental variables along a depth gradient.

4. Results

4.1. Density and species composition

4.1.1. Horizontal

A significant difference in total (depth-integrated) nematode densities ($p \leq 0.05$) was found when comparing all three sites (Table 3). A post hoc multiple comparison revealed only significant differences between site 1 and 2. Highest abundances were recorded at site 1, lowest abundances at site 2 and intermediate abundances at site 3. In addition, DCA, based on species densities of bulk samples, separated all three sites and illustrated the high similarity between the five replicates of each site (Fig. 2). Eigen-values were 0.71 and 0.05 for the first and second axes, respectively.

Total species number was in the same range at all three sites. In total, 76 nematode species were identified: 52 at site 1, 41 at site 2 and 54 at site 3. Site 1 had 11 species exclusive to that site while sites 2 and 3 had 9 and 17

Table 3

Total number of species, vertically integrated nematode density (ind. 10 cm^{-2}) and vertically integrated nematode diversity for the three sites (mean values from five samples \pm SD)

	Site 1	Site 2	Site 3
Total number of species	52	41	54
Nematodes density	2990 \pm 818	1560 \pm 699	2090 \pm 666
N_0 diversity	28.4 \pm 3.58	25.2 \pm 1.92	32.0 \pm 2.90
N_1 diversity	9.1 \pm 0.94	8.0 \pm 2.15	8.6 \pm 0.77

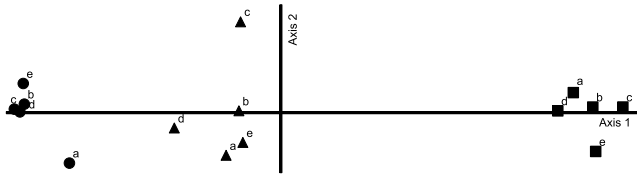


Fig. 2. Results of DCA axes 1 and 2, based on species abundances (site 1, triangles; site 2, squares; site 3, circles; letters represent replicates).

unique species, respectively. Average relative abundances for the dominant species are shown in Table 4. Nematode species were termed dominant when present in at least 25 of the 180 slices or in at least seven slices with a minimum relative abundance of 40% (restrictions based on practical considerations). Based on the dominant species, three different species assemblages could be recognised at the three sites: *Theristus blandicor*, *Ascolaimus elongatus* and *Eleutherolaimus amasi* were most dominant in site 1 (60.5% of total community); *Tripyloides gracilis*, *Viscosia viscosa* and *Ptycholaimellus ponticus* in site 2 (67.8%); *T. blandicor* and *Enoploides longispiculosus* in site 3 (60.6%).

4.1.2. Vertical profiles

ANOVA split-plot analysis (Table 5) showed significant differences in depth distribution of total nematode densities (Fig. 3) and of densities for each dominant species (Figs. 4–6) among the three sites. Nematode density at site 1 (Fig. 3) was highest at the sediment surface and decreased gradually with depth. The maximum density of each of the dominant species at site 1 occurs at different depth layers (Fig. 4), which indicates a gradual shifting of the nematode community with depth in the sediment at site 1. Maximum

Table 4

Relative abundances and feeding type (Wieser, 1953) of the dominant species

	Feeding type	Site 1	Site 2	Site 3
<i>Theristus blandicor</i>	1B	20.1	0.6	30.5
<i>Viscosia viscosa</i>	2B	9.5	15.3	4.0
<i>Ascolaimus elongatus</i>	1B	26.0	2.6	3.1
<i>Eleutherolaimus amasi</i>	1B	14.4	0.2	3.9
<i>Theristus pertenuis</i>	1B	5.2	0.1	5.6
<i>Microlaimus marinus</i>	2A	2.7		2.7
<i>Trefusia helgolandica</i>	1A	1.0		2.3
<i>Enoploides longispiculosus</i>	2B	0.5	0.1	30.1
<i>Daptonema riemanni</i>	1B	0.1	0.2	3.9
<i>Microlaimus acinaces</i>	2A	0.3		2.2
<i>Tripyloides gracilis</i>	1B	2.9	40.1	0.2
<i>Ptycholaimellus ponticus</i>	2B	0.3	12.4	0.1
<i>Daptonema tenuispiculum</i>	1B	0.8	9.7	
<i>Calytronema maxweberi</i>	2B	1.6	2.8	
<i>Theristus acer</i>	1B	0.1	3.1	0.1
<i>Sabatieria pulchra</i>	1B	1.4	0.9	0.1

Values in bold are the dominant species per site.

abundance of *Viscosia viscosa* was found in the upper sediment layer (0–0.5 cm). *Ascolaimus elongates*, *Eleutherolaimus amasi* and *Theristus pertenuis* showed peak abundance at 0.5–1 cm depth; *Microlaimus marinus* and *Trefusia helgolandica* at intermediate sediment depths (2–3; 6–8 cm, respectively), and finally *Theristus blandicor* at 5–10 cm depth in the sediment (Fig. 4).

At site 2, the total nematode density was very high at the sediment surface and decreased gradually with depth (Fig. 3). Here 95% of the nematode community was confined to the upper 2 cm of the sediment, compared to 73% in site 1. This steep gradient in depth is also reflected in the individual species distributions (Fig. 5).

Table 5

Univariate ANOVA tests (df = 2, 11, 18 for site, depth, site×depth, respectively)

	Site		Depth		Site×depth	
	F	p	F	p	F	p
Nematode community	8.982	0.009	32.725	0.000	11.301	0.000
<i>Ascolaimus elongatus</i>	58.182	0.000	47.698	0.000	26.079	0.000
<i>Calytronema maxweberi</i>	16.247	0.002	12.628	0.000	3.637	0.000
<i>Daptonema tenuispiculum</i>	107.517	0.000	23.619	0.000	13.099	0.000
<i>Eleutherolaimus amasi</i>	30.397	0.000	5.511	0.000	7.896	0.000
<i>Enoploides longispiculosus</i>	147.509	0.000	34.552	0.000	18.075	0.000
<i>Microlaimus acinaces</i>	20.651	0.001	3.320	0.005	2.763	0.001
<i>Microlaimus marinus</i>	57.632	0.000	8.647	0.000	8.104	0.000
<i>Ptycholaimellus ponticus</i>	205.658	0.000	49.261	0.000	28.031	0.000
<i>Sabatieria pulchra</i>	10.489	0.006	5.446	0.000	3.431	0.000
<i>Theristus acer</i>	40.125	0.000	14.292	0.000	16.890	0.000
<i>Theristus blandicor</i>	12.632	0.003	22.656	0.000	7.444	0.000
<i>Theristus pertenuis</i>	41.311	0.000	6.514	0.000	5.858	0.000
<i>Theristus riemanni</i>	9.746	0.007	5.386	0.000	4.449	0.000
<i>Trefusia helgolandica</i>	9.802	0.007	8.781	0.000	2.823	0.000
<i>Tripyloides gracilis</i>	23.913	0.000	49.971	0.000	15.236	0.000
<i>Viscosia viscosa</i>	17.188	0.001	40.367	0.000	7.210	0.000
N ₀ diversity	3.929	0.065	8.664	0.000	6.227	0.000
N ₁ diversity	5.806	0.028	10.686	0.000	5.605	0.000

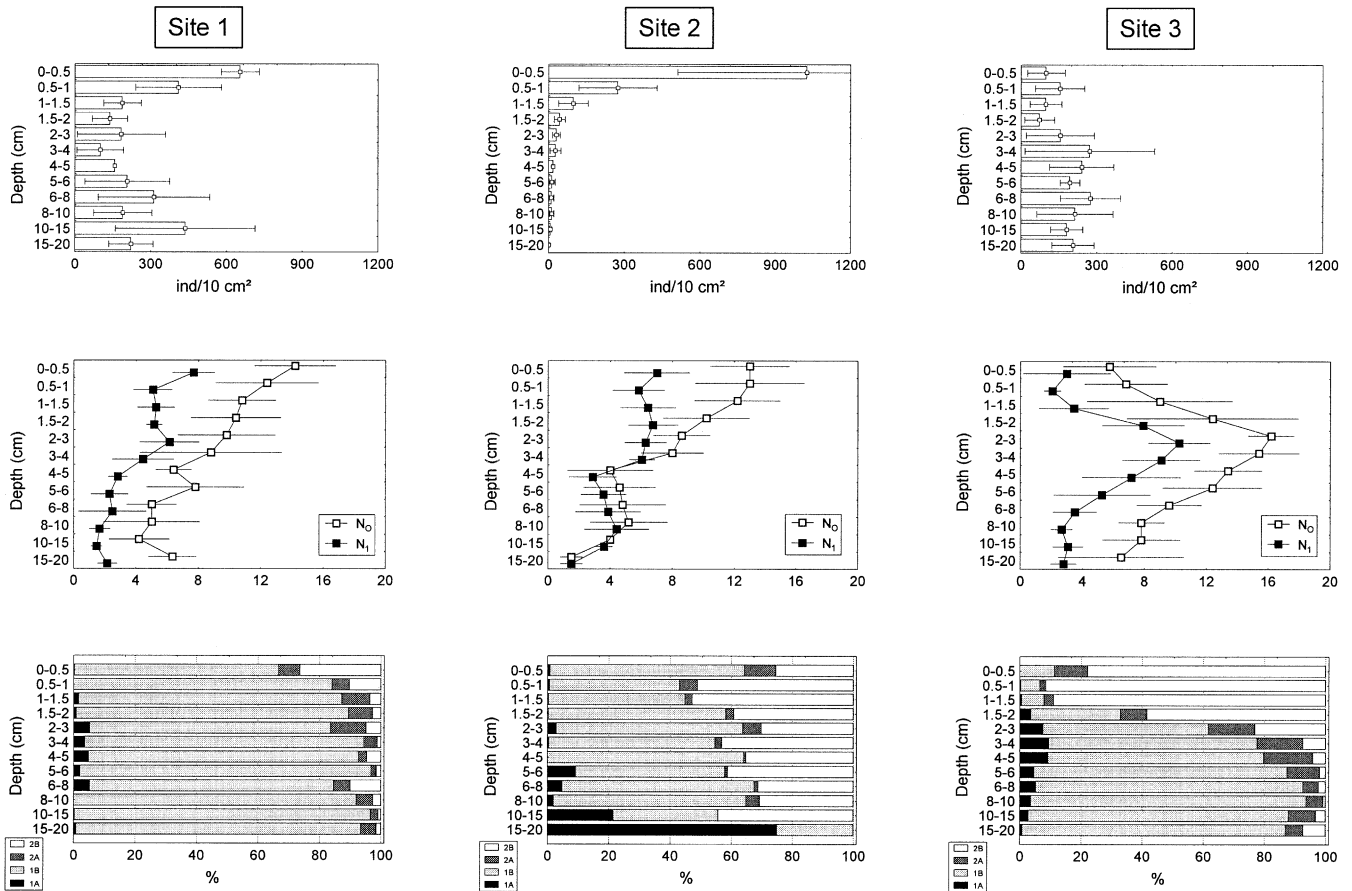


Fig. 3. Nematode abundance, nematode diversity and feeding type distribution with depth.

All dominant species, *Calyptronema maxweberi*, *Daptonema tenuispiculum*, *Ptycholaimellus ponticus*, *Theristus acer*, *Tripyloides gracilis* and *Viscosia viscosa*, showed maximum abundances in the top 1 cm of the sediment, except for *Sabatieria pulchra* and *Theristus blandicor*, which were more uniformly distributed down the core.

For site 3, total nematode density remained low and relatively constant until 10 cm depth (Fig. 3). Deeper down, the abundance tended to decrease very slowly. The individual species (Fig. 6) belong to two vertically segregated species assemblages. The ‘*Enoploides longispiculosus*-assemblage’ situated at the top 1.5–2 cm of site 3, is characterised by high abundances of *E. longispiculosus*, with a maximum abundance recorded at 0.5–1 cm depth and other species that also conform to this zonation. The ‘*Theristus blandicor*-assemblage’ is located in deeper sediment layers (from 3 cm onwards). The most important species here is *T. blandicor*, which has its maximum abundance at 5–6 cm depth. A similar feature is found for the less numerous species *Ascolaimus elongatus*, *Eleutherolaimus amasi*, *Microlaimus marinus*, *Microlaimus acinaces*, *Trefusia helgolandica*, *Theristus pertenuis* and *Theristus riemanni* however species’ preferences are not monotonic in this zone. Their maximum abundance is between 8 and 10 cm. All

species, except *Viscosia viscosa*, can be attributed to one of both subcommunities. *V. viscosa* has highest abundance between 1.5 and 2 cm depth.

The existence of three different nematode assemblages at the three sites (based on dominant species and depth profiles), is confirmed by the DCA (Fig. 7). The first ordination axis has an eigen-value of 0.787; the second axis has an eigen-value of 0.42. The third axis (not depicted) has a low eigen-value (0.098) and yields no additional information. Site 1 consists of a gradually shifting nematode community with depth. This is illustrated by the samples, which are placed along a depth gradient parallel to the first axis. It should be noted that depth layers are plotted relatively far from each other (especially in upper depth layers). The samples of upper sediment layers of site 3 (0–0.5; 0.5–1; 1–1.5 cm) appear separate from those of deeper sediment layers (Fig. 7), pointing again to the existence of two vertically separated subcommunities at site 3. The analysis further indicates that there is a similarity between deeper layers of site 1 and 3 (beneath 2 cm) and the deepest layer of site 2 (15–20 cm), all dominated by *Theristus blandicor*. Site 2 is characterised by a surface nematode association that declines with depth. Vertically sectioned samples, exclusively the deepest sediment layer (15–20 cm), of site 2 are

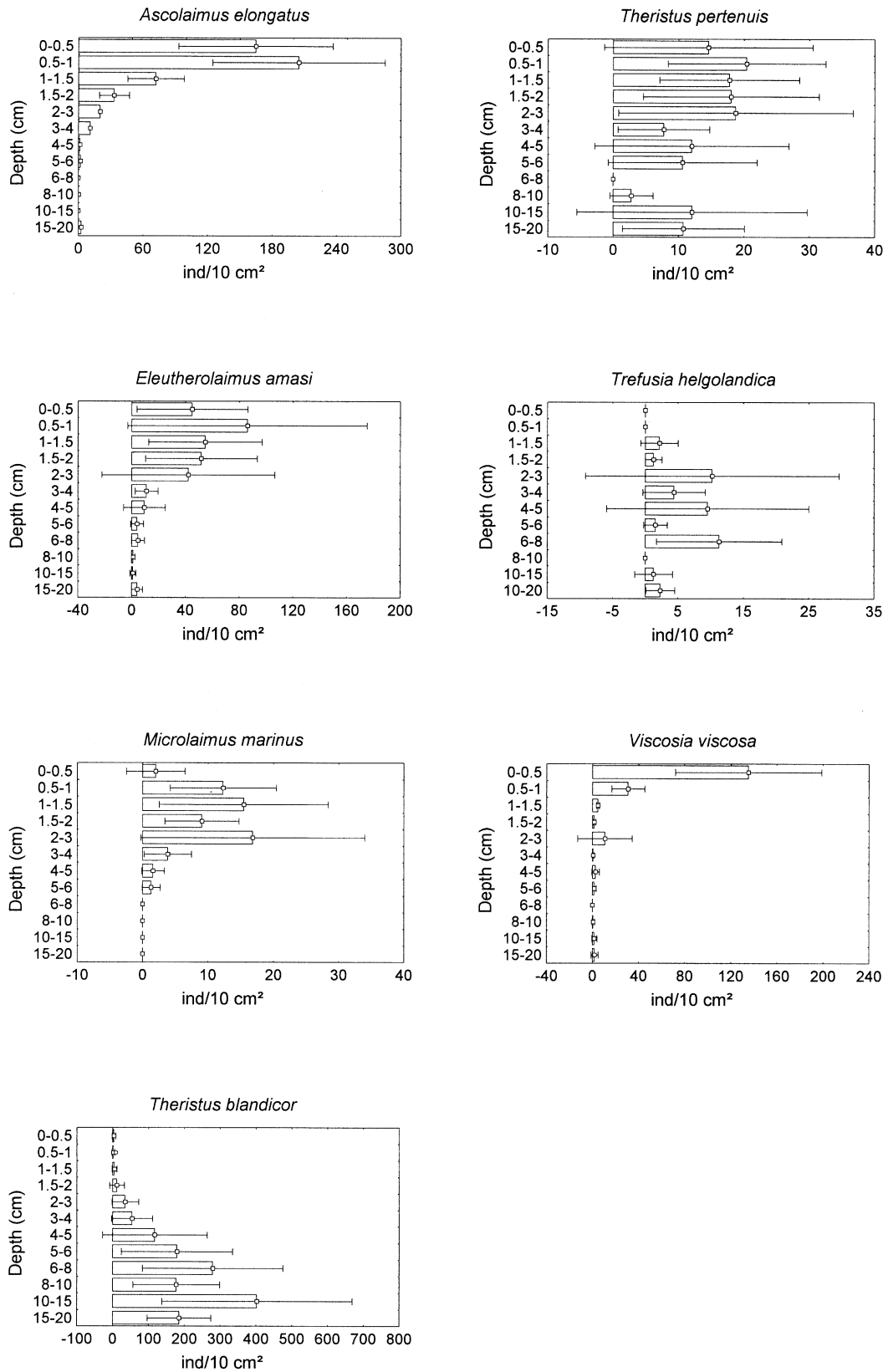


Fig. 4. Depth distribution of the dominant species in site 1 (note different width in sediment slices on Y-axes).

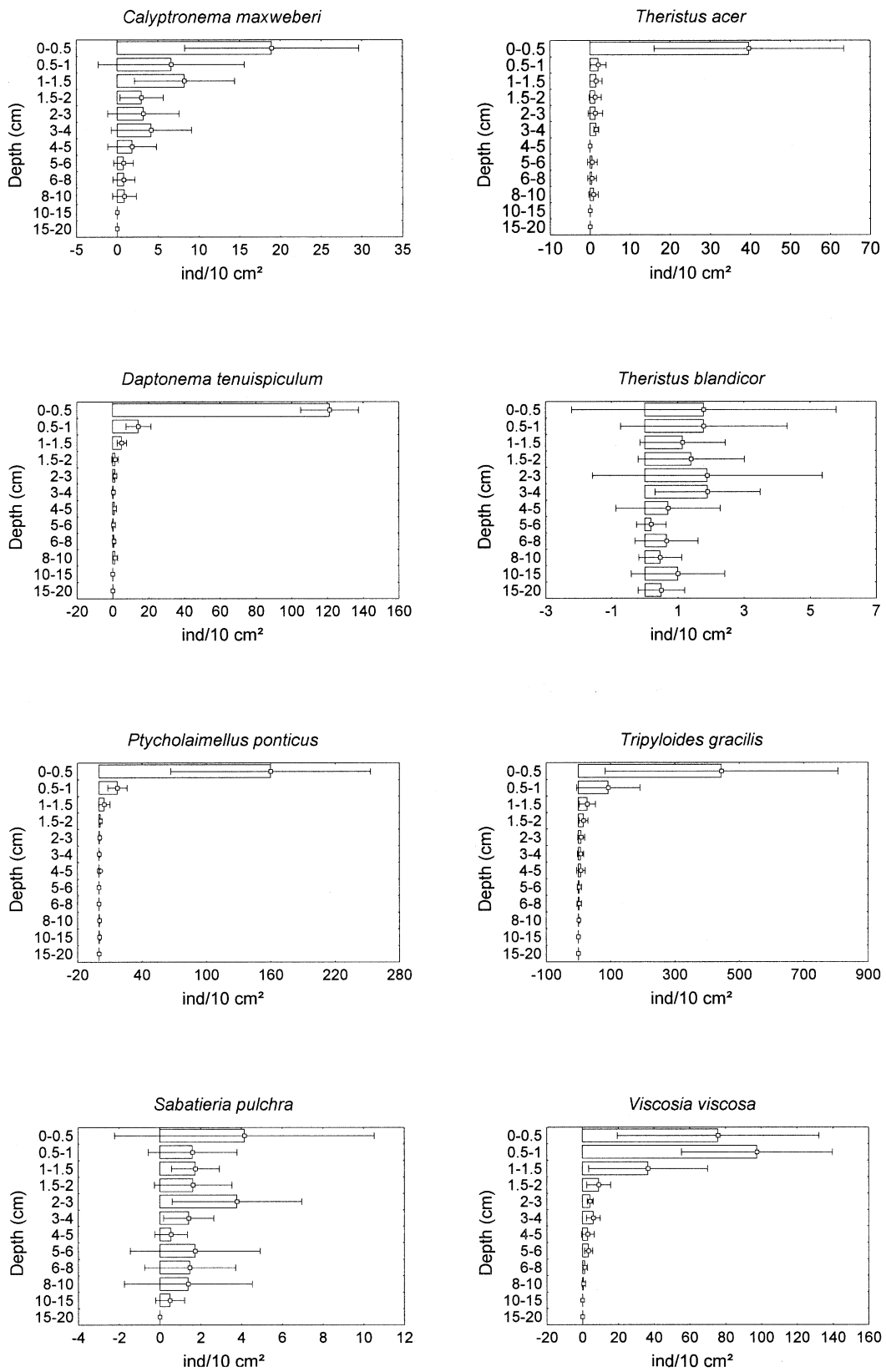


Fig. 5. Depth distribution of the dominant species in site 2 (note different width in sediment slices on Y-axes).

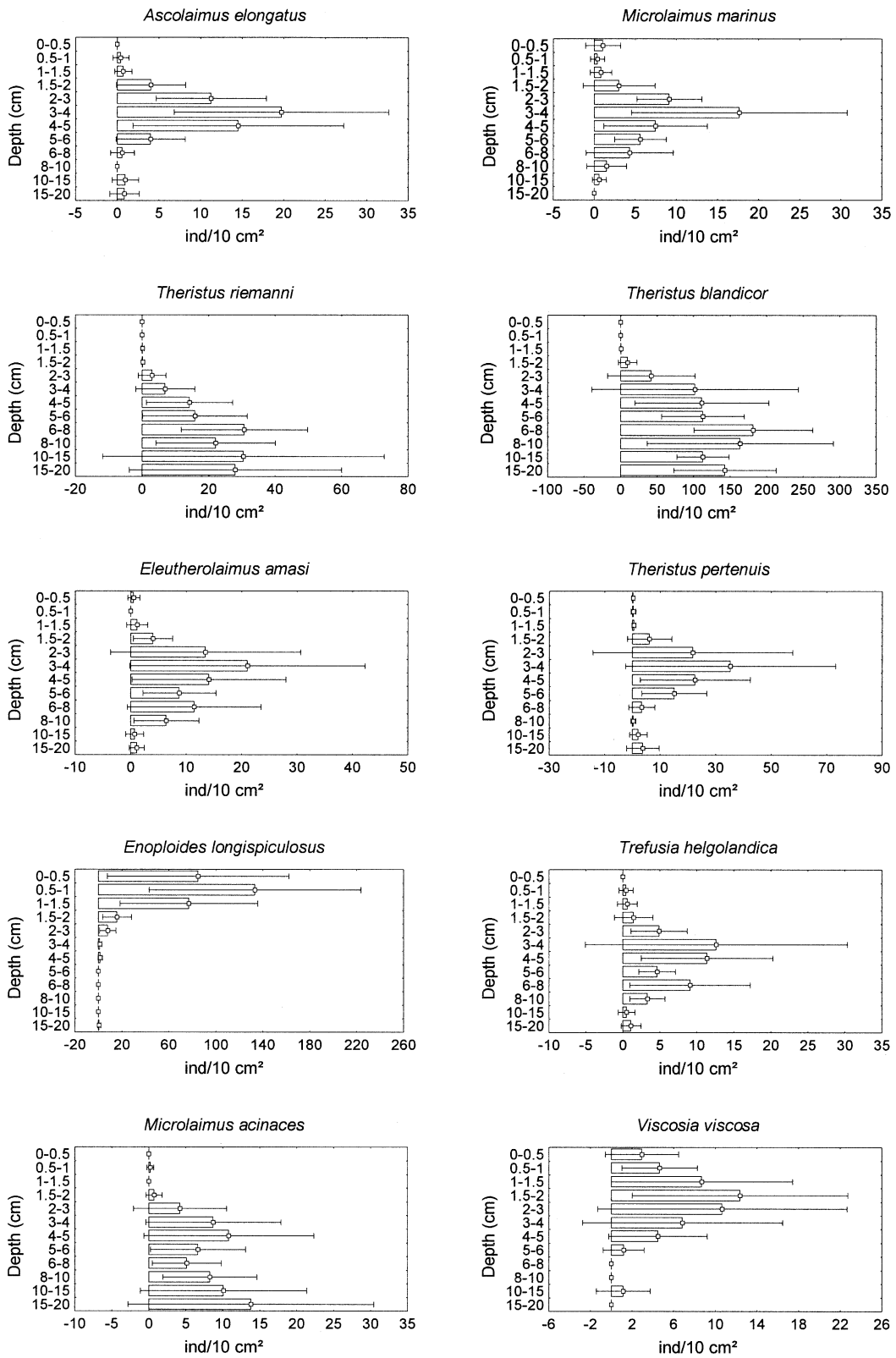


Fig. 6. Depth distribution of the dominant species in site 3 (note different width in sediment slices on Y-axes).

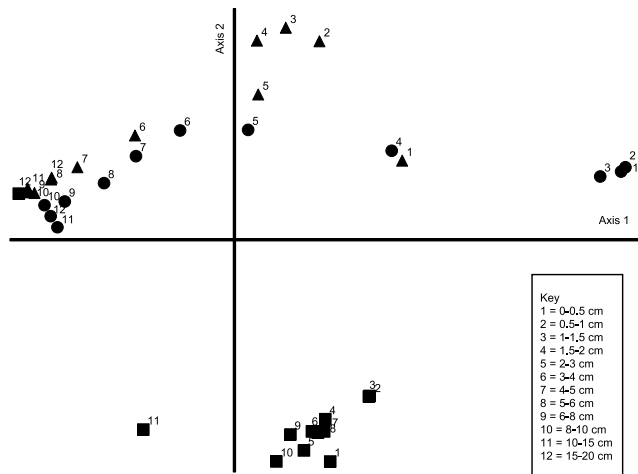


Fig. 7. Results of DCA axes 1 and 2, based on relative abundances (site 1, triangles; site 2, squares; site 3, circles).

clustered closely together and species associations seem to be similar at all depth layers.

4.1.3. Feeding type distribution

Non-selective deposit feeders dominated site 1. In the surface layer of the sediment, non-selective deposit feeders are slightly less important, due to the presence of predators/omnivores, and to a lesser degree, epistrate feeders (Fig. 3). Selective deposit feeders and epistrate feeders form a small fraction of the nematode community in all depth layers. At site 2, two feeding types are well represented: non-selective deposit feeders and predators/omnivores. Selective deposit feeders were only important in deeper layers. At site 3, the communities inhabiting the top layers (to about 1.5 cm) are dominated by the predator/omnivore *Enoploides longispiculosus*. The deeper layers, from 2 cm onwards are characterised by the dominance of the non-selective deposit feeder, *Theristus blandicor*. The changes in trophic strategy with depth reflect the change in single species dominance at site 3.

4.1.4. Diversity

For total (integrated depth layers) number of species (N_0), a significant difference ($p \leq 0.05$) was found between sites 2 and 3 (Table 3). No significant differences between sites existed for N_1 diversity. The ANOVA split-plot analysis demonstrated significant differences among the three sites with respect to the variation in diversity with depth (Fig. 3, Table 5). Site 3 is clearly different from the other sites. At site 3, N_0 and N_1 are low at the surface of the sediment and a maximum diversity exists at around 3–4 cm depth precisely where there is a change from a surface community to a deeper community. At sites 1 and 2, species diversity within the sediment exhibited a different profile. At both sites, diversity was highest at the sediment surface and decreased with depth. At site 2, there was a sharper decrease of diversity below 8 cm compared to site 1; this can be linked to a decrease in density.

At site 1 there was positive correlation between diversity and chlorophyll *a*, silt and a negative correlation between medium sand content (Table 6). At site 2, diversity was positively correlated with chlorophyll *a* and silt content. At site 3, no significant correlations were found between diversity and any of the environmental factors.

5. Discussion

5.1. Nematode densities

Previous studies addressing nematode community structure on intertidal flats, have dealt with diversity, distribution on different scales and production (Hogue & Miller, 1981; Joint et al., 1982; Ott, 1972; Pickney & Sandulli, 1990; Van Es, Van Arkel, Bouwman, & Schroder, 1980; Warwick & Price, 1979). In general, extremely high abundances of meiofauna, with nematodes always the dominant taxon, are characteristic of sheltered muddy regions of estuaries (Heip et al., 1985). The present study shows a significant difference in total

Table 6
Spearman rank order correlations for the three sites

		Site 1		Site 2		Site 3	
		Spearman <i>R</i>	<i>p</i> -Level	Spearman <i>R</i>	<i>p</i> -Level	Spearman <i>R</i>	<i>p</i> -Level
Chlorophyll <i>a</i>	N_0 diversity	0.865	0.003	0.728	0.026	0.134	0.731
Chlorophyll <i>a</i>	N_1 diversity	0.762	0.017	0.711	0.032	0.251	0.515
Organic C	N_0 diversity	0.470	0.202	0.117	0.764	–0.287	0.454
Organic C	N_1 diversity	0.371	0.325	0.134	0.731	–0.226	0.558
Silt	N_0 diversity	0.848	0.004	0.828	0.006	–0.444	0.232
Silt	N_1 diversity	0.895	0.001	0.862	0.003	–0.326	0.391
Fine sand	N_0 diversity	–0.136	0.728	0.142	0.715	0.251	0.515
Fine sand	N_1 diversity	–0.134	0.731	0.126	0.748	0.318	0.404
Medium sand	N_0 diversity	–0.763	0.017	–0.176	0.651	–0.159	0.683
Medium sand	N_1 diversity	–0.828	0.006	–0.192	0.620	–0.226	0.559

nematode abundance between the muddy site (site 2, lowest densities) and site 1, which has fine sandy sediment with high silt content and the highest nematode densities. The higher densities in site 1, together with the higher benthic primary production (Herman et al., 2001), the higher autotrophic biomass (Hamels et al., 1998) and the larger macrobenthos stock (Herman et al., 2000), all point to a higher productivity in site 1. This site is located at the border of the tidal flat and the open water and it is therefore exposed to stronger hydrodynamic forces compared to site 2 (Widdows et al., 2000). As such it is likely that the input and output of fresh organic material may be larger and is the basis of the highly productive system.

The presence of macrofauna may affect the nematode densities although in different ways. Besides alterations of the chemical and physical properties of the sediment by macrobenthos, their effect may also be linked to feeding activities (e.g. Austen, Widdicombe, & Villano-Pitacco, 1998; Ólafsson, Elmgren, & Papakosta, 1993). Site 3, and in minor degree, site 1, with higher current velocities and bottom shear stress (Van De Koppel et al., 2001) are characterised by higher densities of surface deposit feeders, while site 2, where sedimentation rates were high (Schmidt et al., 1999 in Herman et al., 2001), is characterised by suspension feeders (Herman et al., 2001). Deposit feeders may have a predatory effect due to coincidental consumption of nematodes while feeding. Alternatively, the disturbance activity may stimulate microbial growth and increased sediment oxygenation, providing an increase in food and spatial resources, which in turn stimulated the nematodes (Reise, 1983). Suspension feeders may stimulate nematode abundance through biodeposition of organic carbon. However it remains unclear what the macrofauna–meiofauna interaction is at the Molenplaat.

The differences in sediment characteristics, hydrodynamic conditions (reflected by current velocity and bottom shear stress) and productivity did not affect diversity of the three investigated sites and only partially total nematode abundance (only two of the three sites differed in nematode abundance). Detailed investigation of vertical depth profiles however, revealed differences that may relate to environmental factors.

Generally, the vertical distribution pattern of nematodes in silty sediments is well established: abundances are extremely high at the sediment surface or subsurface and subsequently decrease steeply with depth (for an overview, see Heip et al., 1985). This trend was evident in site 1 and was even more pronounced in site 2 of the Molenplaat. In the more sandy conditions of site 3, nematode abundance remains generally lower and fluctuates greatly with depth. Oxygen penetration and the occurrence of sulphide have been linked many times to the depth distribution of nematodes (Giere, 1993; Heip et al., 1985; Hendelberg & Jensen, 1993; Platt, 1977;

Wetzel et al., 1995). In the sediment of site 3, which is highly bioturbated by macrobenthos, oxygen penetration might be several centimetres in the proximity of burrows, whereas site 2 is the least bioturbated (C. Barranguet, personal communication). Besides a number of biotic (e.g. resource availability and distribution) and abiotic (e.g. compaction of sediment) interactions, oxygen distribution is thought to be one of the important regulating factors in explaining the obvious discrepancy in vertical distribution patterns on the Molenplaat.

5.2. Community composition

From studies which have dealt with the vertical distribution of free-living nematodes at the species level (e.g. Blome, 1983; Hendelberg & Jensen, 1993; Jensen, 1987; Joint et al., 1982; Soetaert et al., 1994; Steyaert et al., 1999; Warwick & Gee, 1984), it is clear that some nematode species show a consistent depth distribution in different areas, which suggests species-specific depth preferences. As the auto-ecological information on free-living nematodes is still very scarce, the causal factors for this depth preference are not yet completely clear. It has been suggested that the biogeochemical properties of the sediment might control the depth distribution of some species (Bouwman, Romeyn, Kremer, & Van Es, 1984; Jensen, 1981, 1987; Platt & Lamshead, 1985; Jensen & Aagaard, 1992; Steyaert et al., 1999). This might explain the surface dominated community of the silty sediment (site 2), where oxygen penetrated only into the upper millimetres of the sediment (C. Barranguet, personal communication).

The factors determining the vertical distribution in the fine sandy sediments may act in combination with biotic interactions. Joint et al. (1982) argued that interspecific competition gives rise to vertical niche segregation. As such, fine scale vertical stratification may play a role in allowing species with similar food requirements and feeding behaviour to co-exist in the same locality. The present study supports this finding for the highly productive system of site 1. Here, a gradual shifting of the nematode community was recorded, as a result of the succession of maximum density peaks of dominant species with depth. This sediment was characterised by a high percentage of non-selective deposit feeders (e.g. *Theristus blandicor*, *Ascolaimus elongatus*, *Eleutherolaimus amasi*, *Theristus pertenuis*).

In the sediment of site 3, two vertically segregated species assemblages were observed. The upper assemblage (to about 1.5 cm depth) was dominated by *Enoploides longispiculosus*. The lower assemblage (below 2 cm) was dominated by *Theristus blandicor* and a number of less common species (Fig. 6). In earlier studies, these consistent depth profiles were also reported for some species: *E. longispiculosus* being a true

surface-dweller (Soetaert et al., 1994) while *T. blandicor*, *Microaimus marinus*, *Theristus pertenuis* and *Theristus riemanni* were considered as ‘deep-dwelling’ species (Blome, 1983; Soetaert et al., 1994). The existence of these two vertically segregated assemblages on the Molenplaat is probably due to a combination of factors, of which the most important might be related to food preferences and the strong hydrodynamic regime at the site. As sediment granulometry appears to be of more importance in controlling the fauna in the upper sediment layers, the similarity of the deeper nematode communities—caused by the dominance of *T. blandicor*—at sites 1 and 3 is particularly interesting. The environmental regime at depth seems to result in similar faunal assemblages irrespective of sediment granulometry. From its dominance in deeper sediment layers at site 1 and 3 and the apparent lack of depth preference in site 2 it can be concluded that *T. blandicor* persists in sediments with restrictive conditions for other nematode species. It appears that this species is capable of surviving anoxic conditions and is able to exploit the available food resources at depth. Such life conditions have often been described for *Sabatieria* species, which are typical inhabitants for deeper sediment layers of muddy intertidal and subtidal sediments (Hendelberg & Jensen, 1993; Soetaert & Heip, 1995; Steyaert et al., 1999). One *Sabatieria* species, *Sabatieria pulchra*, was observed in this study.

5.3. Diversity

Nematode community diversity has been associated with sediment composition, oxygen, salinity, stress and organic enrichment (Austen et al., 1998; Essink & Keidel, 1998; Giere, 1993; Soetaert et al., 1994; Steyaert et al., 1999; Warwick & Clarke, 1993; Warwick, McEvoy, & Trush, 1997). This study documented a high nematode species richness on a small area (the Molenplaat) of approximately 2–3 km². This high diversity may be largely the result of the heterogeneous geochemical and physical characteristics on the tidal flat. Moreover, the twofold difference in total (over the whole sediment column) number of species and averaged (over five replicates) total number of species per site, also illustrates the importance of small-scale effects. Thus, besides the mean differences, found on a large scale (between the three sites), local effects (on 10-m scale) of sedimentological characteristics are evident.

The difference in vertical profile of species richness found between site 3 and both site 1 and 2, is expected as a result of the wider range of microhabitats available for meiofauna in sandy sediments (site 3) compared to muddy sediments (site 1 and 2) (Heip & Decraemer, 1974). Within the sediment column diversity in site 3 is strongly variable and is only high at a depth of 2–5 cm. These higher values can be explained by the occur-

rence of both species from the upper ‘*Enoploides longispiculosus*-community’ and the lower ‘*Theristus blandicor*-community’. The lower diversity values in the uppermost layers of the sediment are related to low densities and probably result from the strong hydrodynamic regime and the tidal disturbance that prevail at the upper 2 cm of the sediment at site 3 (Herman et al., 2000; Widdows et al., 2000). It is therefore suggested that the hydrodynamic regime, which influences sediment granulometry, will predominantly affect the number of nematode species at the Molenplaat.

In many studies, correlation tests have been used to illustrate possible relationships between meiobenthos, in particular nematodes and copepods, and possible food sources (e.g. Blanchard, 1990; Danovaro, 1996; Danovaro et al., 1995; Findlay, 1981; Moens et al., 1999; Pickney & Sandulli, 1990; Santos, Castel, & Souza-Santos, 1996; Steyaert et al., 1999). These studies considered, in most cases, horizontal variation. When visualising vertical distribution patterns in the sediment, correlations between individual nematode species or feeding types and biotic variables are in many cases misleading. Moreover, correlation does not imply causation. The vertical distributions of factors such as pigments, bacteria and nutrients are often depth dependent. Therefore caution needs to be used when correlating depth profiles of different variables. In this study the diversity indices of site 1 and 2, both sediments characterised by a high silt content, were positively correlated with this silt content. Such a strong positive correlation of diversity and silt content with depth has also been found in muddy sediments of the Belgian coastal zone (Steyaert et al., 1999). In general, sediment granulometry exerts an important influence on the diversity of nematode communities. On a broad, horizontal scale, coarser sediments will enhance nematode diversity by creating a broad range of microhabitats (Heip & Decraemer, 1974). Diversity on a small spatial scale, within the sediment, is inversely related to the sediment granulometry of muddy sediments. The finer the sediment becomes, the more diverse the nematode community.

Conclusions

Differences in sediment composition in association with different hydrodynamic conditions at separate sites of an estuarine intertidal flat are reflected in total nematode abundances and in species composition. The heterogeneity was much higher at kilometre than at metre scales, at which level environmental conditions seemed more consistent. Species richness in combination with equitability did not differ among the three sites, when integrated over 20 cm depth in the sediment. In contrast, microscale vertical profile analysis illustrated

the presence of three distinctly different distribution patterns of species associations on the tidal flat: (1) In sandy sediment under strong hydrodynamic and food-stressed conditions a surface-dwelling nematode community of mainly a large predatory enoplid was observed living above a deposit feeding xyalid-microlaimid community in deeper sediment layers. These extreme environmental conditions resulted in low density and diversity. Diversity was highest at the interface between the two vertically separated communities as a result of co-existence of species from the upper and lower community. (2) In the finest sediment most nematode species were confined to the surface layers. Only a few could occasionally penetrate into deeper layers, resulting in a sharp decline in diversity and density with depth in the sediment. (3) At intermediate hydrodynamic and granulometric conditions there is a gradual shift from a diverse and abundant nematode community at the surface to a less diverse and less abundant one in the deeper sediment layers. The vertical changes in nematode composition resulted in a community similar to that found in the deeper sediment layers of the sandy site in terms of abundances, diversity and composition. Vertical profile analysis provides additional information over bulk sampling that is key to understanding horizontal patterns and their relation with environmental characteristics in nematode communities.

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References

- Austen, M., Widdicombe, C., & Villano-Pitacco, S. (1998). Effects of biological disturbance on diversity and structure of meiobenthic nematode communities. *Marine Ecology Progress Series* 174, 233–246.
- Barranguet, C., & Kromkamp, J. (2000). Estimating primary production rates from photosynthetic electron transport in estuarine microphytobenthos. *Marine Ecology Progress Series* 204, 39–52.
- Barranguet, C., Herman, P. M. J., & Sinke, J. J. (1997). Microphytobenthos biomass and community composition studied by pigment biomarkers: importance and fate in the carbon cycle of a tidal flat. *Journal of Sea Research* 38, 59–70.
- Barranguet, C., Kromkamp, J., & Peene, J. (1998). Factors controlling primary productivity and photosynthetic characteristics of intertidal microphytobenthos. *Marine Ecology Progress Series* 173, 117–126.
- Blanchard, G. F. (1990). Overlapping microscale dispersion patterns of meiofauna and microphytobenthos. *Marine Ecology Progress Series* 68, 101–111.
- Blome, D. (1983). Oekologie der nematoda eines sandstrandes der Nordseeinsel Sylt. *Mikrofauna des Meeresbodens* 88, 1–76.
- Bouwman, L. A., Romeyn, K., Kremer, D. R., & Van Es, F. B. (1984). Occurrence and feeding biology of some nematode species in estuarine aufwuchscommunities. *Cahiers de Biologie Marine de Roscoff* 25, 287–303.
- Conover, W. J. (1971). *Practical non parametric statistics* (462 pp.). New York: Wiley.
- Danovaro, R. (1996). Detritus–bacteria–meiofauna interactions in a seagrass bed (*Posidonia oceanica*) of the NW-Mediterranean. *Marine Biology* 127, 1–13.
- Danovaro, R., Croce, N. D., Eleftheriou, A., Fabiano, M., Papadopoulou, N., Smith, C., & Tselepidis, A. (1995). Meiofauna of the deep eastern Mediterranean Sea: distribution and abundance in relation to bacterial biomass, organic matter composition and other environmental factors. *Progress in Oceanography* 36, 329–341.
- Essink, K., & Keidel, H. (1998). Changes in estuarine nematode communities following a decrease of organic pollution. *Aquatic Ecology* 32, 195–202.
- Findlay, S. E. G. (1981). Small-scale spatial distribution of meiofauna on a mud- and sandflat. *Estuarine, Coastal and Shelf Science* 12, 471–484.
- Giere, O. (1993). *Meiobenthology: The microscopic fauna in aquatic sediments* (273 pp.). Berlin: Springer.
- Guo, Y., Somerfield, P. J., Warwick, R. M., & Zhang, Z. (2001). Large-scale patterns in the community structure and biodiversity of freeliving nematodes in the Bohai Sea, China. *Journal of the Marine Biological Association of the U.K.* 81, 755–763.
- Hamels, I., Moens, T., Muylaert, K., & Vyverman, W. (2001). Trophic interactions between ciliates and nematodes from an intertidal flat. *Aquatic Microbial Ecology* 26, 61–72.
- Hamels, I., Sabbe, K., Muylaert, K., Barranguet, C., Lucas, C., Herman, P., & Vyverman, W. (1998). Organisation of microbenthic communities in intertidal estuarine flats, a case study from the Molenplaat (Westerschelde Estuary, The Netherlands). *European Journal of Protistology* 34, 308–320.
- Heip, C., & Decraemer, W. (1974). The diversity of nematode communities in the southern North Sea. *Journal of the Marine Biology Association of the U.K.* 54, 251–255.
- Heip, C., Vincx, M., & Vranken, G. (1985). The ecology of marine nematodes. *Oceanography and Marine Biology Annual Review* 23, 399–489.
- Hendelberg, M., & Jensen, J. (1993). Vertical distribution of the nematode fauna in a coastal sediment influenced by seasonal hypoxia in the bottom water. *Ophelia* 37, 83–94.
- Herman, P. M. J., Middelburg, J. J., & Heip, C. H. R. (2001). Benthic community structure and sediment processes on an intertidal flat: results from the ECOFLAT project. *Continental Shelf Research* 21, 2055–2071.
- Herman, P. M. J., Middelburg, J. J., Widdows, J., Lucas, C. H., & Heip, C. H. R. (2000). Stable isotopes as trophic tracers: combining field sampling and manipulative labelling of food resources for macrobenthos. *Marine Ecology Progress Series* 204, 79–92.
- Hill, M. O. (1973). Diversity and evenness: a unifying notation and its consequences. *Ecology* 54, 427–432.
- Hogue, E. W., & Miller, C. B. (1981). Effects of sediment microtopography on small-scale spatial distributions of meiobenthic nematodes. *Journal of Experimental Marine Biology and Ecology* 53, 181–191.

- Jensen, P. (1981). Species distribution and a microhabitat theory for marine mud dwelling Comesomatidae (Nematoda) in European waters. *Hydrobiologia* 108, 201–217.
- Jensen, P. (1987). Differences in microhabitat, abundance, biomass and body size between oxybiotic and thiobiotic free-living marine nematodes. *Oecologia* 71, 564–567.
- Jensen, P., & Aagaard, I. (1992). 'Bubbling reefs' in the Kattegat: submarine landscapes of carbonate-cemented rocks support a diverse ecosystem at methane seeps. *Marine Ecology Progress Series* 83, 103–112.
- Joint, I. R., Gee, J. M., & Warwick, R. M. (1982). Determination of fine-scale vertical distribution of microbes and meiofauna in an intertidal sediment. *Marine Biology* 72, 157–164.
- Kromkamp, J., Barranguet, C., & Peene, J. (1998). Determination of microphytobenthos PSII quantum efficiency and photosynthetic activity by means of variable chlorophyll fluorescence. *Marine Ecology Progress Series* 162, 45–55.
- Li, J., Vincx, M., Herman, P. M. J., & Heip, C. (1997). Monitoring meiobenthos using cm-, m- and km-scales in the Southern Bight of the North Sea. *Marine Environmental Research* 43, 265–278.
- Lucas, C., & Holligan, P. M. (1999). Nature and ecological implications of algal pigment diversity on the Molenplaat tidal flat Westerschelde estuary. *Marine Ecology Progress Series* 180, 51–64.
- Lucas, C. H., Widdows, J., Brinsley, M. D., Salkeld, P. N., & Herman, P. M. J. (2000). Benthic-pelagic exchange of microalgae at a tidal flat. I. Pigment analysis. *Marine Ecology Progress Series* 196, 59–73.
- McCune, B., & Mefford, M. J. (1999). *Multivariate analysis of ecological data*. Glendened Beach, OR: MjM Software.
- Moens, T., Herman, P. M. J., Verbeeck, L., Steyaert, M., & Vincx, M. (2000). Predation rates and prey selectivity of two predacious marine nematode species. *Marine Ecology Progress Series* 205, 185–193.
- Moens, T., Van Gansbeke, D., & Vincx, M. (1999). Linking estuarine nematodes to their suspected food. A case study from the Westerschelde Estuary (south-west Netherlands). *Journal of the Marine Biology Association of the U.K.* 79, 1017–1027.
- Neilson, R., & Boag, B. (2002). Marine nematode associations from an intertidal estuarine biotope. *Russian Journal of Nematology* 10, 113–121.
- Ólafsson, E., Elmgren, R., & Papakosta, O. (1993). Effects of the deposit-feeding benthic bivalve *Macoma balthica* on meiobenthos. *Oecologia* 93, 457–462.
- Ott, J. A. (1972). Determination of fauna boundaries of nematodes in an intertidal sand flat. *Internationale Revue der gesamten Hydrobiologie* 57, 645–663.
- Pickney, J., & Sandulli, R. (1990). Spatial autocorrelation analysis of meiofaunal and microalgal populations on an intertidal sandflat: scale linkage between consumers and resources. *Estuarine, Coastal and Shelf Science* 30, 341–353.
- Platt, H. M. (1977). Vertical and horizontal distribution of free-living marine nematodes from Strangford Lough, Northern Ireland. *Cahiers de Biologie Marine de Roscoff* 18, 261–273.
- Platt, H. M., & Lambshead, P. J. D. (1985). Neutral model analysis of patterns of marine benthic species diversity. *Marine Ecology Progress Series* 24, 75–81.
- Reise, K. (1983). Biotic enrichment of intertidal sediments by experimental aggregates of the deposit-feeding bivalve *Macoma balthica*. *Marine Ecology Progress Series* 12, 229–236.
- Sandulli, R., & Pickney, J. (1999). Patch sizes and spatial patterns of meiobenthic copepods and benthic microalgae in sandy sediments: a microscale approach. *Journal of Sea Research* 41, 179–187.
- Santos, P. J. P., Castel, J., & Souza-Santos, L. P. (1996). Seasonal variability of meiofaunal abundance in the oligo-mesohaline area of the Gironde Estuary, France. *Estuarine, Coastal and Shelf Science* 43, 549–563.
- Soetaert, K., & Heip, C. (1995). Nematode assemblages of deep-sea and shelf break sites in the North Atlantic and Mediterranean Sea. *Marine Ecology Progress Series* 125, 171–183.
- Soetaert, K., Vincx, M., Wittoeck, J., & Tulkens, M. (1995). Meiobenthic distribution and nematode community structure in 5 European estuaries. *Hydrobiologia* 311, 185–206.
- Soetaert, K., Vincx, M., Wittoeck, J., Tulkens, M., & Van Gansbeke, D. (1994). Spatial patterns of Westerschelde meiobenthos. *Estuarine, Coastal and Shelf Science* 39, 367–388.
- Steyaert, M., Garner, N., Van Gansbeke, D., & Vincx, M. (1999). Nematode communities from the North Sea: environmental controls on species diversity and vertical distribution within the sediment. *Journal of the Marine Biology Association of the U.K.* 79, 253–264.
- Steyaert, M., Herman, P. M. J., Moens, T., & Vincx, M. (2001). Variation in depth distribution of a nematode community during a tidal cycle (Molenplaat, Westerschelde estuary, The Netherlands). *Marine Ecology Progress Series* 224, 229–304.
- Tita, G., Desrosiers, G., Vincx, M., & Clement, M. (2002). Intertidal meiofauna of the St Lawrence estuary (Quebec, Canada): diversity, biomass and feeding structure of nematode assemblages. *Journal of the Marine Biology Association of the U.K.* 82, 779–791.
- Van de Koppel, J., Herman, P. M. J., Thoolen, P., & Heip, C. H. R. (2001). Do alternate stable states occur in natural ecosystems? Evidence from a tidal flat. *Ecology* 82, 3449–3461.
- Van Es, F. B., Van Arkel, M. A., Bouwman, L. A., & Schroder, H. G. J. (1980). Influence of organic pollution on bacterial, macrobenthic and meiobenthic populations in intertidal flats of the Dollard. *Netherlands Journal of Sea Research* 14, 288–304.
- Warwick, R. M., & Clarke, K. R. (1993). Increased variability as a symptom of stress in marine communities. *Journal of Experimental Marine Biology and Ecology* 172, 215–226.
- Warwick, R. M., & Gee, J. M. (1984). Community structure of estuarine meiobenthos. *Marine Ecology Progress Series* 18, 97–111.
- Warwick, R. M., McEvoy, A. J., & Trush, S. F. (1997). The influence of *Atrina zelandica* Gray on meiobenthic nematode diversity and community structure. *Journal of Experimental Marine Biology and Ecology* 214, 231–247.
- Warwick, R. M., & Price, R. (1979). Ecological and metabolic studies on free-living nematodes from an estuarine mudflat. *Estuarine and Coastal Marine Sciences* 9, 257–271.
- Wetzel, M. A., Jensen, P., & Giere, O. (1995). Oxygen/sulfide regime and nematode fauna associated with *Arenicola marina* burrows: new insights in the thiobios case. *Marine Biology* 124, 301–312.
- Widdows, J., Brinsley, M. D., Salkeld, P. N., & Lucas, C. H. (2000). Influence of biota on spatial and temporal variation in sediment erodability and material flux on a tidal flat (Westerschelde, The Netherlands). *Marine Ecology Progress Series* 194, 23–37.
- Wiesser, W. (1953). Die Beziehung zwischen Mundhöhlengestalt, Ernährungsweise und Vorkommen bei freilebenden Nematoden. *Arkiv für Zoology* 4(ser. 2), 439–484.