

Assessment of the soft sediment fauna five years after construction of the Princess Amalia wind farm



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CONTENTS

Figures	5
Tables	7
Summary	9
Samenvatting	11
1. Introduction	13
1.1 Princess Amalia Wind Farm	13
1.2 Scope of the study	14
2. Material and Methods	15
2.1 Survey design	15
2.2 Sampling methods	17
2.2.1 Boxcore sampling	17
2.2.2 Triple D Dredge sampling	19
2.3 Laboratory methods	20
2.3.1 Sediment analyses	20
2.3.2 Boxcore samples	21
Sorting	21
Identification	21
Biomass	21
2.3.3 Dredge samples	22
Sorting, identification and biomass	22
Shell damage	22
2.4 Statistical analyses	22
2.5 Quality control	23
3. Results	25
3.1 Physical variables	25
3.2 Boxcore samples	27
General diversity – species composition	27
Density and biomass	31
Multivariate analyses	32
3.3 Dredge samples	35
General diversity – species composition	35

Density and biomass	37
Multivariate analyses.....	39
Shell damage.....	41
4. Discussion.....	43
4.1 Development benthic invertebrate fauna Princess Amalia wind farm.....	43
4.2 Shell damage as indicator for reduced fishery pressure.....	44
4.3 Comparison with other wind farms in the Southern North Sea.....	44
4.4 Remarkable findings	46
5. Conclusions	49
6. Bibliography	51
7. Annex List	53
Annex 1: Species list.....	53
Annex 2: Sampling coordinates	53
Annex 3: Raw data boxcore	53
Annex 4: Raw data dredge.....	53
Annex 5: Physical variables	53

Figures

Figure 1: Projected sampling sites in the Dutch Coastal zone.....	16
Figure 2: Sample positions of the boxcores in the four areas.....	18
Figure 3: Sample positions of the dredges in the four areas.	20
Figure 4: Box-whisker plots of the sampling depth for boxcore samples per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.	25
Figure 5: Box-whisker plots of the median grain size per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.	26
Figure 6: Box-whisker plot of the organic matter content per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.	27
Figure 7: Box-whisker plot of species richness per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.....	28
Figure 8: Proportion of species richness of the taxa present in the boxcore samples. Percentages represent the relative contribution of the different taxa to the species richness.	28
Figure 9: Box-whisker plot of Pielou's Evenness per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.	29
Figure 10: Box-whisker plot of Shannon Index per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.....	29
Figure 11: Box-whisker plot of Gini-Simpson Index per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.	30
Figure 12: Estimated number of species in relation to the number of samples, determined with Bootstrapping.	30
Figure 13: Box-whisker plot of abundance per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.....	31
Figure 14: Box-whisker plot of biomass per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.	32
Figure 15: MDS plot based on density data with indication of the different areas (note the stress factor of 0.29, which suggests a relatively low relevance of the sample clusters). Dark blue circle: QCS; Red circle: QT and QAW; Light blue circle: QCN.....	33
Figure 16: Box-whisker plot of species richness per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.....	35
Figure 17: Box-whisker plot of Pielou's Evenness per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.	36
Figure 18: Box-whisker plot of Shannon Index per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.....	36
Figure 19: Box-whisker plot of Gini-Simpson Index per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.	37

Figure 20: Dredge samples, estimated number of species in relation to the number of samples, determined with Bootstrapping. 37

Figure 21: Box-whisker plot of abundance per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters. 38

Figure 22: Box-whisker plot of biomass per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters. 39

Figure 23: Length-frequency distribution of *Echinocardium cordatum* per area (outliers excluded). 39

Figure 24: MDS plot based on density data in the dredge samples with indication of the different areas (note the stress factor of 0.25, which suggest a relatively low relevance of the sample clusters). 40

Figure 25: Length-frequency distribution of *Chamelea* spp. in the different areas. 42

Figure 26: Length-frequency distribution of *Donax vittatus* in the different areas. 42

Figure 27: Length-frequency distribution of *Spisula* spp. in the different areas. 42

Figure 28: A) Polyp of *Eucheilota maculata*; B) siphon tip of *Lutraria* sp.; C) eggs of *Lunatia catena*. 47

Tables

Table 1: Sampling depth of the boxcore samples.....	25
Table 2: Physical variables of the boxcore samples.....	26
Table 3: Biodiversity data of the boxcore samples per area.....	27
Table 4: Density and biomass data of the boxcore samples.....	31
Table 5: Results ANOSIM and SIMPER analyses of the boxcore samples.....	34
Table 6: Taxa specific to one or two areas, which appear in more than one sample.....	34
Table 7: Biodiversity data of the dredge samples per area.....	35
Table 8: Density and biomass data of the dredge sampling.....	38
Table 9: Results ANOSIM and SIMPER analyses of the dredge samples.....	40
Table 10: Dredge sample taxa specific to one or two areas and appearing in more than one sample.....	41
Table 11: Number of damaged shells showing shell repair and total number of encountered individuals for the frequently encountered Bivalvia per area.....	41

SUMMARY

During the present study, the macrobenthic fauna was investigated five years after the construction of the Princess Amalia Wind Farm. In March 2012, samples were taken with a boxcorer and a dredge in the turbine site (QT), an adjacent buffer zone (QAW), a northern (QCN) and a southern reference site (QCS). These sites were also sampled in 2003 during the baseline study. Samples were taken at least 150 m away from the turbines.

Sediment characteristics of the turbine site were intermediate between the northern and the southern reference area. Density and diversity of the benthic fauna in the boxcore samples of the turbine site were also intermediate between the northern and the southern reference site, whereas biomass values did not differ significantly between areas.

Density, diversity and biomass in the dredge samples, which contain the larger K-strategic species, also did not differ between areas. In addition, no systematic differences in shell damage and length-frequency distribution of the common bivalve species could be detected between the areas. It can therefore be concluded that the installation of the Princess Amalia Wind Farm did not yet affect the soft sediment benthic fauna, at least not for samples taken further than 150 m from the turbines.

SAMENVATTING

In voorliggend onderzoek werd de macrobenthosgemeenschap in het Prinses Amaliawindmolenpark onderzocht vijf jaar na de constructie van het park. In maart 2012 werden met een boxcore en bodemshaaf monsters genomen in het windmolenpark (QT), in een bufferzone rond het windmolenpark (QAW) en in een Noordelijk (QCN) en Zuidelijk (QCS) referentiegebied. Deze gebieden kwamen overeen met de monsternamenpunten uit het baseline onderzoek uit 2003.

De sedimenteigenschappen in het windmolenpark vertoonden een internmediair beeld tussen het Noordelijk en Zuidelijk referentiegebied. Hetzelfde patroon werd gevonden voor de abundantie en diversiteit van macrofauna, verzameld met de boxcore; biomassa was gelijkaardig over alle bemonsterde gebieden.

Densiteit, biodiversiteit en biomassa van de fauna verzameld met de bodemshaaf, een apparaat wat eerder geschikt is voor het bemonsteren van soorten met een K-strategie, was niet verschillend tussen alle bemonsterde gebieden. Daarnaast werden geen significante verschillen gevonden in beschadigingen aan schelpen – een indicatie voor visserijdruk – of lengte-frequentie distributie van de meest voorkomende tweekleppigen.

Uit dit onderzoek wordt geconcludeerd dat de bouw van het Prinses Amaliawindmolenpark, en de daarbij horende maatregelen zoals het verbod op visserij, nog geen effect heeft gehad op de bodemfauna in het gebied, uitgaande van monsters genomen op minstens 150 m afstand van de windmolens.

1. INTRODUCTION

As part of the Monitoring- and Evaluation Program (MEP) of the Princess Amalia Wind Farm (PAWP), a number of research topics were identified. One of those topics was to characterise the benthic invertebrates living in the seabed. In 2003, a baseline study of the subtidal benthic fauna was carried out by the Institute of Estuarine and Coastal Studies (University of Hull, UK). Samples were taken in the proposed Princess Amalia Wind Farm turbine site (QT), an adjacent buffer zone (QAW) and two reference sites situated north (QCN) and south (QCS) of the site. Results from this baseline study were reported in February 2004 (Jarvis *et al.*, 2004).

Five years after the construction of the Princess Amalia Wind Farm (2012), a new monitoring study was carried out in the same four areas (QT, QAW, QCN and QCS). The object of this report is twofold:

- to present the results of the benthic infauna monitoring;
- to contrast the new findings with the baseline condition prior to the construction of the wind farm.

1.1 Princess Amalia Wind Farm

The Princess Amalia Wind Farm is the second offshore wind farm in the Dutch sector of the North Sea and the first to be located outside the 12 nautical mile limit. PAWP is located in Block Q7 of the Dutch Continental Shelf, at a distance of 23 to 26.4 km from the shore (off the coast of IJmuiden, The Netherlands), in water depths between 19 and 24 m. A total of 60 turbines (4.0 m diameter monopole foundations) were placed in an area of 17 km². The minimum distance between turbines is 550 m. The Princess Amalia Wind Farm has a designed power output of 435 GWh per annum.

The installation of the foundations and transition pieces took place from October 2006 till May 2007. Foundations measuring 54 metres, a diameter of 4 metres and 320 tons in weight were sunk into the sea-floor. The transition pieces weighing 115-tons were placed on the foundations using the Jumping Jack. To support the turbine foundations, a 15 m diameter scour-protection consisting of mixed size rocks was deposited on the soft sediment around the base of each monopile. Cables and wind turbines were installed from May 2007 to April 2008. The wind farm is operational since June 2008.

1.2 Scope of the study

The aim of this study is to evaluate the eventual impacts arising from the construction of the Princess Amalia Wind Farm. Specifically, the effects it may have on occurrence and density of the invertebrates living on the seabed. Comparisons are made between the baseline study and the current situation with respect to infaunal assemblage, epifaunal communities, faunal characteristics, evidence of anthropogenic effects (i.e. shell damage and shorter shells due to trawling) and sediment types. By comparing the results obtained in the baseline study with the results of the present study and by comparing the data from the turbine site with the reference areas, we intend to assess whether the installation of the wind farm caused effects on the soft sediment benthos. A second related objective is to assess whether the wind farm provides shelter for certain longer-lived species like large bivalves and sea urchins. Also shell damage and length-frequency distribution of the bivalves are assessed as it can be expected that less shells with scars and larger individuals can be found when the disturbance caused by trawling decreases (Witbaard & Klein, 1994).

2. MATERIAL AND METHODS

2.1 Survey design

For the present study, the same sampling strategy was chosen as used in the baseline study (Jarvis *et al.*, 2004), to facilitate comparison of the data. The baseline design identified reference areas similar in sediment characteristics to those present under the footprint of the wind farm. Sediment texture (i.e. median particle size) was thought to be one of the main structuring factors of benthic fauna distribution (Gray & Elliot, 2009).

Two reference areas were used, one 40 km to the north of the turbine site (QCN) and another 30 km to the south (QCS). Both reference areas have the same distance to the shore as the turbine site. In addition, three pairs of sampling sites were located around the windfarm area (QAW), to reveal either a possible impact of increased fishing activity on the margins of the exclusion zone or a positive spill-over effect from inside the windfarm area. In the turbine site, samples were always taken at least 150 m away from the monopoles.

The projected sampling sites are indicated in Figure 1.

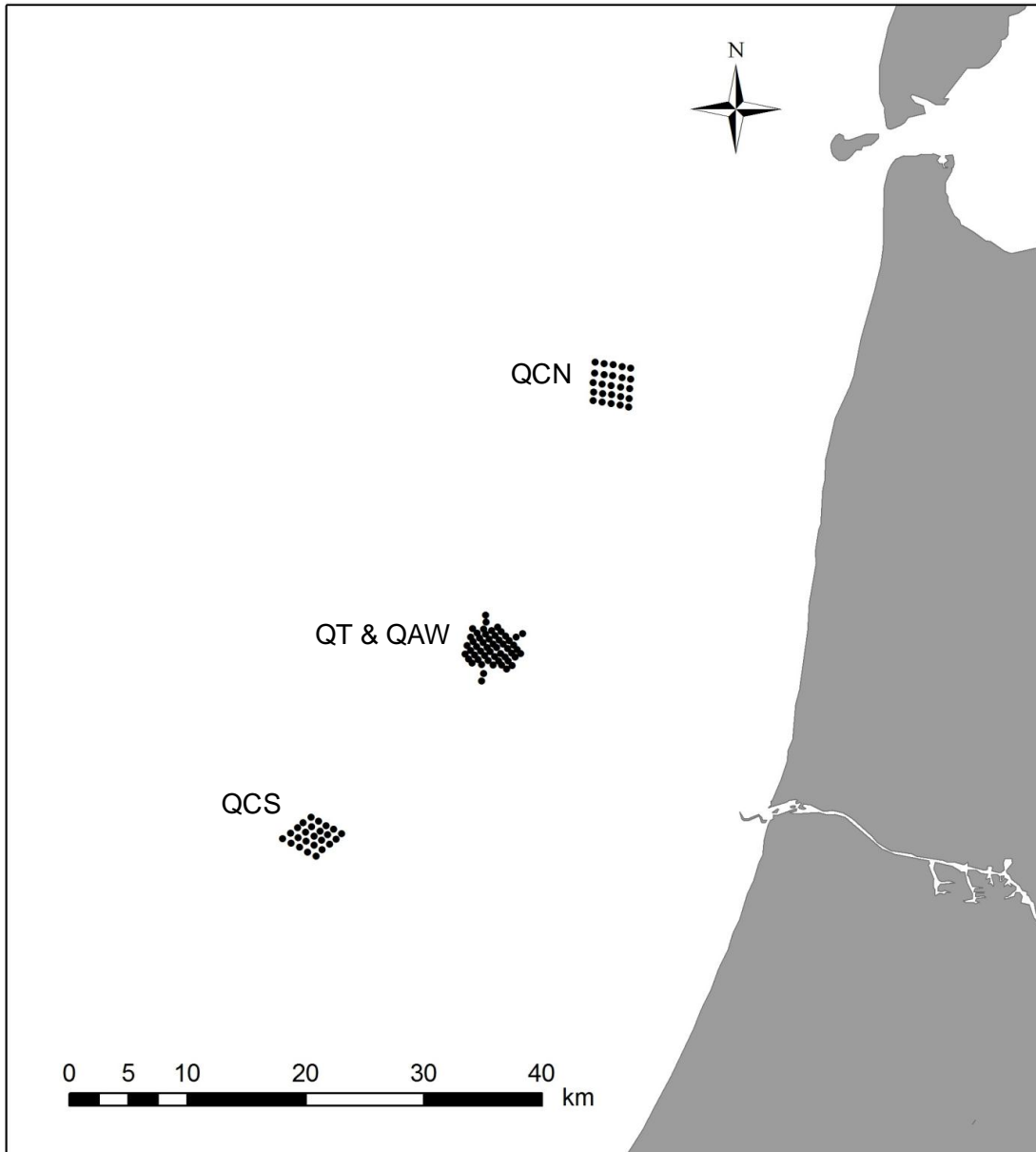


Figure 1: Projected sampling sites in the Dutch Coastal zone.

2.2 Sampling methods

2.2.1 Boxcore sampling

For the survey of the small infaunal invertebrates, the Reineck box with a circular corer (0.078m² sampling surface)¹ was chosen. A total of 199 samples of the projected 200 samples (see further) were collected at 103 stations (79 non-replicated stations and 24 replicated stations) from the turbine site, the adjacent buffer zone and the two reference (control) sites.

The overall sampling effort for the monitoring surveys was as follows (Figure 2):

Non-replicated stations

- Turbine site: 39 stations (39 samples)
- Northern reference area: 20 stations (20 samples)
- Southern reference area: 20 stations (20 samples)

Replicated stations:

- Turbine site: 8 stations (40 samples)
- Adjacent waters: 6 stations (30 samples)
- Northern reference area: 5 stations (25 samples)
- Southern reference area: 5 stations (25 samples)

At every pre-determined sampling station, the Boxcore was lowered and a sample was taken. Upon retrieval of the sampler, the contents of the box corer's sample holder were inspected by a qualified surveyor and the quality of the sample was assessed. Samples disturbed or too low in volume were rejected. Valid samples were washed over a 1 mm sieve. The residue was photographed for possible future reference and stored in a HdPE jar with suitable fixing and preserving solution (4% formo-saline solution).

Boxcore samples were taken during one week, from the 12th of March onwards. Samples were taken in a north to south direction. Only one station (QT17) was skipped, because it was located only 10 m from the cable to the OHVS.

Taking replicate samples turned out to be difficult due to tidal currents, which made it impossible to stay at the exact same location while collecting the replicates. However, thanks to the favourable weather conditions experienced during the survey, all replicate samples could be collected after the non-replicate samples; the extra sampling day planned to collect all possibly remaining replicate samples was hence not used.

¹ The Reineck box corer consists of a round metal box (29,5 cm diameter), which is attached to a weighted tubular steel frame. Upon deployment and when contact is made with the seabed, the projecting box is driven into the sediment by the weight of the corer. Once the tension has been taken by the lifting cable, a mechanism rotates a counterbalanced arm that swings a cutting plate through the sediment and under the box to retain the sediment contained within. The sampling device is then lifted from the seabed and through the water column to the survey vessel. The cutting plate has a rubberised upper surface, which seals the sediment sample within the box corer by sitting flush against the lower edge of the corer during the recovery from the seabed to the vessel.

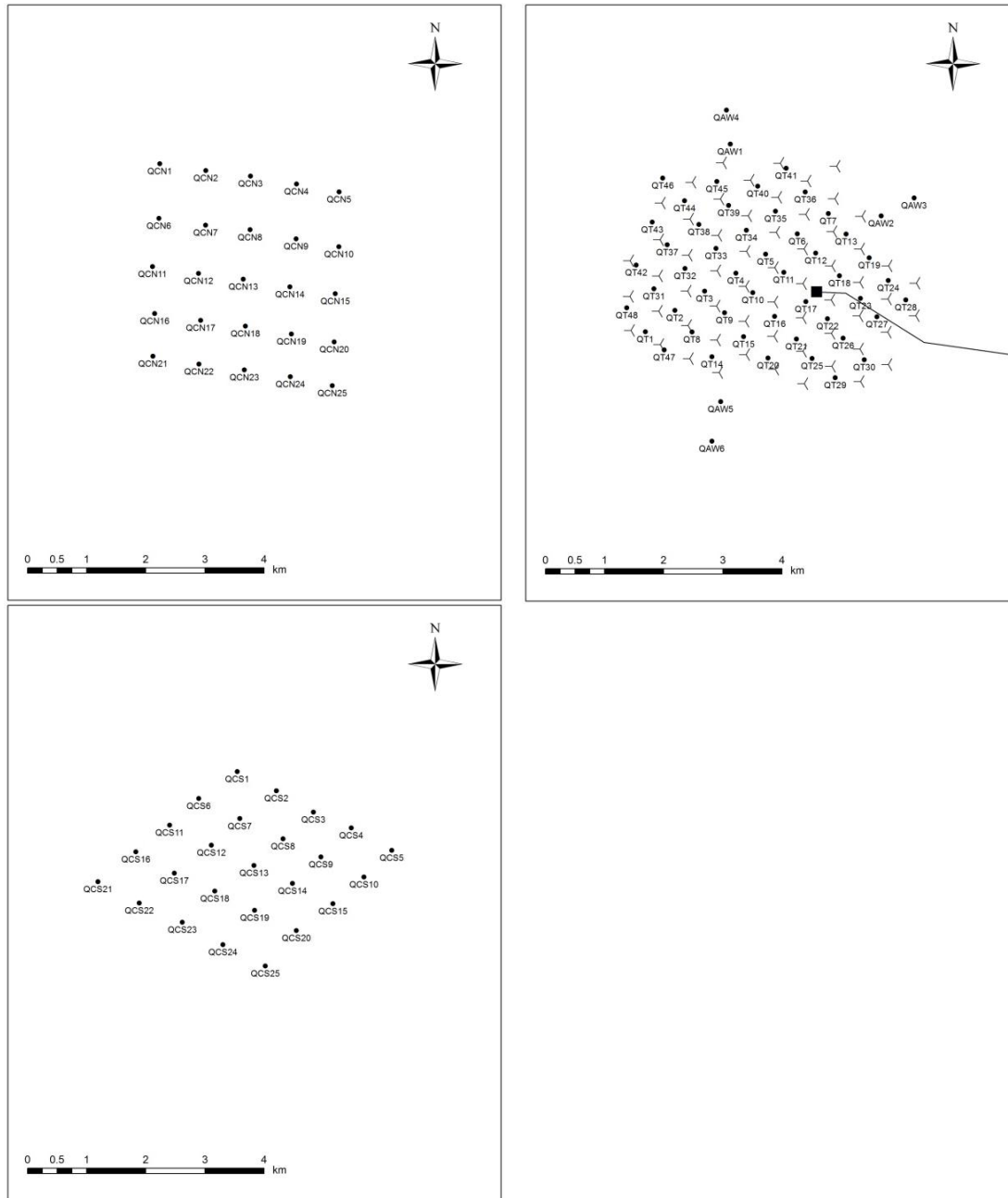


Figure 2: Sample positions of the boxcores in the four areas.

2.2.2 Triple D Dredge sampling

To survey less abundant and larger infaunal invertebrates, the Triple D dredge was chosen. This dredge has a cutting plate with a width of 1 m and a penetration depth of 15 cm, the retaining net has a mesh size of 6 mm. The dredge was towed for a distance of 100 m, which was the optimal sample size. Trial tows of 50 m proved to be too short and trial tows of 200 m were longer than necessary. A total of 39 samples were collected at 39 different stations.

The overall sampling effort for the monitoring surveys was as follows (Figure 3):

- Turbine site: 15 dredge tows
- Adjacent waters: 6 dredge tows
- Northern reference area: 9 dredge tows
- Southern reference area: 9 dredge tows

At the pre-determined sampling stations, the dredge was lowered and towed over a distance of 100 m through the seabed. After 100 m, the dredge was hauled in. Once on deck, the retaining net was emptied and, if approved to be a successful sample, the content of the net was placed in a sample container and photographed for possible future reference. All dredge samples were sorted onboard, identified to species level if possible and measured. Depending on the total volume of the sample, a subsample was taken. The processed part of the sample was fixated with a 4% formalin-saline solution.

Dredge samples were taken from the 25th till 28th of March in the order QCN, QT, QCS and QAW. Samples were temporarily stored on ice in labeled, closed buckets till processing on land. At the end of each sampling day, samples were brought to land to be processed by the processing crew the next day.

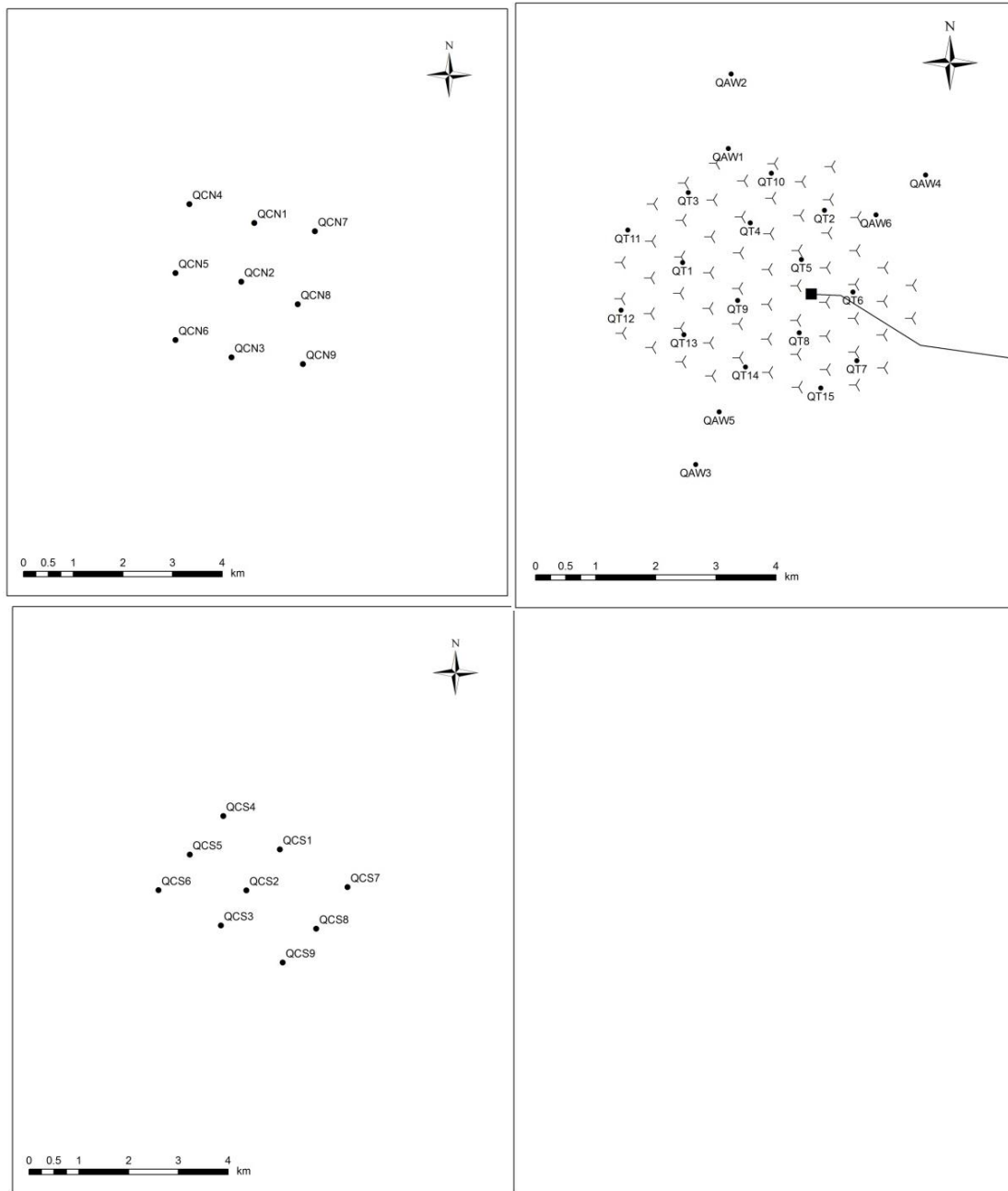


Figure 3: Sample positions of the dredges in the four areas.

2.3 Laboratory methods

2.3.1 Sediment analyses

Sediment particle size of the fraction less than 2000 μm was determined with laser granulometry (Malvern Mastersize 2000 laser granulometer).

To assess the organic matter content, the PrepAsh was used: an instrument for gravimetric analysis, which increases precision while decreasing the time of analysis.

This is mainly accomplished by the completely automatised run of 29 samples and one reference sample. The weight change of the sample is plotted while the temperature gradually increases according a predefined program.

2.3.2 Boxcore samples

In accordance with the research plan, only the non-replicate boxcore samples were sorted in the lab. The replicate samples were collected for possible further reference and would only be analysed if the data from the non-replicated sites suggested there might be patterns at a small spatial scale that were relevant for the research questions.

Sorting

After fixation with a 4% formo-saline solution, the benthos samples were stained with Rose Bengal (1%), which has the ability to stain proteins in animals that were caught alive. After 24h, the organisms were picked out.

The samples were thoroughly rinsed in a 1 mm sieve to remove as much fixation liquid as possible. The rinsed sediment was stored in a triage bin, where the organisms were picked out. Under a binocular microscope, the organisms were sorted to higher taxa levels. From this point, the organisms were preserved in 70% ethanol. The residue was kept for possible future audits.

Identification

All present organisms were identified to species level whenever possible and counted. Identification was performed with a binocular microscope and based on the most recent systematic literature. For nomenclature and taxonomy, the World Register of Marine Species (WoRMS) was followed. All identified organisms were stored per species in separated tubes/jars.

Biomass

In this study, we opted to determine biomass (ash free dry weight) in an indirect way. Depending on the taxonomic class, a different method was used. This is according to the methods used for the T0 data (Jarvis *et al.*, 2004) and the T1 data of OWEZ (Daan *et al.*, 2007).

Molluscs, echinoids and *Ophiura* spp.

Lengths and widths (if possible) were measured of molluscs and echinoids, respectively. The central disk was measured of *Ophiura* spp. All measurements were done with a digital caliper with an accuracy of 0.01 mm. Mollusc lengths were converted to ashfree dry weight (AFDW) using an equation of the form: $W = a.L^b$; where W is AFDW, L is length and a and b are species specific constants supplied by NIOZ.

Polychaetes, larger crustaceans, gastropods and *Asterias rubens*

Biomass data of these organisms were determined with the Blotted wet weight method. Organisms were weighted per sample and per species. Individuals were dried on filter paper until no surface moisture was present. The organisms were gently placed in an aluminium cup (small organisms) or a porcelain cup (larger organisms) and weighted to 0.0001 gram accurate. Wet weight was transformed to ashfree dry weight (AFDW) by using applicable conversion factors for the different taxa (Ricciardi & Bourget, 1998).

2.3.3 Dredge samples

Sorting, identification and biomass

All 39 dredge samples were sorted, identified and weighted (wet weight) in a field laboratory. Some species (polychaetes and bivalves) were re-identified in the laboratory if there was some uncertainty about the identification. In addition, five complete samples were re-sorted and re-identified in the laboratory for quality control purposes.

Shell damage

To assess whether bivalves suffered from shell damage, all species of bivalves that were regularly encountered were examined for scars under a dissecting microscope. In addition, shell lengths were measured to assess length-frequency distributions.

2.4 Statistical analyses

Density and biomass (Ash Free Dry Weight, AFDW) for the boxcore samples were standardised to the number of individuals per m^2 (ind/ m^2) and for the dredge samples to the number of individuals per 100 m^2 . In addition, diversity indices were calculated per sample (Shannon-Wiener, Pielou's Evenness and Gini-Simpson index). The Shannon index is based on the relative abundances of the species in a community and increases with increasing diversity. Pielou's evenness ranges from 0 to 1 and raises with decreasing variation in communities. The Gini-Simpson index equals the probability that two individuals represent different taxa, which raises with increasing diversity. Differences between the four different areas were assessed with Kruskal-Wallis ANOVA followed by post-hoc multiple comparisons using Statistica (StatSoft, 2004).

Multivariate analyses were carried out with the Primer v6 program (Clarke & Gorley, 2006). Before analysis, data were fourth-root transformed. Bray-Curtis similarity matrices were used to build up non-metric multidimensional scaling (MDS) plots. MDS plots give information on relationships between data points. SIMPER analysis allows to detect which species contribute to the distance between certain communities (dissimilarity percentage) and clustering in a community (similarity percentage). ANOSIM analyses (Analysis of Similarities) were performed to determine significant differences ($p < 0.05$) between groups (areas).

2.5 Quality control

During the analyses of benthic samples - sorting and identification - quality controls were carried out. 10% of the boxcore samples and 12% of the dredge samples were re-analysed by a colleague analyst.

All controls were within the 90% similarity level set as a minimum acceptance criterion by NMBAQCS (Worsfold & Hall, 2010).

3. RESULTS

3.1 Physical variables

Sampling depth of the boxcore samples ranged from 16.6 m (QT29) to 23.1 m (QCS14), with a mean of 21.0 ± 1.5 m for all samples (Table 1). Despite the small range, sampling depth was significantly shallower in the northern reference area than in the southern reference area and the turbine site (Figure 4).

Table 1: Sampling depth of the boxcore samples.

Area	Sampling depth (m)			
	Mean	St. Dev.	Max.	Min.
QCN	20,2	0,18	20,7	20,0
QT	21,1	1,87	22,5	16,6
QAW	20,8	1,03	22,2	20,0
QCS	21,7	1,19	23,1	19,1
Total	21,0	1,50	23,1	16,6

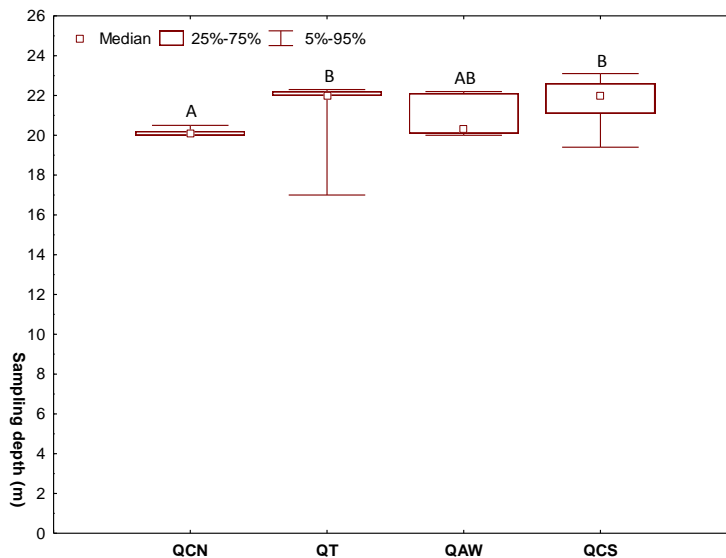


Figure 4: Box-whisker plots of the sampling depth for boxcore samples per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

Median grain size ranged from $210 \mu\text{m}$ (QCS 6) to $364 \mu\text{m}$ (QCN 16), with a mean particle size of $280 \pm 28 \mu\text{m}$ for all samples (Table 2); this can be classified as medium sand according to the Wentworth-scale. There was a slight north-south trend of

coarsening sands, with a significantly higher median grain size in the southern reference area than in the northern reference area and the turbine site (Figure 5).

Table 2: Physical variables of the boxcore samples.

Area	Median particle size (μm)		Clay (%)		Sand (%)		Organic matter (%)	
	Mean	St. Dev.	Mean	St. Dev.	Mean	St. Dev.	Mean	St. Dev.
QCN	263	31,4	1,40	2,22	98,6	2,22	0,57	0,16
QT	277	16,8	0,31	1,36	99,7	1,36	0,50	0,06
QAW	279	5,93	0,00	0,00	100	0,00	0,59	0,19
QCS	303	29,9	0,16	0,46	99,8	0,46	0,49	0,14

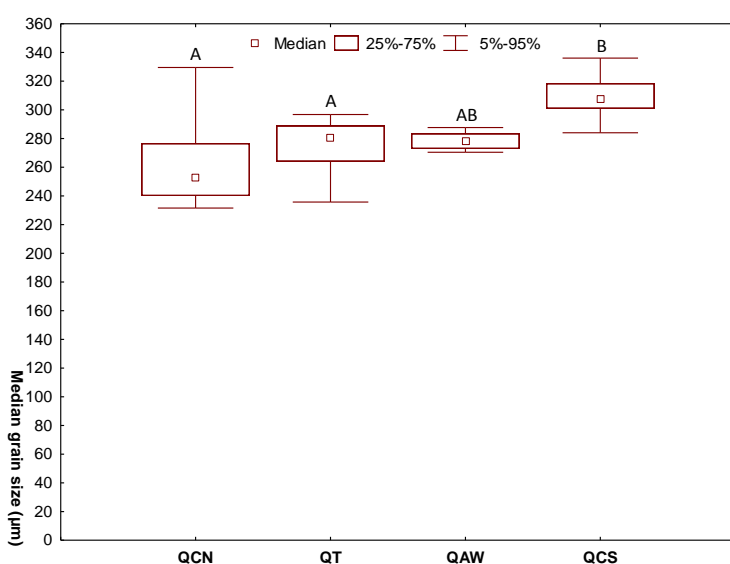


Figure 5: Box-whisker plots of the median grain size per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

Clay percentages (particle size less than $63 \mu\text{m}$) were very low, the mean ranged between 0% (QAW) and 1.4% (QCN). Sandy sediments dominated the three areas, with mean percentages varying between 98.6 and 100.0%. No significant differences in clay and sand percentages were observed between the areas.

Organic matter content was comparable between the different areas, with an overall mean of $0.521 \pm 0.125\%$. The lowest value was measured for QCN 19 with a value of 0.277%, the highest value (1.00%) was measured in QCN 25. Despite the limited range, organic matter content was significantly lower in the southern reference area than in the northern reference area (Figure 6).

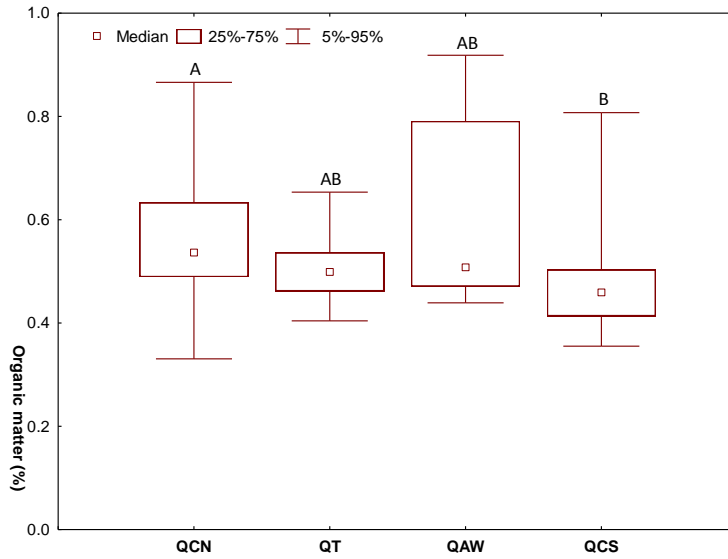


Figure 6: Box-whisker plot of the organic matter content per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

3.2 Boxcore samples

General diversity – species composition

A total number of 58 different invertebrate taxa was found in the windfarm area and the two reference areas. The number of species per sample varied between 3 (QCS 8) and 19 (QT 10) with an average of 10 ± 3 species per sample (Table 3). Species richness was significantly higher in the northern than in the southern reference area (Figure 7). Most of the taxa found were polychaete worms (21 taxa) and crustaceans (20 taxa); the latter contained amongst others 11 amphipod species and 5 decapod species (Figure 8).

Table 3: Biodiversity data of the boxcore samples per area.

Area	Samples	Number of species			Pielou's Evenness	Shannon Index	Gini-Simpson Index
		Total	Mean	St. Dev.			
QCN	25	40	12	3	0,866	2,09	0,834
QT	47	51	9	3	0,879	1,94	0,816
QAW	6	24	10	2	0,849	1,94	0,800
QCS	25	36	8	3	0,864	1,70	0,751
Total	103	58	10	3	0,870	1,92	0,804

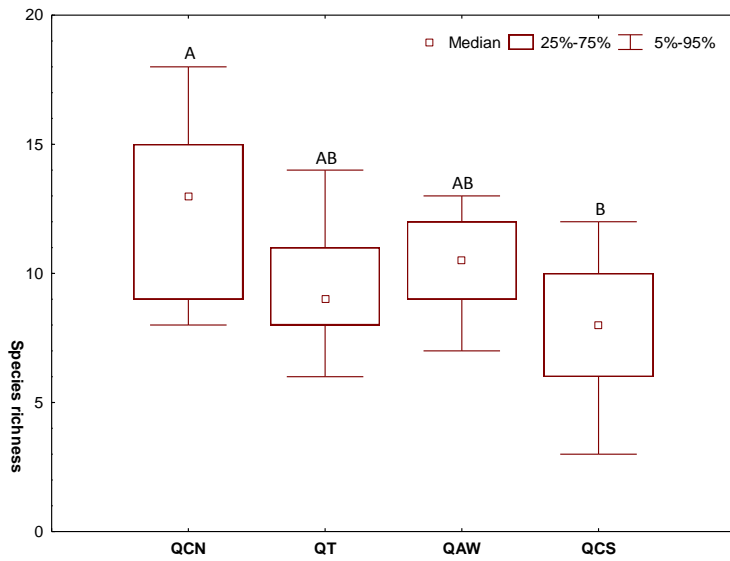


Figure 7: Box-whisker plot of species richness per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

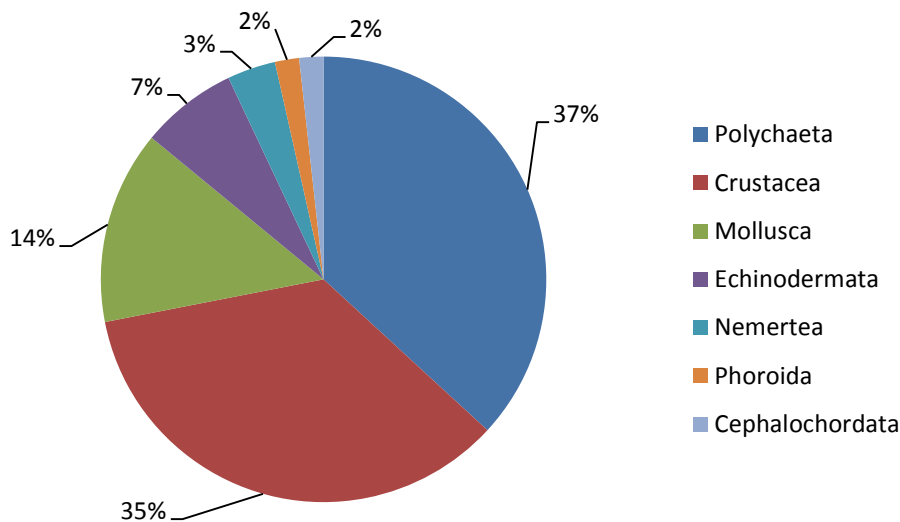


Figure 8: Proportion of species richness of the taxa present in the boxcore samples. Percentages represent the relative contribution of the different taxa the species richness.

Pielou's evenness was similar for the different areas (Figure 9). The Pielou's Evenness values of 0,84 and higher indicate a fairly even community in all areas. Shannon Index (Figure 10) and Gini-Simpson Index (Figure 11) were both significantly higher in the northern reference area than in the southern reference area.

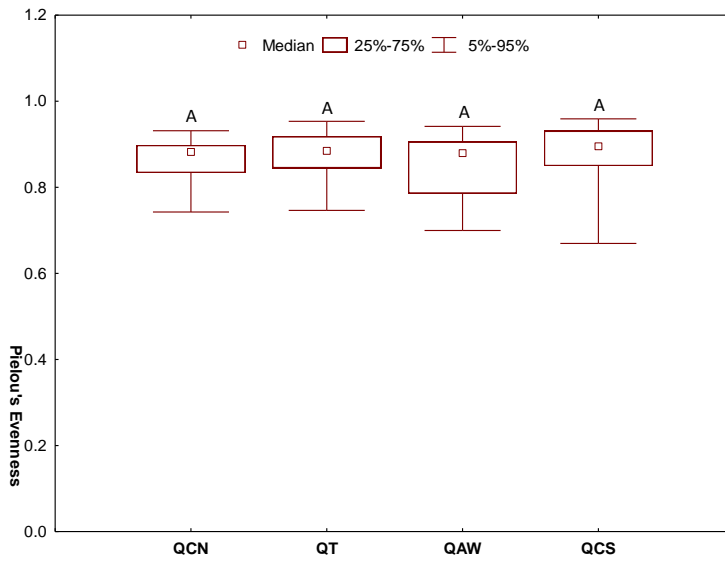


Figure 9: Box-whisker plot of Pielou's Evenness per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

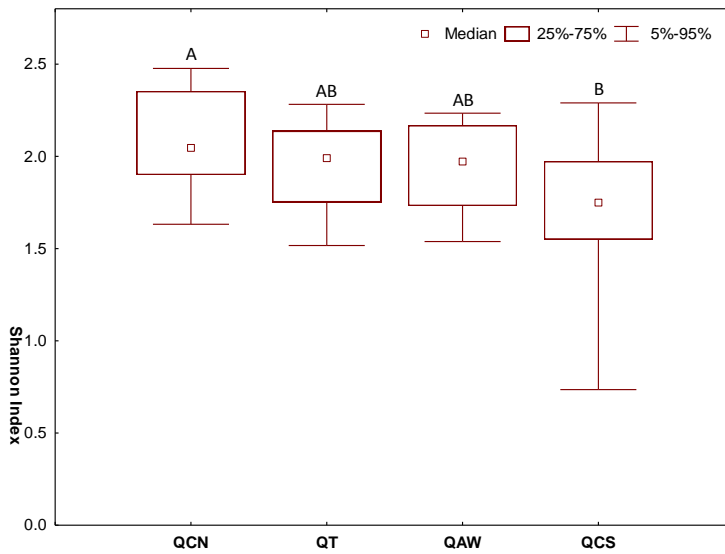


Figure 10: Box-whisker plot of Shannon Index per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

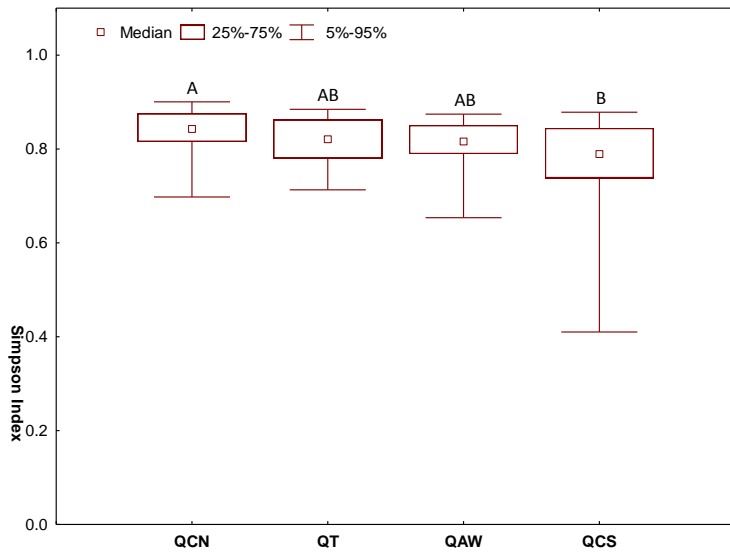


Figure 11: Box-whisker plot of Gini-Simpson Index per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

A ‘species accumulation curve’ was used to examine if the amount of analysed samples was sufficient to get a representative idea of the biodiversity present (Figure 12). The maximum number of species was estimated using an extrapolation technique (Bootstrapping). The graph shows that at 104 samples, the estimated number of species is close to an asymptotic value. This means that the sampling effort was sufficient. Looking at estimated versus observed number of species over all areas together, 95% of the estimated number of species was encountered in the samples. In the northern and southern reference site it was 90%, in the turbine site 86%.

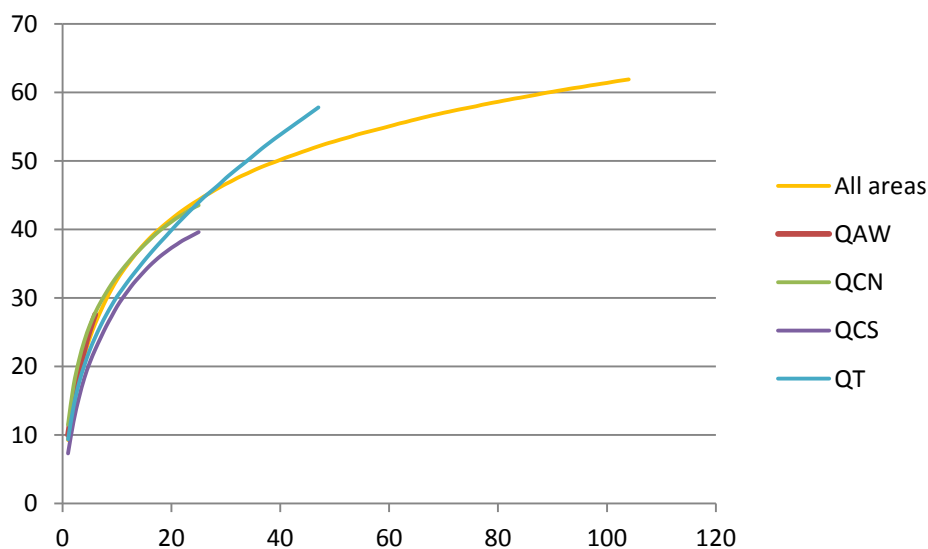


Figure 12: Estimated number of species in relation to the number of samples, determined with Bootstrapping.

Density and biomass

Mean invertebrate densities ranged from 186 ± 80 individuals/m² in QCS to 459 ± 258 individuals/m² in QCN (Table 4). The maximum density was observed in QCN 12 with 1154 individuals/m², which was mainly due to a high density of the tube building polychaete *Lanice conchilega* (423 individuals/m²). A high density in QT 20 was due to *Scoloplos armiger* (295 individuals/m²). Density was significantly lower in the southern reference area than in the other areas and density was also significantly lower in the turbine site than in the northern reference area (Figure 13).

Table 4: Density and biomass data of the boxcore samples.

Area	Density (N/m ²)				Biomass (g AFDW/m ²)			
	Mean	St. Dev.	Max.	Min.	Mean	St. Dev.	Max.	Min.
QCN	459	258	1154	205	16,1	14,3	49,7	0,214
QT	280	132	756	128	7,43	7,98	39,5	0,101
QAW	346	152	577	218	9,50	8,11	18,7	0,626
QCS	186	80	385	77	9,92	12,0	53,9	0,225
Total	305	189	1154	77	10,3	11,2	53,9	0,102

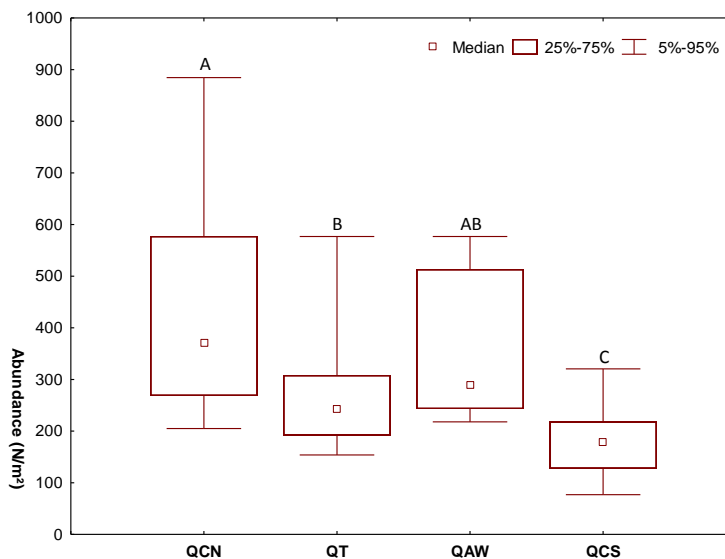


Figure 13: Box-whisker plot of abundance per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

The mean biomass (AFDW) was highest in the northern reference site (16.1 ± 14.3 g/m²) and lowest in the turbine site (7.43 ± 7.98 g/m²). High biomasses were mostly due to the presence of *Echinocardium cordatum* and adult *Ensis directus*. There were no statistically significant differences in biomasses between the areas (Figure 14).

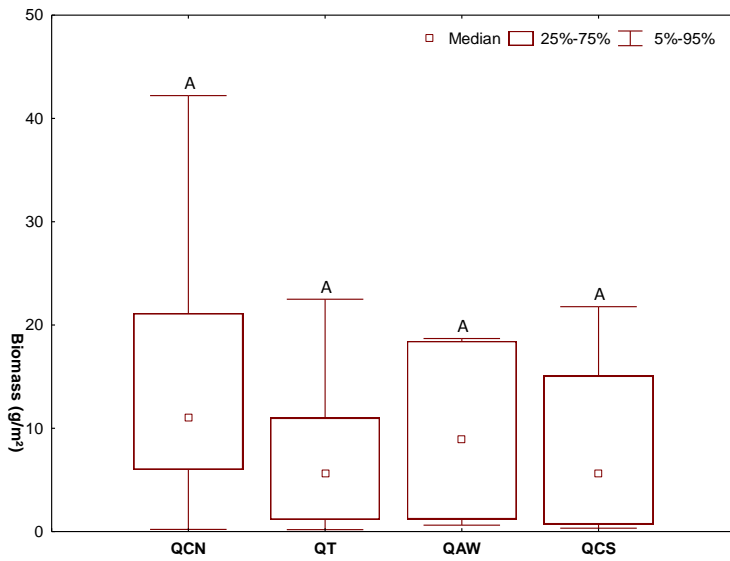


Figure 14: Box-whisker plot of biomass per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

Multivariate analyses

The multivariate ANOSIM data-analysis showed statistical differences in density and biomass between the different areas ($p < 0.001$). However, no difference was found between the QT area and any of the other areas. Looking at the MDS-plot, it is clear that the samples are arranged along a north – south gradient (Figure 15). Dissimilarity between areas was derived from the ANOSIM and SIMPER results. The dissimilarity between the different areas was caused by differences in densities of the following species: *Bathyporeia elegans*, *Chaetozone setosa* and *Echinocardium cordatum*. When these analyses were performed on the biomass data, similar dissimilarity percentages were obtained, although there was a shift in characteristic species. The biomass indicator species were *Echinocardium cordatum*, *Ensis directus* and *Lanice conchilega* (Table 5).

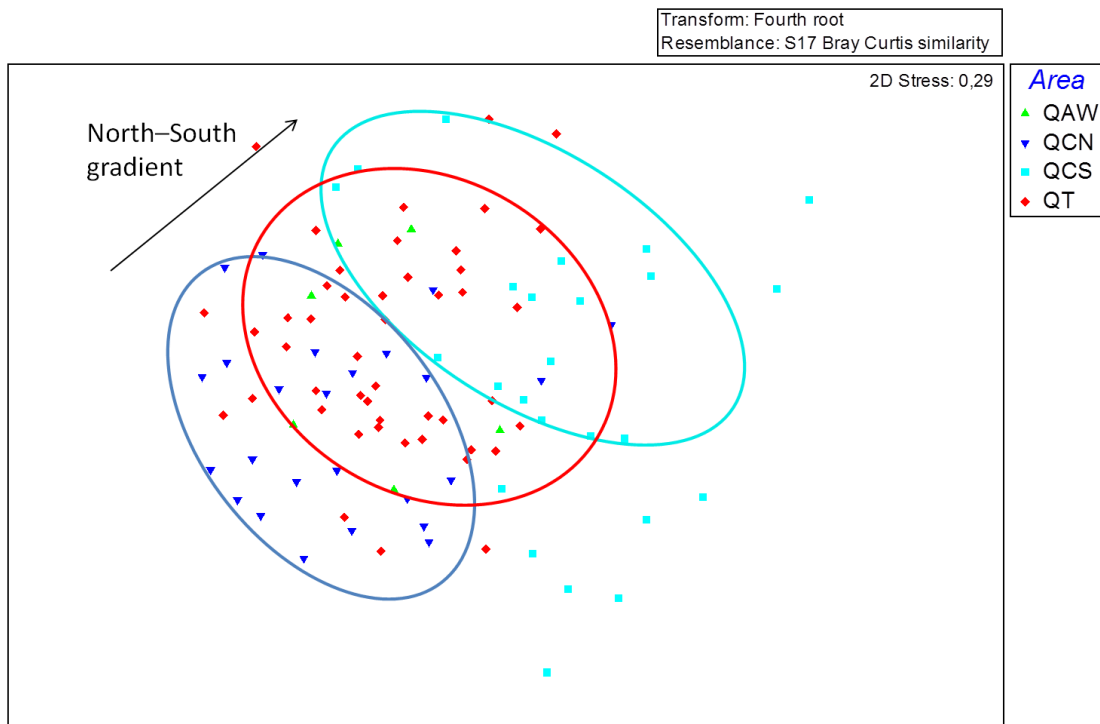


Figure 15: MDS plot based on density data with indication of the different areas (note the stress factor of 0.29, which suggest a relatively low relevance of the sample clusters). Dark blue circle: QCS; Red circle: QT and QAW; Light blue circle: QCN.

The most abundant taxa were *Scoloplos armiger*, *Nephtys cirrosa* and *Bathyporeia elegans*. These species led to a similarity of $\pm 40\%$ in each area². A total of 31 species (53%) occurred in all areas, which indicates that the seabed on a broad scale in this area is comparable. The taxa listed in Table 6 were specific to one or two areas (North-South gradient from left to right) and occurred in more than one sample.

² In multivariate analyses, the areas QT and QAW were treated as one area, because of the low sample size of QAW.

Table 5: Results ANOSIM and SIMPER analyses of the boxcore samples.

Area	<i>R</i>	<i>p</i>	Dissimilarity (%)	Indicator species	Mean density Area 1 – Area 2	
QCN - QCS	0,355	0,001	65,2	<i>Bathyporeia elegans</i>	2,29	0,96
				<i>Chaetozone setosa</i>	1,67	0,23
				<i>Bathyporeia guilliamsoniana</i>	1,98	0,77
				<i>Echinocardium cordatum</i>	1,48	1,16
				<i>Scoloplos armiger</i>	2,29	1,61
				<i>Opbelia borealis</i>	0,56	1,31
QCN - QT	0,239	0,001	57,0	<i>Bathyporeia guilliamsoniana</i>	1,98	0,85
				<i>Chaetozone setosa</i>	1,67	0,90
				<i>Urotroche brevicornis</i>	0,85	1,13
				<i>Echinocardium cordatum</i>	1,48	0,97
				<i>Magelona johnstoni</i>	1,14	0,59
				<i>Eteone longa</i>	0,66	1,16
				<i>Bathyporeia elegans</i>	2,29	2,17
QCS - QT	0,417	0,001	64,0	<i>Bathyporeia elegans</i>	0,96	2,17
				<i>Scoloplos armiger</i>	1,61	2,77
				<i>Opbelia borealis</i>	1,31	0,42
				<i>Echinocardium cordatum</i>	1,16	0,97
				<i>Eteone longa</i>	0,23	1,16
				<i>Urotroche brevicornis</i>	0,15	1,13

Table 6: Taxa specific to one or two areas, which appear in more than one sample.

QCN	QT and QAW	QCS
<i>Nephtys caeca</i>		
	<i>Chamelea striatula</i>	
	<i>Donax vittatus</i>	
	<i>Eunereis longissima</i>	
	<i>Lanice conchilega</i>	
	<i>Malmgreniella darbouxi</i>	
	<i>Nephtys hombergii</i>	
	<i>Phyllodoce mucosa</i>	
	<i>Processa modica</i>	
	<i>Asterias rubens</i>	
	<i>Nototropis falcatus</i>	
	<i>Pinnotheres pisum</i>	
	<i>Leucothoe incisa</i>	
	<i>Spiophanes bombyx</i>	
	<i>Spisula subtruncata</i>	
	<i>Thia scutellata</i>	
	<i>Aricidea spp.</i>	
	<i>Liocarcinus holsatus</i>	

3.3 Dredge samples

General diversity – species composition

During the benthic dredge surveys, a total of 60 different species were identified, with an average of 19 ± 3 species per sample (Table 7). Species richness was similar in the different areas (Figure 16). Eleven taxa were decapods, 11 bivalves, four echinoderms and 14 fish. Also 15 polychaete species were identified, however, this benthic dredge is not optimal to sample polychaetes, because only bigger specimens are retained in the net.

Table 7: Biodiversity data of the dredge samples per area.

Area	Samples	Number of species			Pielou's Evenness	Shannon Index	Gini-Simpson Index
		Total	Mean	St. Dev.			
QCN	9	41	19	3	0,731	2,16	0,823
QT	15	48	19	3	0,694	2,03	0,781
QAW	6	35	19	3	0,712	2,08	0,830
QCS	9	39	18	2	0,745	2,14	0,822
Total	39	60	19	3	0,717	2,09	0,808

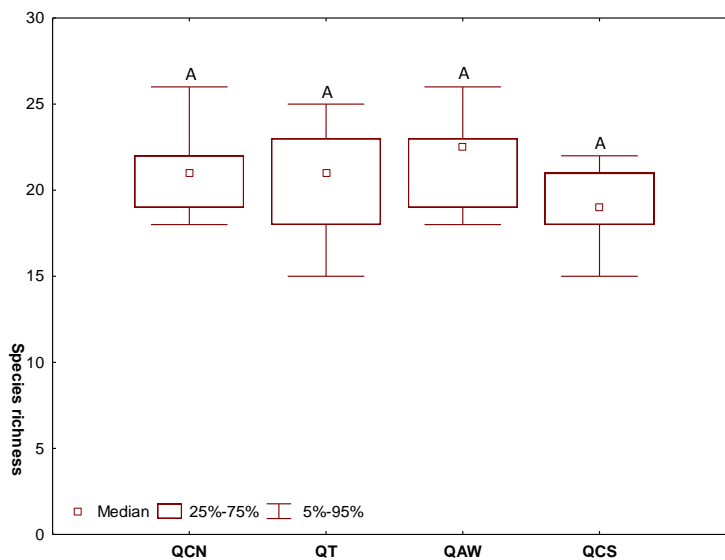


Figure 16: Box-whisker plot of species richness per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

There is little variation in diversity between the different sampling areas. There were no statistically significant differences in Pielou's Evenness (Figure 17), Shannon Index (Figure 18) and Gini-Simpson Index (Figure 19) between the areas.

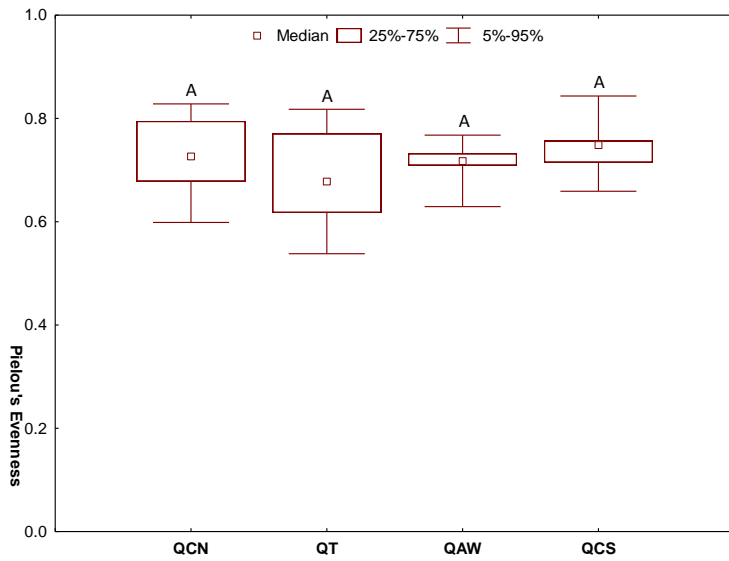


Figure 17: Box-whisker plot of Pielou's Evenness per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

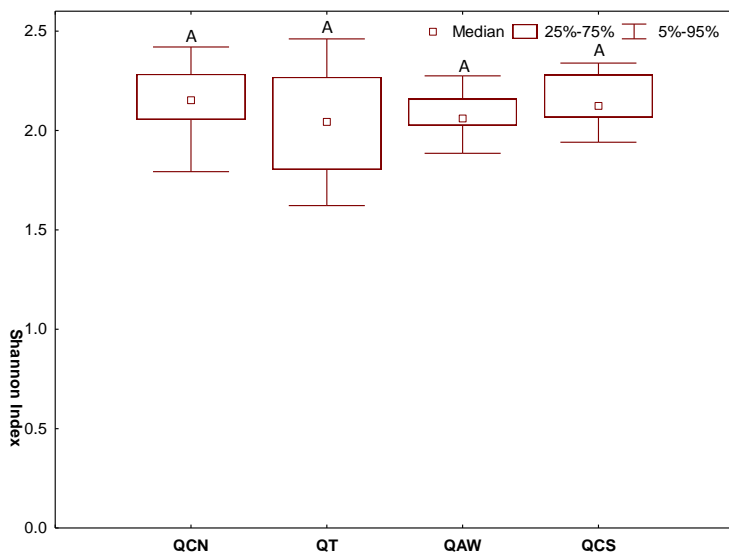


Figure 18: Box-whisker plot of Shannon Index per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

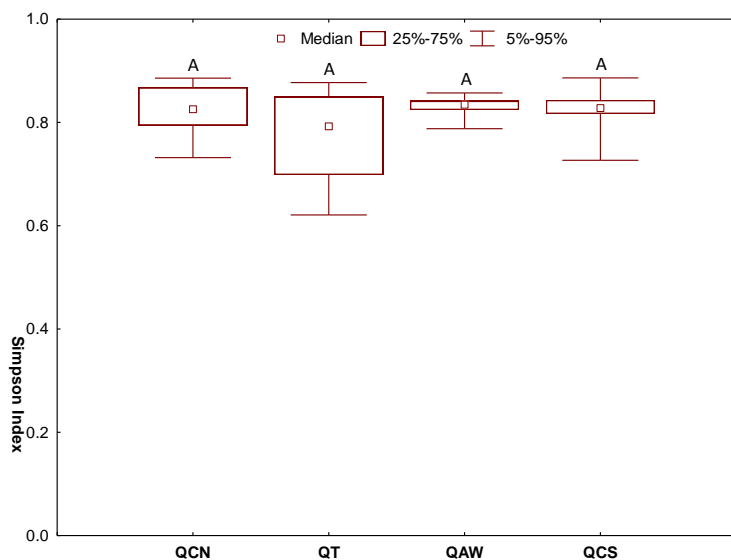


Figure 19: Box-whisker plot of Gini-Simpson Index per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

The species accumulation curve shows that an asymptotic value is almost reached with an effort of 39 samples (Figure 20). At that point, 88% of the estimated number of species was encountered in the samples.

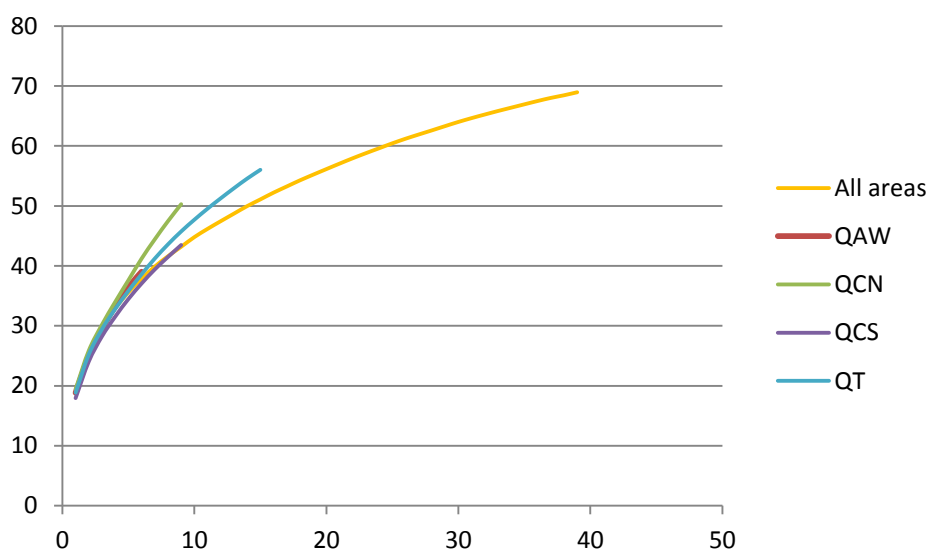


Figure 20: Dredge samples, estimated number of species in relation to the number of samples, determined with Bootstrapping.

Density and biomass

The mean dredge densities ranged from 140 ± 47 individuals/100 m² in QCS to 213 ± 36 individuals/100 m² in QAW. The highest density was observed in QT8 with 332 individuals/100 m², which was due to a high density of *Echinocardium cordatum* (171 individuals/100 m²). There were no statistically significant differences in density between

the areas (Figure 21). The most abundant species was *Echinocardium cordatum*, followed by *Ophiura ophiura*, *Spisula* spp. and *Ophiura albida*.

Table 8: Density and biomass data of the dredge sampling.

Area	Density (N/100m ²)				Biomass (g AFDW/100m ²)			
	Mean	St. Dev.	Max.	Min.	Mean	St. Dev.	Max.	Min.
QCN	158	58	247	82	80	34	128	31
QT	178	74	332	102	106	57	219	36
QAW	213	36	278	176	74	30	127	38
QCS	140	47	235	87	85	64	238	33
Total	170	63	332	82	91	52	238	31

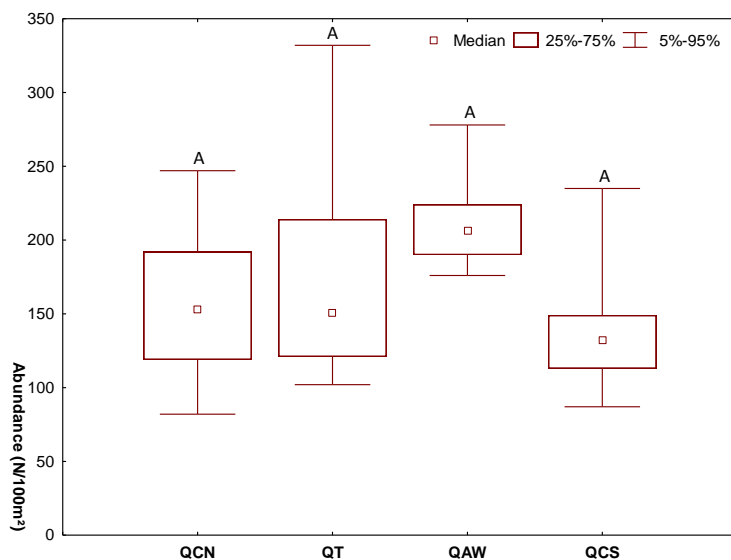


Figure 21: Box-whisker plot of abundance per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

The mean dredge biomass ranged from 74 ± 30 g AFDW/100 m² in QAW to 106 ± 57 g AFDW/100 m² in QT (Table 8). The highest biomass was observed in QSC6 with 238 g AFDW/100 m², which was due to a high density of *Echinocardium cordatum* (194 g AFDW/100 m²). There were no statistically significant differences in biomass between the areas (Figure 22). In all areas, *Echinocardium cordatum* was the dominant species in terms of biomass. The length-frequency distribution of *Echinocardium cordatum* in the different areas was similar (Figure 23), although slightly more large specimens of around 4 cm were captured in QT and QAW.

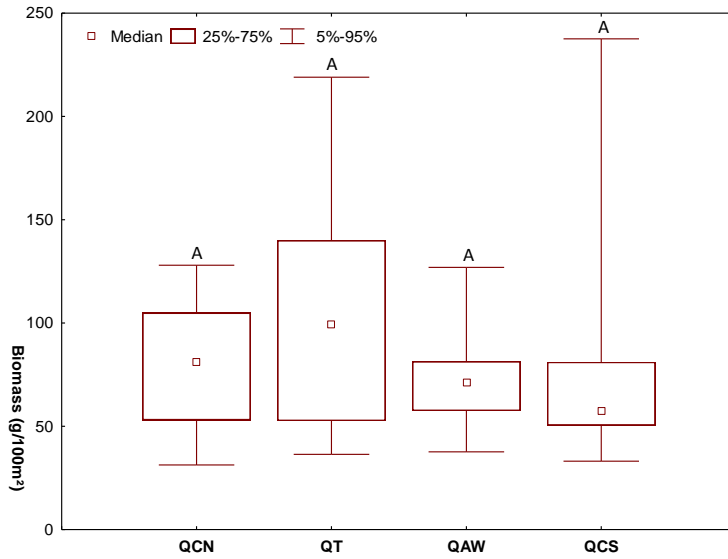


Figure 22: Box-whisker plot of biomass per area. Statistically significant differences (post-hoc multiple comparisons; $p < 0.05$) are indicated with different letters.

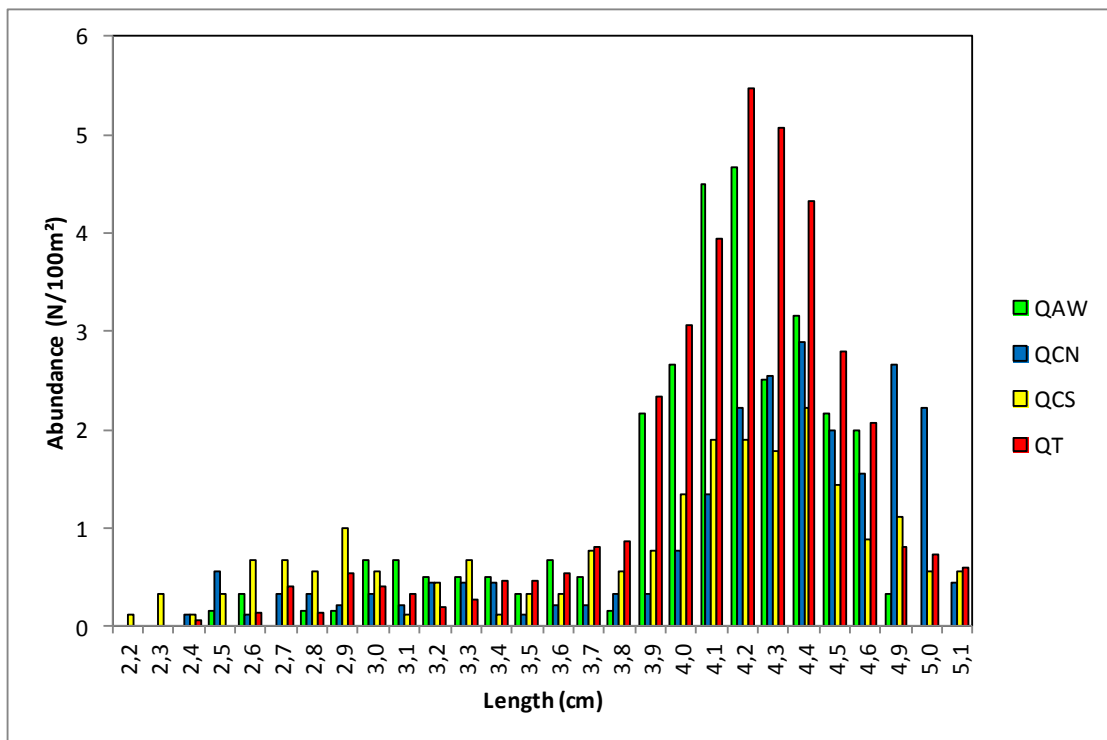


Figure 23: Length-frequency distribution of *Echinocardium cordatum* per area (outliers excluded).

Multivariate analyses

The ANOSIM analysis showed, as with the boxcore analysis, differences in densities between the different areas (Table 9). As with the boxcore samples, the MDS-plot also revealed a gradient from north to south (Figure 24). Dissimilarity between areas were mainly caused by differences in densities of *Donax vittatus*, *Ophelia borealis*, *Pomatoschistus* spp. and *Chamelea* spp.

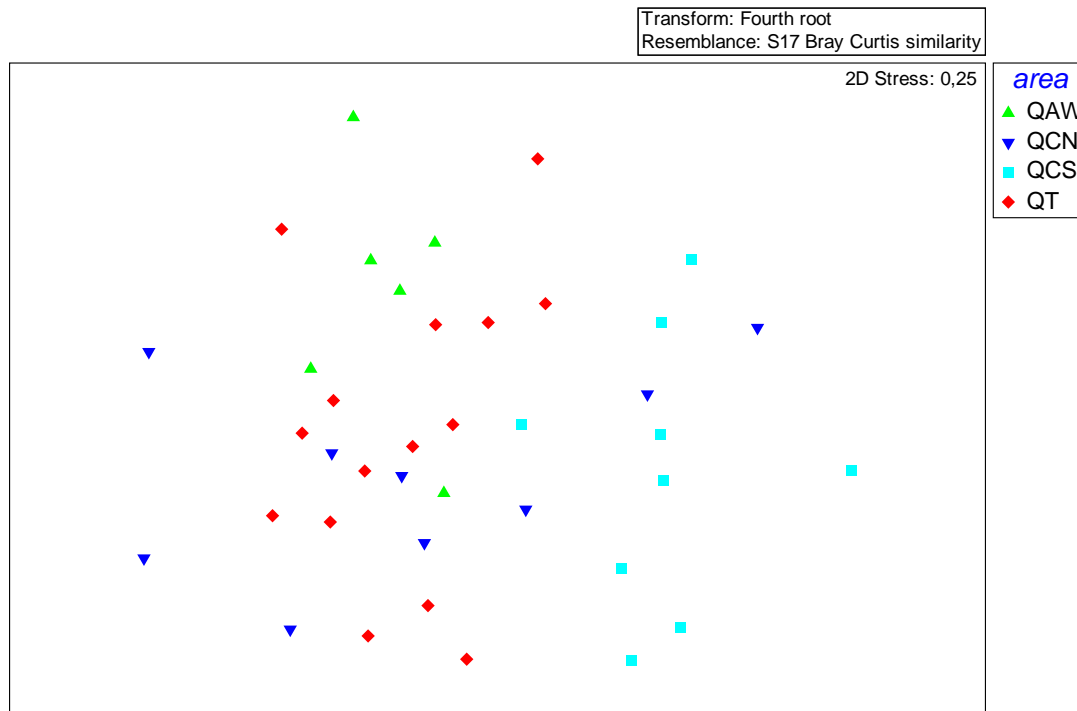


Figure 24: MDS plot based on density data in the dredge samples with indication of the different areas (note the stress factor of 0.25, which suggest a relatively low relevance of the sample clusters).

Table 9: Results ANOSIM and SIMPER analyses of the dredge samples.

Area	R	p	Dissimilarity (%)	Indicator species	Mean density Area 1 – Area 2	
QCN - QCS	0,499	0,001	41,7	<i>Donax vittatus</i>	2,14	0,42
				<i>Opbelia borealis</i>	1,06	1,96
				<i>Pomatoschistus spp.</i>	1,3	0,47
				<i>Asterias rubens</i>	1,06	0,22
				<i>Chamelea spp.</i>	1,17	0,35
QCN - QT	0,207	0,012	37,2	<i>Donax vittatus</i>	2,14	1,32
				<i>Opbelia borealis</i>	1,06	0,88
				<i>Pomatoschistus spp.</i>	1,30	0,78
				<i>Echinocardium cordatum</i>	2,12	2,67
				<i>Euspira pulchella</i>	1,04	0,91
QCS – QT	0,541	0,001	40,8	<i>Opbelia borealis</i>	1,96	0,88
				<i>Donax vittatus</i>	0,42	1,32
				<i>Asterias rubens</i>	0,22	1,14
				<i>Chamelea spp.</i>	0,35	1,21
				<i>Nephtys cirrosa</i>	0,89	0,30

Within the areas, there was a similarity of $\pm 40\%$, due to the presence of *Echinocardium cordatum*, *Ophiura* spp., *Donax vittatus* and *Spisula* spp. Species listed in Table 10 were specific to one or two areas and occurred in more than one sample. Three species (*Nemertea* spp. C, *Nephtys caeca*, *Arnoglossus laterna*) occurred only in the reference areas (north and south), but not in the turbine site.

Table 10: Dredge sample taxa specific to one or two areas and appearing in more than one sample.

QCN	QT and QAW	QCS
	<i>Sigalion mathildae</i>	
	<i>Nemertea</i> spp.	
	<i>Crangon almanni</i>	
	<i>Echiichthys vipera</i>	
	<i>Solea solea</i>	
	<i>Nereidae</i> spp.	
	<i>Callionymus reticulatus</i>	
	<i>Corystes cassivelaunus</i>	

Shell damage

Only four species of Bivalvia were regularly encountered: *Angulus fabula*, *Chamelea* spp., *Donax vittatus* and *Spisula* spp. *Ensis* spp. was also abundant, however, most shells were severely damaged during sampling and therefore, possible shell repair could not be evaluated for this genus. The number of damaged shells was very low overall (Table 11). *Angulus fabula*, the most fragile species, most frequently showed shell repair. For *Angulus fabula*, shell repair was only observed in the northern reference area, however, this species hardly occurred in the other areas. *Spisula* spp. also showed shell repair on a few occasions. Only *Spisula elliptica* showed shell repair, while the more robust *Spisula solida* never showed shell repair. Also *Donax vittatus* and *Chamelea* spp. never showed shell repair.

Table 11: Number of damaged shells showing shell repair and total number of encountered individuals for the frequently encountered Bivalvia per area.

	QCN		QT		QAW		QCS	
	Repair	Total	Repair	Total	Repair	Total	Repair	Total
<i>Angulus fabula</i>	6	172			3		1	
<i>Chamelea</i> spp.		30		46	27		4	
<i>Donax vittatus</i>		272		73	56		8	
<i>Spisula</i> spp.	2	102	1	264	176	1	193	

The length-frequency distribution of *Chamelea* spp. was similar in QT and QCN, while the species was hardly found in QCS and only small individuals were present in QAW (Figure 25). A higher abundance of *Donax vittatus* was observed in QCN, while the species was almost absent in QCS, however, the length was similar in the different areas (Figure 26). The length-frequency of *Spisula* spp. was similar in the different areas (Figure 27).

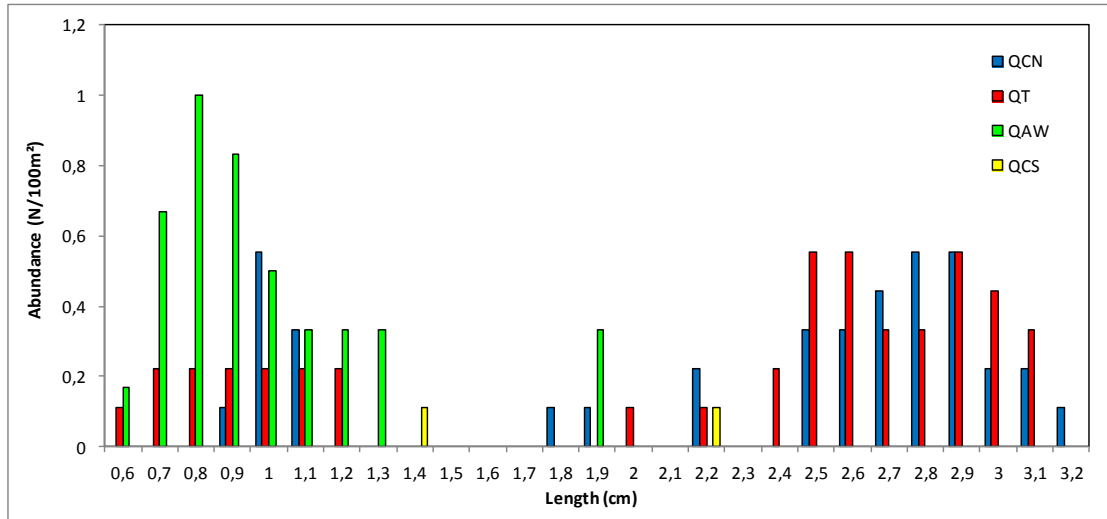


Figure 25: Length-frequency distribution of *Chamelea* spp. in the different areas.

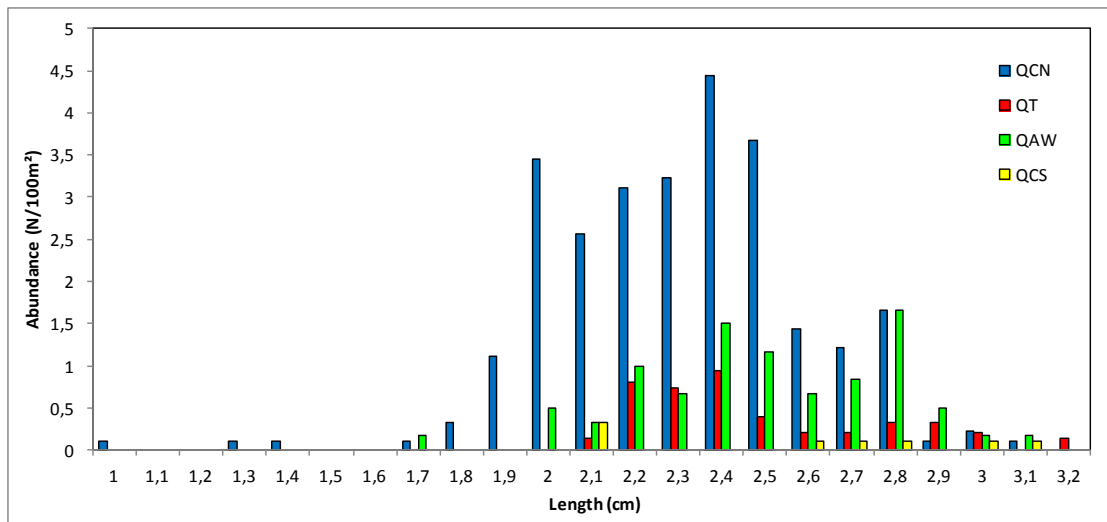


Figure 26: Length-frequency distribution of *Donax vittatus* in the different areas.

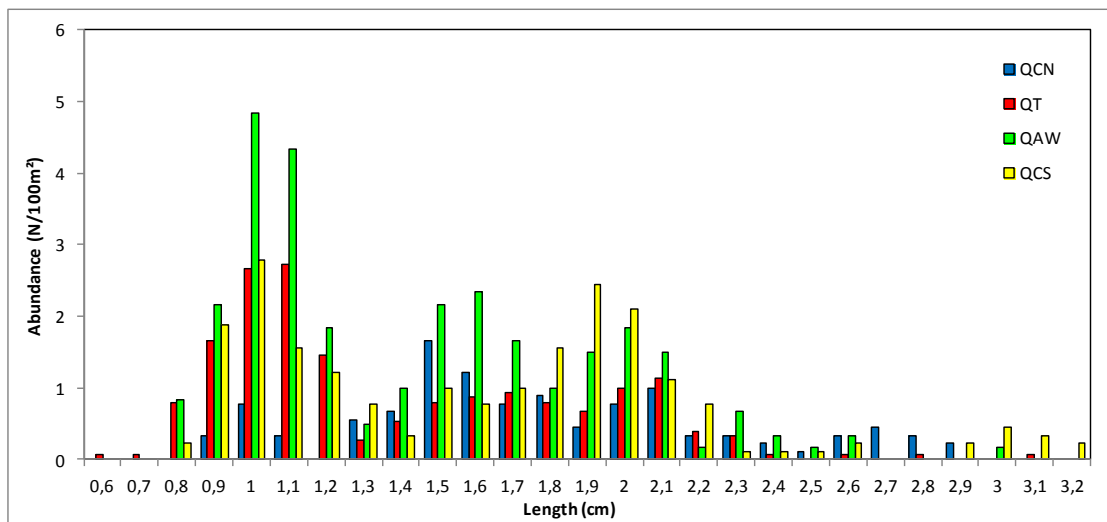


Figure 27: Length-frequency distribution of *Spisula* spp. in the different areas.

4. DISCUSSION

4.1 Development benthic invertebrate fauna Princess Amalia wind farm

In the present study, all sediments could be classified as medium sands and there was a small south to north transition from coarse to finer sediment. The finer sediment also contained slightly more organic matter.

The southern reference area, characterised by coarser sediment, contained the lowest total number of species and the lowest density in the boxcore samples. The sediments in the turbine site (QT) were intermediate in grain size. The mean invertebrate density and species richness in the boxcores were also intermediate. In the northern reference area, the sediment was the finest and also the highest density and species richness were found in the boxcore samples. The high densities were on account of *Urothoe poseidonis*, *Scoloplos armiger* and *Lanice conchilega*. *Asterias rubens*, *Nototropis falcatus* and *Pinnotheres pisum* were only found in the turbine area.

In the dredge samples, no significant differences were observed in density, biomass and species richness. *Echinocardium cordatum* dominated in terms of density as well as biomass. *Crangon almanni*, *Echiichthys vipera* and *Solea solea* were only found in the turbine area, although only a few individuals were encountered. As the dredge samples the larger, K-strategic species, it can be concluded that these species could not yet profit from the shelter from fishing activities five years after the installation of the wind farm. The vessel movement data confirm that no fishing activities are taking place in the wind farm (Koldenhof and de Jong, 2013). It thus seems that for the long-living species, five years is probably too short to show the recovery that was expected.

The species identified in the present study were typical for a sandy subtidal area. Such areas are often dominated by polychaete worms such as *Nephtys* spp., *Scoloplos armiger*, *Magelona* spp., *Chaetozone setosa*, *Ophelia borealis* and *Lanice conchilega*; amphipod genera such as *Urothoe* and *Bathyporeia* and the echinoid *Echinocardium cordatum* (Heip & Craeymeersch, 1995), which is in accordance with the present study.

Some of the typical sandy sediment species are classified as K-strategy species. K-strategy species are characterised by a slower growth rate, are longer-lived, grow bigger and generally start later with reproduction than r-strategy species. For example *Nephtys caeca* and other big polychaetes may get seven years old, *Chamelia striatula* 12 years and *Echinocardium cordatum* 10 or maybe 20 years (Van Moorsel, 2005). The latter species were found in comparable densities and length classes in the turbine area and in the reference areas. This means that no effect of the presence of the wind farm on K-strategy species of the area could yet be demonstrated.

4.2 Shell damage as indicator for reduced fishery pressure

Witbaard & Klein (1994) found that the incidence of sand grains incorporated into repairs in the shell matrix of bivalves increased coincidentally with the expansion of the Dutch Beam trawling fleet in the southern North Sea. Not only bivalves are damaged by beam trawling, also starfish could be damaged (Kaiser, 1996). Examination of shell scars of long-lived species provides a historical picture of fishing disturbance, whereas damaged starfish or specimens with regenerating arms are a useful short term (1 – 2 year) indicator (Witbaard & Klein, 1994; Kaiser, 1996). Since the closure of wind farms for fisheries is expected to increase opportunities for long-living bivalve species, this method was – qualitatively – used in the present study.

During the present study, only a few shells of the species *Angulus fabula* and *Spisula elliptica* were encountered that possessed scars. The number of scarred shells was therefore not sufficient for statistical analyses. The length-frequency distribution of the shells did not indicate that larger specimens were present in the turbine site compared to the reference sites.

4.3 Comparison with other wind farms in the Southern North Sea

The results of the Princess Amalia Wind Farm were compared with results from the baseline study T0 (Jarvis *et al.*, 2004), the T1 results of the OWEZ wind farm (Daan *et al.*, 2007), the Horns Rev wind farm (Bech *et al.*, 2005) and the wind farms on the Bligh Bank and the Thornton Bank (Coates & Vincx, 2010; Coates *et al.*, 2011, 2012). A direct comparison between the data collected during this T1 study and the T0 study is possible, because the same sample areas and stations were monitored. Also in T1 of OWEZ, the same sampling technique was used, while in the Horns Rev wind farm, the samples were collected by SCUBA divers.

PAWP T1 versus baseline study PAWP T0

The mean median particle size was significantly lower in present study than in the baseline T0 study: respectively 280.46 ± 28.07 and 582 ± 58.98 . The sediment in the baseline was classified as coarse sands, while in present study, it was median sands. The difference in sediment grain size is so substantial, that we have to argue that the results of the baseline study are not reliable, especially since the area is known to have median sands (Tempelman *et al.*, 2009). The mean percentage sand was comparable between both studies: around 99%. In the present study, we observed a small south to north transition from coarser to finer sediment; in the baseline, the same trend was observed. This gradual south to north transition along the Dutch coast was described earlier by Creutzberg *et al.* (1984). The organic matter content was slightly higher in T1 than in T0.

In the baseline boxcore samples, 109 benthic invertebrate taxa were identified, while in the present study only 58 taxa were found. There were almost twice as many polychaete and crustacean species found in T0. However, the number of treated samples was 200 in T0 versus 102 in T1. The mean species richness per sample was only slightly lower in T1 than in T0. A possible explanation for the observed difference in biodiversity could be due to the sampling season: samples were taken in May during T0 and in March during T1.

Densities in the boxcore samples were higher in T0, which was due to high densities of i.e. *Spiophanes bombyx*. During T1, this species was encountered on only three occasions. Notwithstanding the higher densities in T0, the biomasses were similar in both studies if large crabs (*Liocarcinus* spp.), razor clams (*Ensis* spp.) and sea urchins (*Echinocardium cordatum*) were not taken into account.

If polychaetes, Nemertea and fishes were not taken into account, 27 taxa were encountered in dredge samples in T1 versus 32 taxa in T0. In contrast to the boxcore samples, the mean species richness was slightly higher in dredge samples in T1 than in T0.

Densities from dredge samples in QCN were higher in T0 than in T1, due to high densities of *Donax vittatus* in T0. However, densities in the other areas were higher in T1 than in T0, which was mainly caused by higher densities of *Echinocardium cordatum* in T1.

OWEZ T1

Possible short-term effects of the construction of Offshore Windfarm Egmond aan Zee (OWEZ) on the composition of the local benthic fauna living in or on top of the sediment was evaluated a few months after the installation of the wind farm. During this study, benthic fauna was sampled with a boxcorer and a dredge within the wind farm and in six reference areas north and south of the wind farm. A high similarity between the fauna within the wind farm and the majority of the reference areas was observed. Density and diversity within the wind farm were well within the range of values observed in the reference areas. There was no indication that OWEZ differed in some way from the reference areas and there did not seem to be short-term effects of the construction of the wind farm on the local benthic fauna composition. These results are in accordance with the results obtained during the present study.

Horns Rev

The wind farm Horns Rev is situated 14-20 km off Blåvands Huk, which is the most western point of Denmark. One and two years after the wind farm became operational, benthic fauna was monitored. The sediment consisted of medium sand without organic matter. The benthic community at Horns Rev did not show signs of stress response as a consequence of possible impact from construction and operating activities. The density of the most abundant bivalves and bristle worms increased in the wind farm area, whereas the reference areas remained unchanged. This indicated that the potential

decrease in predation pressure from birds contributed to increasing differences between densities for their favoured prey. Bird observations showed predominant foraging activity outside the wind farm area. Sampling depth in Horns Rev was maximum 10.4 m, which is much less than the sampling depth during the present study, where the influence of birds is expected to be much less important.

Although the difference in sampling strategy - in Horns Rev the samples were taken by SCUBA divers at distances of 5, 25 and 100 m from the turbine, while in PAWP, the samples were taken with boxcores at a minimum distance of 150 m from the turbine – almost half of the soft substrate species found in Horns Rev were also encountered in this study. Only a few species from Horns Rev were related to hard substrate, such as *Eulalia viridis*, *Jassa marmorata*, *Balanus* spp. and *Mytilus edulis*. The most abundant species in Horns Rev, *Goodallia triangularis* and *Pisione remota*, were not found in the present study. High abundances as recorded for those species (in some years averages higher than 600 ind./m²) were not recorded in PAWP T1.

Bligh Bank and Thornton Bank

Results from the wind farms on the Bligh Bank and the Thornton Banks in Belgium show that with standard methods, no differences in sediment and benthos between turbine areas and reference sites could yet be found in the first years after construction (Coates & Vincx, 2010). These results are in line with the present study. However, a lower median grain size and higher macrobenthic densities were observed in 2010 one and seven meter from the scour protection system (boulders) of one turbine on the Thornton Bank (Coates *et al.*, 2011). These changes in the close vicinity of the turbines were explained by the decreased current speeds due to changing hydrodynamics or the relatively stable sand pits created during dredging activities. In 2011, an accumulation of juvenile starfish, together with the polychaetes *Spio* spp. and *Spiophanes bombyx* was obvious at stations less than 50 m from the scour protection of one turbine on the Thornton Bank (Coates *et al.*, 2012). Lower currents speeds and changing granulometric characteristics appeared to create a substantial change in the macrobenthic community six years after construction, especially on the Southwest gradient. This means that changes in soft sediment benthos could start from the immediate vicinity of the turbines and possibly spread from there. This research has evident implications for future sampling designs, where it might be advisable to include distance to the turbine foundations as a parameter for positioning sampling locations.

4.4 Remarkable findings

Some species found in the samples were previously unknown to the northern Dutch coast or rarely recorded in the area.

The hydromedusa (small jellyfish) *Eucheilota maculata* is common in coastal waters. Hydromedusae originate from a sedentary polyp stage (Figure 28). Polyps of this

particular medusa have rarely been recorded. In dredge samples of PAWP, the polyp was found several times on the inside of empty cockle valves. The delicate polyps are easily abraded during routine sample processing and difficult to identify. If the inside of empty bivalve shells is a preferred habitat, this could be an important cause of the paucity of records for *E. maculata*.

The scale worm *Malmgreniella darbouxi* was found both in boxcore and dredge samples. The species had been recorded from the coast of The Netherlands only once before (Wijsman & Verduin, 2011).

Paramysis arenosa is a hyperbenthic mysid, only caught with the dredge. This mysid has long been considered a southern species, not occurring in the North Sea (Tattersall & Tattersall, 1951). Recently, it was found off the coast of Belgium and Zeeland (De Wicke *et al.*, 2003). Apart from a doubtful old record from Northumberland (Tattersall & Tattersall, 1951), our observation near IJmuiden constitutes the most northerly record in the North Sea.

The necklace shell *Lunatia catena* and its eggs were only found in the dredge samples. The eggs have rarely been recorded (Figure 28).

In the dredge samples, a number of cut-off bivalve siphon tips with conspicuous reddish markings were found (Figure 28). They agree well with the siphon tips of *Lutraria* sp. (ottershell). Probably they belong to *Lutraria lutraria*. This bivalve lives up to 40 cm deep into the sediment, which is probably the reason why only part of the siphons was collected. It was absent from the boxcore samples. Apparently, its density is too low to appear regularly in boxcores. *Lutraria lutraria* has colonised coastal waters of the southern North Sea during the last decades.



Figure 28: A) Polyp of *Eucheilota maculata*; B) siphon tip of *Lutraria* sp.; C) eggs of *Lunatia catena*.

Striarca lactea is a southern bivalve reaching its northern limit in the southern North Sea and living only locally along the Dutch coast near Zeeland, mainly below 20 m (De Bruyne *et al.*, in prep.). Eisma (1966) mentions only one additional northerly record from a dredge sample.

Two rare species of bryozoans were found in a dredge sample: *Aspidelectra melolontha* and *Membranipora tenuis*. There are no recent records off the Dutch coast north of Zeeland. Both colonies may have been collected dead.

5. CONCLUSIONS

Biodiversity and abundance of soft sediment benthos vary between different wind farms in the Southern North Sea, yet densities within a windfarm are in most cases not consequently higher/lower than in reference areas, which indicates no measurable effect of the wind farm on densities of common benthos species. This pattern was confirmed during the present study.

Density and diversity of the macrobenthic fauna in the boxcore samples of the turbine site were intermediate between the northern and the southern reference area, while no significant differences were observed in terms of biomass. No significant differences in the density, diversity and biomass of the dredge samples, which samples larger K-strategic species, were observed between the areas. These results indicate that five years after installation of the wind farm, K-strategic species could not yet profit from the shelter from fishing activities. Bivalves with scars were rarely observed during the present study and the possible influence of reduced trawling could thus not be evaluated. The length-frequency distribution of the common bivalve species did not indicate that larger shells were found in the turbine site and no effects of reduced fishing activities could therefore be detected.

The used sampling strategy allowed to assess the effect of the installation of wind park on the macroinvertebrate community. We see no reasons to change the research program for the following campaigns.

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7. ANNEX LIST

Annex 1: Species list

Annex 2: Sampling coordinates

Annex 3: Raw data boxcore

Annex 4: Raw data dredge

Annex 5: Physical variables

Annex 1: Species list

¹: Species identified in the boxcore samples; ²: species identified in the dredge samples.

Phylum	Class	Species		
Annelida	Polychaeta	<i>Aricidea minuta</i> ¹		
		<i>Chaetozone setosa</i> ¹		
		<i>Eteone foliosa</i> ^{1, 2}		
		<i>Eteone longa</i> ¹		
		<i>Eunereis longissima</i> ^{1, 2}		
		<i>Lanice conchilega</i> ^{1, 2}		
		<i>Magelona filiformis</i> ¹		
		<i>Magelona johnstoni</i> ¹		
		<i>Magelona mirabilis</i> ¹		
		<i>Malmgreniella darbouxi</i> ¹		
		<i>Nephtys assimilis</i> ²		
		<i>Nephtys caeca</i> ^{1, 2}		
		<i>Nephtys cirrosa</i> ^{1, 2}		
		<i>Nephtys hombergii</i> ^{1, 2}		
		<i>Nereidae spp.</i> ²		
		<i>Ophelia borealis</i> ^{1, 2}		
		<i>Parexogone spp.</i> ¹		
		<i>Phyllodoce groenlandica</i> ²		
		<i>Phyllodoce lineata</i> ²		
		<i>Phyllodoce mucosa</i> ¹		
		<i>Poecilochaetus serpens</i> ¹		
		<i>Scolelepis bonnieri</i> ^{1, 2}		
		<i>Scoloplos armiger</i> ¹		
		<i>Sigalion mathildae</i> ²		
		<i>Spio gonioccephala</i> ¹		
		<i>Spio spp.</i> ²		
		<i>Spiophanes bombyx</i> ¹		
		Arthropoda	Malacostraca (Amphipoda)	<i>Abludomelita obtusata</i> ¹
				<i>Bathyporeia elegans</i> ¹
<i>Bathyporeia guilliamsoniana</i> ¹				
<i>Bathyporeia nana</i> ¹				
<i>Leucothoe incisa</i> ¹				
<i>Megaluropus agilis</i> ¹				
<i>Nototropis falcatus</i> ¹				
<i>Perioculodes longimanus</i> ¹				
<i>Pontocrates arcticus</i> ¹				
<i>Urothoe brevicornis</i> ¹				
<i>Urothoe poseidonis</i> ¹				
Malacostraca (Cumacea)	<i>Diastylis cf. rugosa</i> ¹			
	<i>Monopseudocuma gilsoni</i> ¹			
	<i>Pseudocuma longicorne</i> ¹			

Annex 1: species list

Phylum	Class	Species
	Malacostraca (Decapoda)	<i>Corystes cassivelaunus</i> ² <i>Crangon almanni</i> ² <i>Crangon crangon</i> ² <i>Liocarcinus holsatus</i> ^{1, 2} <i>Liocarcinus marmoreus</i> ² <i>Macropodia rostrata</i> ² <i>Pagurus bernhardus</i> ² <i>Pestarella tyrrhena</i> ¹ <i>Philocheras trispinosus</i> ² <i>Pinnotheres pisum</i> ^{1, 2} <i>Processa modica</i> ¹ <i>Processa spp.</i> ² <i>Thia scutellata</i> ^{1, 2}
	Malacostraca (Mysida)	<i>Gastrosaccus spinifer</i> ^{1, 2}
	Maxillopoda	<i>Sacculina carcini</i> ²
Chordata	Actinopterygii	<i>Ammodytes spp.</i> ² <i>Arnoglossus laterna</i> ² <i>Buglossidium luteum</i> ² <i>Callionymus lyra</i> ² <i>Callionymus reticulatus</i> ² <i>Echiichthys vipera</i> ² <i>Gymnammodytes semisquamatus</i> ² <i>Hyperoplus lanceolatus</i> ² <i>Limanda limanda</i> ² <i>Pleuronectes platessa</i> ² <i>Pomatoschistus spp.</i> ² <i>Solea solea</i> ² <i>Taurulus bubalis</i> ²
	Leptocardii	<i>Branchiostoma lanceolatum</i> ¹
Echinodermata	Asteroidea	<i>Asterias rubens</i> ^{1, 2}
	Echinoidea	<i>Echinocardium cordatum</i> ^{1, 2}
	Ophiuroidea	<i>Ophiura albida</i> ^{1, 2} <i>Ophiura ophiura</i> ^{1, 2}
Mollusca	Bivalvia	<i>Angulus fabula</i> ^{1, 2} <i>Angulus tenuis</i> ² <i>Chamelea striatula</i> ^{1, 2} <i>Donax vittatus</i> ^{1, 2} <i>Ensis directus</i> ^{1, 2} <i>Ensis ensis</i> ² <i>Lutraria spp.</i> ² <i>Macoma balthica</i> ¹ <i>Spisula spp.</i> ^{1, 2} <i>Spisula subtruncata</i> ¹ <i>Striarca lactea</i> ² <i>Tellimya ferruginosa</i> ^{1, 2}
	Gastropoda	<i>Euspira pulchella</i> ^{1, 2}

Phylum	Class	Species
Nemertea	/	<i>Nemertea spp. A</i> ^{1, 2}
		<i>Nemertea spp. B</i> ^{1, 2}
		<i>Nemertea spp. C</i> ²
Phoronida	/	<i>Phoronis spp.</i> ¹

Station:	Time	Date	Coordinate
QCN1	12u50	12/03/2012	N52 48.350 E4 21.190
QCN2	13u49	12/03/2012	N52 48.238 E4 21.893
QCN3.0	13u58	12/03/2012	N52 48.216 E4 22.553
QCN3.1	14u07	12/03/2012	N52 48.216 E4 22.553
QCN3.2	14u13	12/03/2012	N52 48.216 E4 22.553
QCN3.3	14u41	12/03/2012	N52 48.216 E4 22.553
QCN3.4	15u08	12/03/2012	N52 48.216 E4 22.553
QCN4	16u08	12/03/2012	N52 48.185 E4 23.228
QCN5	16u13	12/03/2012	N52 48.105 E4 23.878
QCN10	16u25	12/03/2012	N52 47.589 E4 23.835
QCN9	16u37	12/03/2012	N52 47.681 E4 23.178
QCN8	16u44	12/03/2012	N52 47.763 E4 22.500
QCN7	16u54	12/03/2012	N52 47.825 E4 21.795
QCN6	17u03	12/03/2012	N52 47.902 E4 21.086
QCN11.0	17u13	12/03/2012	N52 47.436 E4 21.031
QCN11.1	17u19	12/03/2012	N52 47.467 E4 21.030
QCN11.2	17u29	12/03/2012	N52 47.467 E4 21.030
QCN11.3	17u39	12/03/2012	N52 47.467 E4 21.030
QCN11.4	17u48	12/03/2012	N52 47.467 E4 21.030
QCN12	17u57	12/03/2012	N52 47.392 E4 21.776
QCN13.0	18u08	12/03/2012	N52 47.341 E4 22.426
QCN13.1	18u16	12/03/2012	N52 47.341 E4 22.426
QCN13.2	18u26	12/03/2012	N52 47.341 E4 22.426
QCN13.3	18u37	12/03/2012	N52 47.341 E4 22.426
QCN13.4	18u46	12/03/2012	N52 47.341 E4 22.426
QCN14	11u33	13/03/2012	N52 47.234 E4 23.096
QCN15.4	12u16	13/03/2012	N52 47.149 E4 23.826
QCN15.0	11u41	13/03/2012	N52 47.149 E4 23.826
QCN15.1	11u50	13/03/2012	N52 47.149 E4 23.826
QCN15.2	11u57	13/03/2012	N52 47.149 E4 23.826
QCN15.3	12u06	13/03/2012	N52 47.149 E4 23.826
QCN20	12u26	13/03/2012	N52 46.698 E4 23.738
QCN19	12u37	13/03/2012	N52 46.803 E4 23.089
QCN18	12u48	13/03/2012	N52 46.888 E4 22.397
QCN17	12u56	13/03/2012	N52 46.942 E4 21.724
QCN16	13u07	13/03/2012	N52 47.013 E4 21.020
QCN21	14u24	13/03/2012	N52 46.625 E4 21.012
QCN22	14u34	13/03/2012	N52 46.497 E4 21.734
QCN23.2	15u00	13/03/2012	N52 46.476 E4 22.407
QCN23.3	15u09	13/03/2012	N52 46.476 E4 22.407
QCN23.4	15u16	13/03/2012	N52 46.476 E4 22.407
QCN23.0	14u42	13/03/2012	N52 46.476 E4 22.407
QCN23.1	14u52	13/03/2012	N52 46.476 E4 22.407
QCN24	15u25	13/03/2012	N52 46.413 E4 23.087
QCN25	15u38	13/03/2012	N52 46.315 E4 23.735
QAW4.0	17u00	13/03/2012	N52 36.800 E4 12.985
QAW4.1	17u06	13/03/2012	N52 36.800 E4 12.985

Annex 2: Coordinates

QAW4.2	17u12	13/03/2012	N52 36.800 E4 12.985
QAW4.3	17u17	13/03/2012	N52 36.800 E4 12.985
QAW4.4	17u25	13/03/2012	N52 36.800 E4 12.985
QAW1.0	17u35	13/03/2012	N52 36.514 E4 13.031
QAW1.1	17u41	13/03/2012	N52 36.514 E4 13.031
QAW1.2	17u47	13/03/2012	N52 36.514 E4 13.031
QAW1.3	17u53	13/03/2012	N52 36.514 E4 13.031
QAW1.4	18u00	13/03/2012	N52 36.514 E4 13.031
QAW2.0	18u19	13/03/2012	N52 35.858 E4 15.258
QAW2.1	18u28	13/03/2012	N52 35.858 E4 15.258
QAW2.2	18u37	13/03/2012	N52 35.858 E4 15.258
QAW2.3	18u46	13/03/2012	N52 35.858 E4 15.258
QAW2.4	18u55	13/03/2012	N52 35.858 E4 15.258
QAW3.0	19u02	13/03/2012	N52 36.011 E4 15.818
QAW3.1	19u15	13/03/2012	N52 36.011 E4 15.818
QAW3.2	19u26	13/03/2012	N52 36.011 E4 15.818
QAW3.3	19u38	13/03/2012	N52 36.011 E4 15.818
QAW3.4	19u50	13/03/2012	N52 36.011 E4 15.818
QAW6.1	18u17	15/03/2012	N52 33.805 E4 12.689
QAW6.2	18u23	15/03/2012	N52 33.805 E4 12.689
QAW6.3	18u28	15/03/2012	N52 33.805 E4 12.689
QAW6.4	18u32	15/03/2012	N52 33.805 E4 12.689
QAW6.0	9u53	14/03/2012	N52 33.805 E4 12.689
QAW5.1	18u40	15/03/2012	N52 34.168 E4 12.823
QAW5.2	18u47	15/03/2012	N52 34.168 E4 12.823
QAW5.3	18u54	15/03/2012	N52 34.168 E4 12.823
QAW5.4	18u59	15/03/2012	N52 34.168 E4 12.823
QAW5.0	10u03	14/03/2012	N52 34.168 E4 12.823
QT42	10u25	14/03/2012	N52 35.431 E4 11.601
QT37	10u32	14/03/2012	N52 35.624 E4 12.089
QT38	10u37	14/03/2012	N52 35.811 E4 12.563
QT39	10u45	14/03/2012	N52 35.971 E4 13.014
QT40.1	9u05	16/03/2012	N52 36.173 E4 13.547
QT40.2	9u10	16/03/2012	N52 36.173 E4 13.547
QT40.3	9u15	16/03/2012	N52 36.173 E4 13.547
QT40.4	9u20	16/03/2012	N52 36.173 E4 13.547
QT40.0	10u50	14/03/2012	N52 36.173 E4 13.547
QT41	10u58	14/03/2012	N52 36.279 E4 13.870
QT45	11u11	14/03/2012	N52 36.170 E4 12.827
QT44.1	8u34	16/03/2012	N52 36.011 E4 12.328
QT44.2	8u46	16/03/2012	N52 36.011 E4 12.328
QT44.3	8u53	16/03/2012	N52 36.011 E4 12.328
QT44.4	8u59	16/03/2012	N52 36.011 E4 12.328
QT44.0	11u16	14/03/2012	N52 36.011 E4 12.328
QT43	11u25	14/03/2012	N52 35.808 E4 11.835
QT46	11u34	14/03/2012	N52 36.210 E4 12.022
QT36	12u08	14/03/2012	N52 36.050 E4 14.122
QT35	12u14	14/03/2012	N52 35.890 E4 13.681
QT34	12u23	14/03/2012	N52 35.720 E4 13.245
QT33.1	9u27	16/03/2012	N52 35.557 E4 12.787

QT33.2	9u34	16/03/2012	N52 35.557 E4 12.787
QT33.3	9u39	16/03/2012	N52 35.557 E4 12.787
QT33.4	9u45	16/03/2012	N52 35.557 E4 12.787
QT33.0	12u30	14/03/2012	N52 35.557 E4 12.787
QT32	12u37	14/03/2012	N52 35.375 E4 12.319
QT31	12u43	14/03/2012	N52 35.198 E4 11.855
QT48	12u54	14/03/2012	N52 35.028 E4 11.440
QT1	13u00	14/03/2012	N52 34.800 E4 11.719
QT2.1	9u57	16/03/2012	N52 35.006 E4 12.173
QT2.2	10u41	16/03/2012	N52 35.006 E4 12.173
QT2.3	10u47	16/03/2012	N52 35.006 E4 12.173
QT2.4	10u53	16/03/2012	N52 35.006 E4 12.173
QT2.0	13u07	14/03/2012	N52 35.006 E4 12.173
QT3	13u14	14/03/2012	N52 35.168 E4 12.622
QT4	13u20	14/03/2012	N52 35.325 E4 13.096
QT5	13u28	14/03/2012	N52 35.495 E4 13.546
QT6.1	12u30	16/03/2012	N52 35.680 E4 14.032
QT6.2	12u36	16/03/2012	N52 35.680 E4 14.032
QT6.3	12u43	16/03/2012	N52 35.680 E4 14.032
QT6.4	12u48	16/03/2012	N52 35.680 E4 14.032
QT6.0	13u36	14/03/2012	N52 35.680 E4 14.032
QT7	13u44	14/03/2012	N52 35.860 E4 14.481
QT13	14u58	14/03/2012	N52 35.673 E4 14.735
QT12	15u03	14/03/2012	N52 35.495 E4 14.271
QT11	15u10	14/03/2012	N52 35.325 E4 13.785
QT10.1	11u06	16/03/2012	N52 35.149 E4 13.324
QT10.2	11u12	16/03/2012	N52 35.149 E4 13.324
QT10.3	11u16	16/03/2012	N52 35.149 E4 13.324
QT10.4	11u24	16/03/2012	N52 35.149 E4 13.324
QT10.0	15u18	14/03/2012	N52 35.149 E4 13.324
QT9	15u24	14/03/2012	N52 34.955 E4 12.873
QT8	15u29	14/03/2012	N52 34.794 E4 12.407
QT47	15u34	14/03/2012	N52 34.635 E4 11.976
QT14	15u42	14/03/2012	N52 34.564 E4 12.705
QT15	15u50	14/03/2012	N52 34.743 E4 13.183
QT16	15u57	14/03/2012	N52 34.927 E4 13.660
QT18.1	12u06	16/03/2012	N52 35.292 E4 14.638
QT18.2	12u12	16/03/2012	N52 35.292 E4 14.638
QT18.3	12u17	16/03/2012	N52 35.292 E4 14.638
QT18.4	12u21	16/03/2012	N52 35.292 E4 14.638
QT18.0	16u05	14/03/2012	N52 35.292 E4 14.638
QT19	16u12	14/03/2012	N52 35.446 E4 15.103
QT24	16u18	14/03/2012	N52 35.213 E4 15.349
QT23	16u24	14/03/2012	N52 35.067 E4 14.913
QT22	16u31	14/03/2012	N52 34.891 E4 14.422
QT21.1	11u25	16/03/2012	N52 34.713 E4 13.957
QT21.2	11u40	16/03/2012	N52 34.713 E4 13.957
QT21.3	11u47	16/03/2012	N52 34.713 E4 13.957
QT21.4	11u54	16/03/2012	N52 34.713 E4 13.957
QT21.0	16u36	14/03/2012	N52 34.713 E4 13.957

Annex 2: Coordinates

QT20	16u42	14/03/2012	N52 34.545 E4 13.525
QT25	16u50	14/03/2012	N52 34.537 E4 14.218
QT30	17u26	14/03/2012	N52 34.526 E4 14.979
QT29	17u34	14/03/2012	N52 34.353 E4 14.519
QT26	17u42	14/03/2012	N52 34.719 E4 14.671
QT27	17u48	14/03/2012	N52 34.909 E4 15.180
QT28	17u53	14/03/2012	N52 35.055 E4 15.630
QCS5	9u59	15/03/2012	N52 26.817 E4 02.260
QCS10	10u06	15/03/2012	N52 26.577 E4 01.854
QCS15.0	10u13	15/03/2012	N52 26.334 E4 01.382
QCS15.1	10u20	15/03/2012	N52 26.334 E4 01.382
QCS15.2	10u27	15/03/2012	N52 26.334 E4 01.382
QCS15.3	10u32	15/03/2012	N52 26.334 E4 01.382
QCS15.4	10u39	15/03/2012	N52 26.334 E4 01.382
QCS20	10u45	15/03/2012	N52 26.092 E4 00.837
QCS25	10u56	15/03/2012	N52 25.765 E4 00.376
QCS24	11u04	15/03/2012	N52 25.983 E3 59.724
QCS19	11u10	15/03/2012	N52 26.292 E4 00.239
QCS14	11u16	15/03/2012	N52 26.535 E4 00.800
QCS9	11u23	15/03/2012	N52 26.767 E4 01.213
QCS4	11u30	15/03/2012	N52 27.022 E4 01.687
QCS3.0	11u40	15/03/2012	N52 27.168 E4 01.098
QCS3.1	11u48	15/03/2012	N52 27.168 E4 01.098
QCS3.2	11u53	15/03/2012	N52 27.168 E4 01.098
QCS3.3	12u00	15/03/2012	N52 27.168 E4 01.098
QCS3.4	12u05	15/03/2012	N52 27.168 E4 01.098
QCS8	12u30	15/03/2012	N52 26.930 E4 00.654
QCS13.0	12u36	15/03/2012	N52 26.685 E4 00.198
QCS13.1	12u43	15/03/2012	N52 26.685 E4 00.198
QCS13.2	12u48	15/03/2012	N52 26.685 E4 00.198
QCS13.3	12u53	15/03/2012	N52 26.685 E4 00.198
QCS13.4	12u59	15/03/2012	N52 26.685 E4 00.198
QCS18	13u07	15/03/2012	N52 26.456 E3 59.619
QCS23.0	13u14	15/03/2012	N52 26.171 E3 59.131
QCS23.1	13u19	15/03/2012	N52 26.171 E3 59.131
QCS23.2	13u25	15/03/2012	N52 26.171 E3 59.131
QCS23.3	13u31	15/03/2012	N52 26.171 E3 59.131
QCS23.4	13u39	15/03/2012	N52 26.171 E3 59.131
QCS22	13u39	15/03/2012	
QCS17	14u45	15/03/2012	N52 26.352 E3 58.478
QCS12	14u58	15/03/2012	N52 26.625 E3 59.014
QCS7	15u07	15/03/2012	N52 26.877 E3 59.570
QCS2	15u14	15/03/2012	N52 27.101 E4 00.002
QCS1	15u21	15/03/2012	N52 27.365 E4 00.560
QCS6	15u30	15/03/2012	N52 27.549 E3 59.955
QCS11.0	15u37	15/03/2012	N52 27.282 E3 59.366
QCS11.1	15u47	15/03/2012	N52 27.067 E3 58.959
QCS11.2	15u53	15/03/2012	N52 27.067 E3 58.959
QCS11.2	16u00	13/03/2012	N52 27.067 E3 58.959
QCS11.3	16u06	13/03/2012	N52 27.067 E3 58.959

Annex 2: Coordinates

QCS11.4	16u12	13/03/2012	N52 27.067 E3 58.959
QCS16	16u18	13/03/2012	N52 26.825 E3 58.438
QCS21	16u26	13/03/2012	N52 26.545 E3 57.849

Annex 3: Box core data

Area Sample Species	QCN							
	QCN 1	QCN 2	QCN 3.0	QCN 4	QCN 5	QCN 6	QCN 7	QCN 8
<i>Periocolodes longimanus</i>								
<i>Pestarella tyrrhena</i>								
<i>Phoronis</i> spp.								
<i>Phyllodoce mucosa</i>								
<i>Pinnotheres pisum</i>								
<i>Poecilochaetus serpens</i>								
<i>Pontocrates arcticus</i>				13			13	
<i>Processa modica</i>								
<i>Pseudocuma longicorne</i>								
<i>Scolecopsis bonnieri</i>					26	13		
<i>Scoloplos armiger</i>	26	26	51	26	13	51	115	13
<i>Spio goniocephala</i>		26						
<i>Spiophanes bombyx</i>								
<i>Spisula</i> spp.								
<i>Spisula subtruncata</i>								
<i>Tellimya ferruginosa</i>			13	13		13	26	
<i>Thia scutellata</i>								
<i>Tubularia indivisa</i>								
<i>Urothoe brevicornis</i>		13				13	26	
<i>Urothoe poseidonis</i>					372		13	
<i>Urothoe</i> spp.			13					

Annex 3: Box core data

Area	QC	QT	QT	QT	QT	QT	QT	QT
Sample	25	1	2	3	4	5	6.0	7
Species								
<i>Pontocrates arcticus</i>		13		13				26
<i>Processa modica</i>						13		
<i>Pseudocuma longicorne</i>								
<i>Scolecopsis bonnierii</i>	13							
<i>Scoloplos armiger</i>	51	13	26	115	103	77	38	103
<i>Spio goniocephala</i>			13					
<i>Spiophanes bombyx</i>				13				
<i>Spisula</i> spp.								
<i>Spisula subtruncata</i>								
<i>Tellimya ferruginosa</i>	13							
<i>Thia scutellata</i>								
<i>Tubularia indivisa</i>								
<i>Urothoe brevicornis</i>			13				13	26
<i>Urothoe poseidonis</i>								
<i>Urothoe</i> spp.	13							

Area	QT 8	QT 9	QT 10.0	QT 11	QT 12	QT 13	QT 14	QT 15
Abludomelita obtusata								
Angulus fabula								
Aricidea minuta								
Asterias rubens			13					
Bathyporeia cf. elegans	13	13						
Bathyporeia cf. nana								
Bathyporeia elegans	13	13	38	90	51	192	26	38
Bathyporeia guilliamsoniana			26	13	26	13		38
Bathyporeia nana								
Bathyporeia spp.						13		
Branchiostoma lanceolatum				13				
Chaetozone setosa			26		51			38
Chamelea striatula								
Diastylis cf. rugosa	13							
Donax vittatus								
Echinocardium cordatum	26	13	26		13		13	
Ensis directus								
Eteone foliosa			13					
Eteone longa	13	13	13					13
Eunereis longissima			13					13
Euspira pulchella			26					
Exogone spp.								
Gastrosaccus spinifer	13							
Gastrosaccus spp.		13						
Lanice conchilega	13							
Leucothoe incisa	13		13					
Liocarcinus cf. holsatus								
Liocarcinus holsatus								
Macoma balthica						13		
Magelona filiformis	13		90		13		26	13
Magelona johnstoni		13	13	13		13		
Magelona mirabilis					26			
Magelona spp.								
Malmgreniella darbouxi								
Megaluropus agilis								
Monopseudocuma gilsoni								
Nemertea spp. A	38		26				13	
Nemertea spp. B								13
Nephtys caeca								
Nephtys cirrosa	26	64	26	38	77	64	26	64
Nephtys hombergii				13				
Nototropis falcatus						13		
Ophelia borealis		13				13		
Ophiura albida			13					
Ophiura ophiura								
Perioculodes longimanus								
Pestarella tyrrhena				13				
Phoronis spp.					13			
Phyllodoce mucosa								
Pinnotheres pisum						13		
Poecilochaetus serpens			13					167

Area	QT 16	QT 18.0	QT 19	QT 20	QT 21.0	QT 22	QT 23	QT 24
Sample								
Species								
<i>Abludomelita obtusata</i>								
<i>Angulus fabula</i>								
<i>Aricidea minuta</i>								
<i>Asterias rubens</i>								
<i>Bathyporeia</i> cf. <i>elegans</i>								
<i>Bathyporeia</i> cf. <i>nana</i>								
<i>Bathyporeia elegans</i>	51	115	77	90	77	51	51	51
<i>Bathyporeia guilliamsoniana</i>	13	26	26	13	38			13
<i>Bathyporeia nana</i>								
<i>Bathyporeia</i> spp.					13			
<i>Branchiostoma lanceolatum</i>								
<i>Chaetozone setosa</i>	26		13	26	38		13	13
<i>Chamelea striatula</i>	13							
<i>Diastylis</i> cf. <i>rugosa</i>								
<i>Donax vittatus</i>								13
<i>Echinocardium cordatum</i>		13	13	26	13	13		
<i>Ensis directus</i>								
<i>Eteone foliosa</i>							13	
<i>Eteone longa</i>				13	13	38	26	
<i>Eunereis longissima</i>								
<i>Euspira pulchella</i>								
<i>Exogone</i> spp.								
<i>Gastrosaccus spinifer</i>								
<i>Gastrosaccus</i> spp.								
<i>Lanice conchilega</i>								
<i>Leucothoe incisa</i>								
<i>Liocarcinus</i> cf. <i>holsatus</i>								
<i>Liocarcinus holsatus</i>								
<i>Macoma balthica</i>								13
<i>Magelona filiformis</i>	51		13	38		13		26
<i>Magelona johnstoni</i>				13		13	13	
<i>Magelona mirabilis</i>		13			13		13	
<i>Magelona</i> spp.								
<i>Malmgreniella darbouxi</i>								
<i>Megaluropus agilis</i>								13
<i>Monopseudocuma gilsoni</i>								
<i>Nemertea</i> spp. A	38			26		26		26
<i>Nemertea</i> spp. B								
<i>Nephtys caeca</i>								
<i>Nephtys cirrosa</i>		13	26	90	26	26	38	38
<i>Nephtys hombergii</i>								
<i>Nototropis falcatus</i>								
<i>Ophelia borealis</i>	13			13				
<i>Ophiura albida</i>								
<i>Ophiura ophiura</i>								
<i>Perioculodes longimanus</i>								
<i>Pestarella tyrrhena</i>								
<i>Phoronis</i> spp.								
<i>Phyllodoce mucosa</i>							13	
<i>Pinnotheres pisum</i>								
<i>Poecilochaetus serpens</i>				38				

Annex 3: Box core data

Area Sample Species	QT 25	QT 26	QT 27	QT 28	QT 29	QT 30	QT 31	QT 32
<i>Pontocrates arcticus</i>			13	13				
<i>Processa modica</i>								
<i>Pseudocuma longicorne</i>								
<i>Scolecopsis bonnieri</i>					13			
<i>Scoloplos armiger</i>	51	51	38	13	38	26	64	51
<i>Spio goniocephala</i>								
<i>Spiophanes bombyx</i>								
<i>Spisula</i> spp.								
<i>Spisula subtruncata</i>								
<i>Tellimya ferruginosa</i>								
<i>Thia scutellata</i>					13			
<i>Tubularia indivisa</i>								
<i>Urothoe brevicornis</i>		26	64	77	13	38		
<i>Urothoe poseidonis</i>								
<i>Urothoe</i> spp.					13	13		

Area Sample Species	QAW						
	QCS 25	QAW 1.0	QAW 2.0	QAW 3.0	QAW 4.0	QAW 5.0	QAW 6.0
<i>Abludomelita obtusata</i>							
<i>Angulus fabula</i>							
<i>Aricidea minuta</i>							
<i>Asterias rubens</i>							
<i>Bathyporeia</i> cf. <i>elegans</i>							
<i>Bathyporeia</i> cf. <i>nana</i>							13
<i>Bathyporeia elegans</i>	13	77	51	26	13	38	26
<i>Bathyporeia guilliamsoniana</i>	13					77	
<i>Bathyporeia nana</i>							
<i>Bathyporeia</i> spp.			13				
<i>Branchiostoma lanceolatum</i>							
<i>Chaetozone setosa</i>			13		38		13
<i>Chamelea striatula</i>							
<i>Diastylis</i> cf. <i>rugosa</i>							
<i>Donax vittatus</i>							
<i>Echinocardium cordatum</i>	13			38		13	26
<i>Ensis directus</i>			13				13
<i>Eteone foliosa</i>							
<i>Eteone longa</i>		13	13		13	77	
<i>Eunereis longissima</i>							
<i>Euspira pulchella</i>							
<i>Exogone</i> spp.							
<i>Gastrosaccus spinifer</i>		13					
<i>Gastrosaccus</i> spp.							
<i>Lanice conchilega</i>							
<i>Leucothoe incisa</i>							
<i>Liocarcinus</i> cf. <i>holsatus</i>							
<i>Liocarcinus holsatus</i>							
<i>Macoma balthica</i>		13			13		
<i>Magelona filiformis</i>			26		13	38	
<i>Magelona johnstoni</i>				13		13	
<i>Magelona mirabilis</i>	13				38		
<i>Magelona</i> spp.							
<i>Malmgreniella darbouxi</i>							
<i>Megaluropus agilis</i>					13		
<i>Monopseudocuma gilsoni</i>							13
<i>Nemertea</i> spp. A		26	13		26	13	
<i>Nemertea</i> spp. B					13		
<i>Nephtys caeca</i>							
<i>Nephtys cirrosa</i>	77	26	38	77	13	13	51
<i>Nephtys hombergii</i>							
<i>Nototropis falcatus</i>							
<i>Ophelia borealis</i>	26	38	13	13	13		
<i>Ophiura albida</i>							
<i>Ophiura ophiura</i>			26				
<i>Perioculodes longimanus</i>							
<i>Pestarella tyrrhena</i>							
<i>Phoronis</i> spp.						13	
<i>Phyllodoce mucosa</i>							
<i>Pinnotheres pisum</i>							
<i>Poecilochaetus serpens</i>						38	

Annex 3: Box core data

Area Sample Species	QAW						
	QCS 25	QAW 1.0	QAW 2.0	QAW 3.0	QAW 4.0	QAW 5.0	QAW 6.0
<i>Pontocrates arcticus</i>		13			13		
<i>Processa modica</i>							
<i>Pseudocuma longicorne</i>							
<i>Scolelepis bonnierii</i>							
<i>Scoloplos armiger</i>	13	38	26	38	103	231	282
<i>Spio goniocephala</i>							
<i>Spiophanes bombyx</i>							
<i>Spisula</i> spp.							
<i>Spisula subtruncata</i>							
<i>Tellimya ferruginosa</i>				13			38
<i>Thia scutellata</i>							
<i>Tubularia indivisa</i>							
<i>Urothoe brevicornis</i>						13	38
<i>Urothoe poseidonis</i>							
<i>Urothoe</i> spp.							

Annex 3: Box core data

Area Sample Species	QCN							
	QCN 1	QCN 2	QCN 3.0	QCN 4	QCN 5	QCN 6	QCN 7	QCN 8
<i>Periculodes longimanus</i>								
<i>Pestarella tyrrhena</i>								
<i>Pboronis</i> spp.								
<i>Phyllococe mucosa</i>								
<i>Pinnotheres pisum</i>								
<i>Poecilochaetus serpens</i>								
<i>Pontocrates arcticus</i>				0,0038			0,0038	
<i>Processa modica</i>								
<i>Pseudocuma longicorne</i>								
<i>Scolecopsis bonnieri</i>					0,2353	0,0843		
<i>Scoloplos armiger</i>	0,0416	0,1120	0,0441	0,0133	0,0031	0,2652	0,3664	0,0033
<i>Spio goniocephala</i>		0,0051						
<i>Spiophanes bombyx</i>								
<i>Spisula</i> spp.								
<i>Spisula subtruncata</i>								
<i>Tellinomya ferruginosa</i>			0,0395	0,0191		0,0081	0,0461	
<i>Thia scutellata</i>								
<i>Tubularia indivisa</i>								
<i>Urotroche brevicornis</i>		0,0038				0,0038	0,0077	
<i>Urotroche poseidonis</i>					0,1115		0,0038	
<i>Urotroche</i> spp.			0,0038					

Annex 3: Box core data

Area Sample Species	QT							
	QCN 25	QT 1	QT 2	QT 3	QT 4	QT 5	QT 6.0	QT 7
<i>Poecilochaetus serpens</i>								
<i>Pontocrates arcticus</i>		0,0038		0,0038				0,0077
<i>Processa modica</i>						0,0985		
<i>Pseudocuma longicorne</i>								
<i>Scolelepis bonnierii</i>	0,0492							
<i>Scoloplos armiger</i>	0,1298	0,0002	0,0599	0,0297	0,0468	0,0484	0,0158	0,2119
<i>Spio goniocephala</i>			0,0014					
<i>Spiophanes bombyx</i>				0,0055				
<i>Spisula</i> spp.								
<i>Spisula subtruncata</i>								
<i>Tellinmya ferruginosa</i>	0,0304							
<i>Thia scutellata</i>								
<i>Tubularia indivisa</i>								
<i>Urotboe brevicornis</i>			0,0038				0,0038	0,0077
<i>Urotboe poseidonis</i>								
<i>Urotboe</i> spp.	0,0038							

Area	QT 8	QT 9	QT 10.0	QT 11	QT 12	QT 13	QT 14	QT 15
<i>Abludomelita obtusata</i>								
<i>Angulus fabula</i>								
<i>Aricidea minuta</i>								
<i>Aricidea spp.</i>								
<i>Asterias rubens</i>			1,4857					
<i>Bathyporeia cf. elegans</i>	0,0038	0,0038						
<i>Bathyporeia cf. nana</i>								
<i>Bathyporeia elegans</i>	0,0038	0,0038	0,0115	0,0269	0,0154	0,0577	0,0077	0,0115
<i>Bathyporeia guilliamsoniana</i>			0,0077	0,0038	0,0077	0,0038		0,0115
<i>Bathyporeia nana</i>								
<i>Bathyporeia spp.</i>						0,0038		
<i>Branchiostoma lanceolatum</i>				0,0000				
<i>Chaetozone setosa</i>			0,0076		0,0207			0,0119
<i>Chamelea striatula</i>								
<i>Diastylis cf. rugosa</i>	0,0038							
<i>Donax vittatus</i>								
<i>Echinocardium cordatum</i>	19,2820	5,8144	14,5970		5,3321		5,3321	
<i>Ensis directus</i>								
<i>Eteone foliosa</i>			0,0008					
<i>Eteone longa</i>	0,0008	0,0012	0,0201					0,0012
<i>Eunereis longissima</i>			0,0256					1,8885
<i>Euspira pulchella</i>			0,4118					
<i>Gastrosaccus spinifer</i>	0,0128							
<i>Gastrosaccus spp.</i>		0,0128						
<i>Lanice conchilega</i>	0,0595							
<i>Leucothoe incisa</i>	0,0038		0,0038					
<i>Liocarcinus cf. bolsatus</i>								
<i>Liocarcinus bolsatus</i>								
<i>Macoma balthica</i>						0,5817		
<i>Magelona filiformis</i>	0,0014		0,0045		0,0016		0,0148	0,0018
<i>Magelona johnstoni</i>		0,0098	0,0958	0,2397		0,0316		
<i>Magelona mirabilis</i>					0,0197			
<i>Magelona spp.</i>								
<i>Malmgreniella darbonxi</i>								
<i>Megaluropus agilis</i>								
<i>Monopseudocuma gilsoni</i>								
<i>Nemertea spp. A</i>	0,0410		0,0238				0,0018	
<i>Nemertea spp. B</i>								0,0164
<i>Nephtys caeca</i>								
<i>Nephtys cirrosa</i>	0,0224	0,1863	0,6818	0,0032	0,1701	0,1586	0,0708	0,0301
<i>Nephtys bombergii</i>				3,3046				
<i>Nototropis falcatus</i>						0,0038		
<i>Ophelia borealis</i>		0,0406				0,3272		
<i>Ophiura albida</i>			0,0008					
<i>Ophiura ophiura</i>								
<i>Perioculodes longimanus</i>								
<i>Pestarella tyrrhena</i>				3,0872				
<i>Phoronis spp.</i>					0,0185			
<i>Phyllodoce mucosa</i>								
<i>Pinnotheres pisum</i>						0,0120		

Annex 3: Box core data

Area	QT 25	QT 26	QT 27	QT 28	QT 29	QT 30	QT 31	QT 32
Sample								
Species								
<i>Poecilochaetus serpens</i>								
<i>Pontocrates arcticus</i>			0,0038	0,0038				
<i>Processa modica</i>								
<i>Pseudocuma longicorne</i>								
<i>Scolecopsis bonnieri</i>					0,1479			
<i>Scoloplos armiger</i>	0,0127	0,0677	0,0094	0,0277	0,0544	0,0049	0,3424	0,1346
<i>Spio gonocephala</i>								
<i>Spiophanes bombyx</i>								
<i>Spisula</i> spp.								
<i>Spisula subtruncata</i>								
<i>Tellinomya ferruginosa</i>								
<i>Thia scutellata</i>					1,0394			
<i>Tubularia indivisa</i>								
<i>Urotroche brevicornis</i>		0,0077	0,0192	0,0231	0,0038	0,0115		
<i>Urotroche poseidonis</i>								
<i>Urotroche</i> spp.					0,0038	0,0038		

Area Sample Species	QAW						
	QCS 25	QAW 1.0	QAW 2.0	QAW 3.0	QAW 4.0	QAW 5.0	QAW 6.0
<i>Abludomelita obtusata</i>							
<i>Angulus fabula</i>							
<i>Aricidea minuta</i>							
<i>Aricidea spp.</i>							
<i>Asterias rubens</i>							
<i>Bathyporeia cf. elegans</i>							
<i>Bathyporeia cf. nana</i>							0,0038
<i>Bathyporeia elegans</i>	0,0038	0,0231	0,0154	0,0077	0,0038	0,0115	0,0077
<i>Bathyporeia guilliamsoniana</i>	0,0038					0,0231	
<i>Bathyporeia nana</i>							
<i>Bathyporeia spp.</i>			0,0038				
<i>Branchiostoma lanceolatum</i>							
<i>Chaetozone setosa</i>			0,0103		0,0207		0,0008
<i>Chamelea striatula</i>							
<i>Diastylis cf. rugosa</i>							
<i>Donax vittatus</i>							
<i>Echinocardium cordatum</i>	1,1640			18,3102		5,5386	5,3321
<i>Ensis directus</i>			15,3977				5,6658
<i>Eteone foliosa</i>							
<i>Eteone longa</i>		0,0016	0,0014		0,0008	0,0119	
<i>Eunereis longissima</i>							
<i>Euspira pulchella</i>							
<i>Gastrosaccus spinifer</i>		0,0128					
<i>Gastrosaccus spp.</i>							
<i>Lanice conchilega</i>							
<i>Leucothoe incisa</i>							
<i>Liocarcinus cf. bolsatus</i>							
<i>Liocarcinus bolsatus</i>							
<i>Macoma balthica</i>		0,5616			0,0682		
<i>Magelona filiformis</i>			0,0131		0,0023	0,0131	
<i>Magelona johnstoni</i>				0,0082		0,0310	
<i>Magelona mirabilis</i>	0,0275				0,0806		
<i>Magelona spp.</i>							
<i>Malmgreniella darbonxi</i>							
<i>Megaluropus agilis</i>					0,0038		
<i>Monopseudocuma gilsoni</i>							0,0038
<i>Nemertea spp. A</i>		0,0363	0,0049		0,0111	0,0088	
<i>Nemertea spp. B</i>					0,0045		
<i>Nephtys caeca</i>							
<i>Nephtys cirrosa</i>	0,8523	0,2086	0,1723	0,2652	0,2948	0,2062	0,4267
<i>Nephtys bombergii</i>							
<i>Nototropis falcatus</i>							
<i>Ophelia borealis</i>	0,0008	0,1403	0,0014	0,0630	0,0012		
<i>Ophiura albida</i>							
<i>Ophiura ophiura</i>			2,7223				
<i>Perioculodes longimanus</i>							
<i>Pestarella tyrrhena</i>							
<i>Phoronis spp.</i>						0,0088	
<i>Phyllococe mucosa</i>							
<i>Pinnotheres pisum</i>							

Annex 3: Box core data

Area Sample Species	QAW						
	QCS 25	QAW 1.0	QAW 2.0	QAW 3.0	QAW 4.0	QAW 5.0	QAW 6.0
<i>Poecilochaetus serpens</i>						0,0821	
<i>Pontocrates arcticus</i>		0,0038			0,0038		
<i>Processa modica</i>							
<i>Pseudocuma longicorne</i>							
<i>Scolecopsis bonnieri</i>							
<i>Scoloplos armiger</i>	0,2306	0,2314	0,0952	0,0090	0,1305	0,1906	0,3136
<i>Spio gonocephala</i>							
<i>Spiophanes bombyx</i>							
<i>Spisula</i> spp.							
<i>Spisula subtruncata</i>							
<i>Tellinomya ferruginosa</i>				0,0307			0,0987
<i>Thia scutellata</i>							
<i>Tubularia indivisa</i>							
<i>Urotroche brevicornis</i>						0,0038	0,0115
<i>Urotroche poseidonis</i>							
<i>Urotroche</i> spp.							

Annex 4: Dredge sample data

Area Sample Species	QCN							
	QCN 1	QCN 2	QCN 3	QCN 4	QCN 5	QCN 6	QCN 7	QCN 8
<i>Paguridae</i> spp.	1	1				1		
<i>Pagurus bernhardus</i>	5		2					1
<i>Philocheras trispinosus</i>	1			1	2	3		
<i>Phyllodoce groenlandica</i>								
<i>Phyllodoce lineata</i>								
<i>Pinnotheres pisum</i>								
<i>Pleuronectes platessa</i>	1						1	
<i>Pomatoschistus</i> spp.	22	3	2		3	2	5	5
<i>Processa</i> spp.		2		4				
<i>Scolelepis bonnierii</i>								
<i>Scolelepis</i> spp.				1				
<i>Scoloplos armiger</i>						2		
<i>Sigalion mathildae</i>							1	1
<i>Solea solea</i>								
<i>Spio</i> spp.						3		
<i>Spisula</i> spp.	9	18	19	14	11	15	1	4
<i>Striarca lactea</i>								
<i>Taurulus bubalis</i>								
<i>Tellinmya ferruginosa</i>								
<i>Thia scutellata</i>		6		1	5	3	2	6

Annex 4: Dredge sample data

Area Sample Species	QT							
	QCN 9	QT 1	QT 2	QT 3	QT 4	QT 5	QT 6	QT 7
<i>Phyllodoce lineata</i>		1						
<i>Pinnotheres pisum</i>				2	1			
<i>Pleuronectes platessa</i>								
<i>Pomatoschistus</i> spp.	6	1	5	1		1		6
<i>Processa</i> spp.			1	1	1	1		
<i>Scolecipis bonnieri</i>								
<i>Scolecipis</i> spp.								
<i>Scoloplos armiger</i>		1						
<i>Sigalion matbildae</i>								
<i>Solea solea</i>				1				
<i>Spio</i> spp.								
<i>Spisula</i> spp.	11	43	3	12	22	6	9	7
<i>Striarca lactea</i>								
<i>Taurulus bubalis</i>								
<i>Tellimya ferruginosa</i>								
<i>Tibia scutellata</i>	4	3	2	3	1		4	

Annex 4: Dredge sample data

Area	QT 8	QT 9	QT 10	QT 11	QT 12	QT 13	QT 14	QT 15
Sample								
Species								
<i>Ammodytes</i> spp.					1			1
<i>Angulus fabula</i>								
<i>Angulus tenuis</i>								
<i>Armoglossus laterna</i>								
<i>Asterias rubens</i>	7	10			9	1	1	3
<i>Buglossidium luteum</i>	5	8		3	7	2	2	
<i>Callionymus lyra</i>				1		1		
<i>Callionymus reticulatus</i>		1		1				
<i>Chamelea</i> spp.	7	4	1	2	5	1	3	5
<i>Corystes cassivelaunus</i>	1	2						
<i>Crangon almanni</i>							1	
<i>Crangon crangon</i>	1	6	11	8	4	3	4	5
<i>Donax vittatus</i>	7	1	3	7	1	7	3	11
<i>Echüchthys vipera</i>								
<i>Echinocardium cordatum</i>	171	62	13	30	124	112	26	116
<i>Ensis directus</i>								
<i>Ensis ensis</i>	2	2	23	2		4	3	6
<i>Ensis</i> spp.								
<i>Eteone foliosa</i>				2			1	
<i>Eunereis longissima</i>					9	2		
<i>Euspira pulchella</i>	1	9		1	15	4	2	
<i>Gastrosaccus spinifer</i>					1			
<i>Gymnammodytes semisumatus</i>				1				
<i>Hyperoplus lanceolatus</i>	2		1	1	2		1	
<i>Hyperoplus</i> spp.								
<i>Lanice conchilega</i>					1		2	
<i>Limanda limanda</i>			1	2				1
<i>Liocarcinus holsatus</i>	1			2	1	2		
<i>Liocarcinus marmoreus</i>								
<i>Liocarcinus</i> spp.							1	
<i>Lutraria</i> spp.								
<i>Macropodia rostrata</i>								
<i>Nemertea</i> spp. A								
<i>Nemertea</i> spp. B	1							
<i>Nemertea</i> spp. C								
<i>Nephtys assimilis</i>								
<i>Nephtys caeca</i>								
<i>Nephtys cirrosa</i>	8			3		6		
<i>Nephtys hombergii</i>	1			1				
<i>Nephtys</i> spp.	3	3	14		14		9	9
<i>Nereidae</i> spp.		1			1			
<i>Ophelia borealis</i>			1	1	2	2	3	4
<i>Ophiura albida</i>	16	14	5	3	31	15	12	4
<i>Ophiura ophiura</i>	43	83	9	19	46	30	10	8
<i>Ophiuridae</i> spp.								
<i>Paguridae</i> spp.	1			1				
<i>Pagurus bernhardus</i>	2	2						
<i>Philocheras trispinosus</i>			2				2	1
<i>Phyllodoce groenlandica</i>			1					

Annex 4: Dredge sample data

Area	QT 8	QT 9	QT 10	QT 11	QT 12	QT 13	QT 14	QT 15
Sample								
Species								
<i>Phyllodoce lineata</i>								
<i>Pinnotheres pisum</i>								
<i>Pleuronectes platessa</i>			1				2	1
<i>Pomatoschistus</i> spp.	1	4	2		1		1	
<i>Processa</i> spp.		2			6			1
<i>Scolecopsis bonnieri</i>							1	
<i>Scolecopsis</i> spp.	1		1					
<i>Scoloplos armiger</i>	1	1						2
<i>Sigalion matbildae</i>								
<i>Solea solea</i>		1			1	1		
<i>Spio</i> spp.								
<i>Spisula</i> spp.	45	19	7	19	30	15	11	16
<i>Striarca lactea</i>								
<i>Taurulus bubalis</i>			1					
<i>Tellinomya ferruginosa</i>	1							
<i>Thia scutellata</i>	3	3	6	2	5	6	1	8

Annex 4: Dredge sample data

Area Sample Species	QCS							
	QCS 1	QCS 2	QCS 3	QCS 4	QCS 5	QCS 6	QCS 7	QCS 8
<i>Phyllodoce lineata</i>								
<i>Pinnotheres pisum</i>				1	1			
<i>Pleuronectes platessa</i>								
<i>Pomatoschistus</i> spp.		1		1	1			2
<i>Processa</i> spp.	4	1		2	2		6	2
<i>Scolecopsis bonnieri</i>								
<i>Scolecopsis</i> spp.				1				
<i>Scoloplos armiger</i>			1	1	3		2	1
<i>Sigalion matbildae</i>								
<i>Solea solea</i>								
<i>Spio</i> spp.		1	4					
<i>Spisula</i> spp.	19	21	16	37	32	20	14	18
<i>Striarca lactea</i>	1							
<i>Taurulus bubalis</i>								
<i>Tellinomya ferruginosa</i>								
<i>Thia scutellata</i>	16	2	5	20	9	2	12	3

Annex 4: Dredge sample data

Area Sample Species	QAW						
	QCS 9	QAW 1	QAW 2	QAW 3	QAW 4	QAW 5	QAW 6
<i>Ammodytes</i> spp.					2		
<i>Angulus fabula</i>				2	1		
<i>Angulus tenuis</i>							
<i>Armoglossus laterna</i>							
<i>Asterias rubens</i>		1	3	3		1	1
<i>Buglossidium luteum</i>	4	4	1	5	3	8	2
<i>Callionymus lyra</i>			1				
<i>Callionymus reticulatus</i>							
<i>Chamelea</i> spp.	2			9		18	
<i>Corystes cassivelaunus</i>			1				1
<i>Crangon almanni</i>							1
<i>Crangon crangon</i>	6	17	17	9	9	7	7
<i>Donax vittatus</i>		4	3	9	14	13	13
<i>Echüchthys vipera</i>		1					
<i>Echinocardium cordatum</i>	17	28	8	27	33	79	44
<i>Ensis directus</i>							
<i>Ensis ensis</i>	2			4	1	1	
<i>Ensis</i> spp.							
<i>Eteone foliosa</i>							
<i>Eunereis longissima</i>				3		3	
<i>Euspira pulchella</i>	2		1	1	4	2	
<i>Gastrosaccus spinifer</i>							
<i>Gymnammodytes semisumatus</i>							
<i>Hyperoplus lanceolatus</i>		1	1	1	2		1
<i>Hyperoplus</i> spp.							
<i>Lanice conchilega</i>							
<i>Limanda limanda</i>							
<i>Liocarcinus holsatus</i>				3		1	1
<i>Liocarcinus marmoreus</i>	1						
<i>Liocarcinus</i> spp.							
<i>Lutraria</i> spp.			1			1	
<i>Macropodia rostrata</i>							
<i>Nemertea</i> spp. A							
<i>Nemertea</i> spp. B						1	1
<i>Nemertea</i> spp. C	1						
<i>Nephtys assimilis</i>				1			
<i>Nephtys caeca</i>							
<i>Nephtys cirrosa</i>			2			6	2
<i>Nephtys hombergii</i>						1	
<i>Nephtys</i> spp.	21	12		6	6	9	3
<i>Nereidae</i> spp.							
<i>Ophelia borealis</i>	17	5	1	1	2	1	
<i>Ophiura albida</i>	10	25	60	51	55	36	35
<i>Ophiura ophiura</i>	6	41	56	38	49	35	48
<i>Ophiuridae</i> spp.		1	2	1	1		2
<i>Paguridae</i> spp.		2	3	2	3	2	1
<i>Pagurus bernhardus</i>				1			
<i>Philocheras trispinosus</i>	2	1	2	1	3	1	3
<i>Phyllodoce groenlandica</i>							

Annex 4: Dredge sample data

Area Sample Species	QAW						
	QCS 9	QAW 1	QAW 2	QAW 3	QAW 4	QAW 5	QAW 6
<i>Phyllodoce lineata</i>							
<i>Pinnotheres pisum</i>			1				
<i>Pleuronectes platessa</i>	1			1			1
<i>Pomatoschistus</i> spp.		1	3	1	1		2
<i>Processa</i> spp.	6		1	2		3	3
<i>Scolelepis bonnierii</i>							
<i>Scolelepis</i> spp.		1					
<i>Scoloplos armiger</i>	1			1			
<i>Sigalion matbildae</i>							
<i>Solea solea</i>							
<i>Spio</i> spp.			1				
<i>Spisula</i> spp.	23	27	33	41	16	47	12
<i>Striarca lactea</i>							
<i>Taurulus bubalis</i>							
<i>Tellimya ferruginosa</i>							
<i>Thia scutellata</i>	10	4	1		4	2	6

Annex 4: Dredge sample data

Area Sample Species	QCN QCN 1	QCN 2	QCN 3	QCN 4	QCN 5	QCN 6	QCN 7	QCN 8
<i>Paguridae</i> spp.	0,001	0,001				0,001		
<i>Pagurus bernhardus</i>	1,212		0,242					0,242
<i>Philocheras trispinosus</i>	0,028			0,053	0,079	0,122		
<i>Phyllodoce groenlandica</i>								
<i>Phyllodoce lineata</i>								
<i>Pinnotheres pisum</i>								
<i>Pleuronectes platessa</i>	2,600						36,000	
<i>Pomatoschistus</i> spp.	3,800	0,400	0,272		0,800	0,400	1,000	0,600
<i>Processa</i> spp.		0,001		0,001				
<i>Scolecipis bonnieri</i>								
<i>Scolecipis</i> spp.				0,010				
<i>Scoloplos armiger</i>						0,020		
<i>Sigalion matbildae</i>							0,010	0,010
<i>Solea solea</i>								
<i>Spio</i> spp.						0,030		
<i>Spisula</i> spp.	7,273	2,000	4,909	4,182	4,727	9,455	0,182	1,636
<i>Striarca lactea</i>								
<i>Taurulus bubalis</i>								
<i>Tellimya ferruginosa</i>								
<i>Thia scutellata</i>		0,545		0,063	0,303	0,364	0,061	0,727

Annex 4: Dredge sample data

Area Sample Species	QCN 9	QT QT 1	QT 2	QT 3	QT 4	QT 5	QT 6	QT 7
<i>Phyllodoce lineata</i>		0,010						
<i>Pinnotheres pisum</i>				0,001	0,001			
<i>Pleuronectes platessa</i>								
<i>Pomatoschistus</i> spp.	1,200	0,200	0,400	0,400		0,086		0,800
<i>Processa</i> spp.			0,001	0,001	0,001	0,001		
<i>Scolecopsis bonnieri</i>								
<i>Scolecopsis</i> spp.								
<i>Scoloplos armiger</i>		0,010						
<i>Sigalion matbildae</i>								
<i>Solea solea</i>				25,800				
<i>Spio</i> spp.								
<i>Spisula</i> spp.	1,636	2,909	2,727	1,818	2,545	1,818	2,000	2,182
<i>Striarca lactea</i>								
<i>Taurulus bubalis</i>								
<i>Tellimya ferruginosa</i>								
<i>Thia scutellata</i>	0,242	0,121	0,117	0,121	0,063		0,121	

Annex 4: Dredge sample data

Area Sample Species	QT 8	QT 9	QT 10	QT 11	QT 12	QT 13	QT 14	QT 15
<i>Ammodytes</i> spp.					1,400			0,010
<i>Angulus fabula</i>								
<i>Angulus tenuis</i>								
<i>Arnoglossus laterna</i>								
<i>Asterias rubens</i>	2,728	3,720			1,488	0,200	0,200	4,340
<i>Buglossidium luteum</i>	8,000	10,000		2,800	7,600	3,400	2,400	
<i>Callionymus lyra</i>				0,600		0,400		
<i>Callionymus reticulatus</i>		0,200		0,200				
<i>Chamelea</i> spp.	4,909	1,455	0,909	2,364	2,727	1,455	2,727	5,636
<i>Corystes cassivelaunus</i>	0,667	1,394						
<i>Crangon almanni</i>							0,165	
<i>Crangon crangon</i>	0,061	0,424	1,091	0,848	0,061	0,485	0,606	0,424
<i>Donax vittatus</i>	2,364	0,364	1,818	3,455	0,182	2,364	1,273	5,636
<i>Echiichthys vipera</i>								
<i>Echinocardium cordatum</i>	188,930	75,740	16,555	38,500	120,505	156,485	29,155	93,555
<i>Ensis directus</i>								
<i>Ensis ensis</i>	0,727	1,273	10,182	1,636		2,727	5,818	4,364
<i>Ensis</i> spp.								
<i>Eteone foliosa</i>				0,020			0,010	
<i>Eunereis longissima</i>					0,090	0,020		
<i>Euspira pulchella</i>	0,025	0,202		0,025	0,411	0,102	0,052	
<i>Gastrosaccus spinifer</i>					0,001			
<i>Gymnammodytes semisumatus</i>				0,400				
<i>Hyperoplus lanceolatus</i>	0,010		3,200	0,274	3,400		0,010	
<i>Hyperoplus</i> spp.								
<i>Lanice conchilega</i>					0,010		0,020	
<i>Limanda limanda</i>			0,010	8,200				0,010
<i>Liocarcinus holsatus</i>	0,606			0,848	0,424	1,394		
<i>Liocarcinus marmoreus</i>								
<i>Liocarcinus</i> spp.							0,036	
<i>Lutraria</i> spp.								
<i>Macropodia rostrata</i>								
<i>Nemertea</i> spp. A								
<i>Nemertea</i> spp. B	0,010							
<i>Nemertea</i> spp. C								
<i>Nephtys assimilis</i>								
<i>Nephtys caeca</i>								
<i>Nephtys cirrosa</i>	0,080			0,030		0,060		
<i>Nephtys hombergii</i>	0,010			0,010				
<i>Nephtys</i> spp.	0,030	0,030	0,140		0,140		0,090	0,090
<i>Nereidae</i> spp.		0,010			0,010			
<i>Ophelia borealis</i>			0,010	0,010	0,020	0,020	0,030	0,040
<i>Ophiura albida</i>	0,462	0,923	0,768	0,165	1,538	0,873	0,462	0,383
<i>Ophiura ophiura</i>	2,615	7,077	2,615	3,538	2,154	2,923	1,692	2,615
<i>Ophiuridae</i> spp.								
<i>Paguridae</i> spp.	0,001			0,001				
<i>Pagurus bernhardus</i>	1,091	1,939						
<i>Philocheras trispinosus</i>			0,083				0,100	0,059
<i>Phyllodoce groenlandica</i>			0,010					

Annex 4: Dredge sample data

Area Sample Species	QT 8	QT 9	QT 10	QT 11	QT 12	QT 13	QT 14	QT 15
<i>Phyllodoce lineata</i>								
<i>Pinnotheres pisum</i>								
<i>Pleuronectes platessa</i>			5,800				8,000	15,600
<i>Pomatoschistus</i> spp.	0,204	1,600	0,800		0,136		0,400	
<i>Processa</i> spp.		0,001			0,001			0,001
<i>Scolecopsis bonnieri</i>							0,010	
<i>Scolecopsis</i> spp.	0,010		0,010					
<i>Scoloplos armiger</i>	0,010	0,010						0,020
<i>Sigalion mathildae</i>								
<i>Solea solea</i>		16,400			18,200	23,000		
<i>Spio</i> spp.								
<i>Spisula</i> spp.	5,273	1,273	3,091	4,727	1,273	2,364	4,000	6,909
<i>Striarca lactea</i>								
<i>Taurulus bubalis</i>			4,000					
<i>Tellimya ferruginosa</i>	0,003							
<i>Thia scutellata</i>	0,182	0,182	0,242	0,063	0,121	0,364	0,049	0,303

Annex 4: Dredge sample data

Area Sample Species	QCS QCS 1	QCS 2	QCS 3	QCS 4	QCS 5	QCS 6	QCS 7	QCS 8
<i>Phyllodoce lineata</i>								
<i>Pinnotheres pisum</i>				0,001	0,001			
<i>Pleuronectes platessa</i>								
<i>Pomatoschistus</i> spp.		0,200		0,204	0,136			0,200
<i>Processa</i> spp.	0,001	0,001		0,001	0,001		0,001	0,001
<i>Scolecopsis bonnieri</i>								
<i>Scolecopsis</i> spp.				0,010				
<i>Scoloplos armiger</i>			0,010	0,010	0,030		0,010	0,010
<i>Sigalion matbildae</i>								
<i>Solea solea</i>								
<i>Spio</i> spp.		0,010	0,040					
<i>Spisula</i> spp.	4,000	13,636	6,000	7,818	5,273	12,545	5,818	5,273
<i>Striarca lactea</i>	0,040							
<i>Taurulus bubalis</i>								
<i>Tellimya ferruginosa</i>								
<i>Thia scutellata</i>	0,606	0,242	0,242	1,212	0,303	0,121	0,727	0,242

Annex 4: Dredge sample data

Area Sample Species	QCS 9	QAW QAW 1	QAW 2	QAW 3	QAW 4	QAW 5	QAW 6
<i>Ammodytes</i> spp.					1,400		
<i>Angulus fabula</i>				0,062	0,101		
<i>Angulus tenuis</i>							
<i>Arnoglossus laterna</i>							
<i>Asterias rubens</i>		0,200	0,248	1,860		0,200	0,200
<i>Buglossidium luteum</i>	2,600	3,600	1,200	6,400	2,600	9,400	1,000
<i>Callionymus lyra</i>			7,600				
<i>Callionymus reticulatus</i>							
<i>Chamelea</i> spp.	0,020			1,636		1,455	
<i>Corystes cassivelaunus</i>			0,606				0,606
<i>Crangon almanni</i>							0,206
<i>Crangon crangon</i>	0,303	1,636	1,576	0,727	1,030	0,485	0,606
<i>Donax vittatus</i>		2,545	1,273	3,455	6,545	3,636	6,545
<i>Echiichthys vipera</i>		0,086					
<i>Echinocardium cordatum</i>	19,145	31,885	9,240	31,640	34,930	87,710	54,845
<i>Ensis directus</i>							
<i>Ensis ensis</i>	2,000			2,182	0,545	0,364	
<i>Ensis</i> spp.							
<i>Eteone foliosa</i>							
<i>Eunereis longissima</i>				0,030		0,030	
<i>Euspira pulchella</i>	0,032		0,025	0,025	0,100	0,050	
<i>Gastrosaccus spinifer</i>							
<i>Gymnammodytes semisumatus</i>							
<i>Hyperoplus lanceolatus</i>		3,400	0,200	0,400	0,200		0,010
<i>Hyperoplus</i> spp.							
<i>Lanice conchilega</i>							
<i>Limanda limanda</i>							
<i>Liocarcinus holsatus</i>				2,061		0,788	0,545
<i>Liocarcinus marmoreus</i>	0,606						
<i>Liocarcinus</i> spp.							
<i>Lutraria</i> spp.			0,010			0,010	
<i>Macropodia rostrata</i>							
<i>Nemertea</i> spp. A							
<i>Nemertea</i> spp. B						0,010	0,010
<i>Nemertea</i> spp. C	0,010						
<i>Nephtys assimilis</i>				0,010			
<i>Nephtys caeca</i>							
<i>Nephtys cirrosa</i>			0,020			0,060	0,020
<i>Nephtys hombergii</i>						0,010	
<i>Nephtys</i> spp.	0,210	0,120		0,060	0,060	0,090	0,030
<i>Nereidae</i> spp.							
<i>Ophelia borealis</i>	0,170	0,050	0,010	0,010	0,020	0,010	
<i>Ophiura albida</i>	1,231	1,846	4,000	2,769	2,308	3,538	1,385
<i>Ophiura ophiura</i>	3,538	13,077	5,538	7,385	5,077	12,615	6,154
<i>Ophiuridae</i> spp.		0,046	0,119	0,046	0,046		0,146
<i>Paguridae</i> spp.		0,002	0,003	0,002	0,003	0,002	0,001
<i>Pagurus bernhardus</i>				0,143			
<i>Philocheras trispinosus</i>	0,051	0,032	0,078	0,036	0,120	0,047	0,136
<i>Phyllodoce groenlandica</i>							

Annex 4: Dredge sample data

Area Sample Species	QAW						
	QCS 9	QAW 1	QAW 2	QAW 3	QAW 4	QAW 5	QAW 6
<i>Phyllodoce lineata</i>							
<i>Pinnotheres pisum</i>			0,001				
<i>Pleuronectes platessa</i>	34,000			10,000			3,400
<i>Pomatoschistus</i> spp.		0,086	0,600	0,086	0,136		0,200
<i>Processa</i> spp.	0,001		0,001	0,001		0,001	0,001
<i>Scolecopsis bonnierii</i>							
<i>Scolecopsis</i> spp.		0,010					
<i>Scoloplos armiger</i>	0,010			0,010			
<i>Sigalion mathildae</i>							
<i>Solea solea</i>							
<i>Spio</i> spp.			0,010				
<i>Spisula</i> spp.	7,091	7,818	5,273	5,273	2,182	6,364	5,273
<i>Striarca lactea</i>							
<i>Taurulus bubalis</i>							
<i>Tellimya ferruginosa</i>							
<i>Thia scutellata</i>	0,545	0,121	0,028		0,182	0,077	0,061

Annex 5: Physical variables

Sample	Median particle size (µm)	Clay (%)	Sand (%)	Organic matter (%)
QCN 1	278,916	5,325	94,675	0,8463
QCN 2	329,572	0	100	0,5008
QCN 3.0	247,885	0	100	0,4983
QCN 4	246,031	4,824	95,176	0,4892
QCN 5	231,152	5,501	94,499	0,5030
QCN 6	296,385	0	100	0,4362
QCN 7	254,28	0	100	0,5075
QCN 8	251,575	0	100	0,5517
QCN 9	238,465	4,744	95,256	0,6510
QCN 10	231,559	5,399	94,601	0,8660
QCN 11 .0	297,264	0	100	0,6462
QCN 12	261,22	1,233	98,767	0,7360
QCN 13.0	255,157	0	100	0,4896
QCN 13.4	266,077	4,187	95,813	0,4862
QCN 14	242,843	4,592	95,408	0,5278
QCN 15.0	237,937	0	100	0,5841
QCN 16	364,789	0,685	99,315	0,5677
QCN 17	247,676	0	100	0,5898
QCN 18	269,246	0	100	0,6258
QCN 19	240,135	0	100	0,2773
QCN 20	237,764	0	100	0,4818
QCN 21	267,293	0	100	0,5473
QCN 22	277,418	0	100	0,6332
QCN 23.0	276,598	0	100	0,4253
QCN 24	235,316	0	100	0,3306
QCN 25	246,502	0	100	1,0046
QT 1	295,089	0	100	0,4423
QT 2.0	293,44	1,862	98,138	0,5156
QT 3	291,718	0	100	0,4701
QT 4	291,354	0	100	0,4413
QT 5	276,275	0	100	0,3988
QT 6.0	270,426	0	100	0,4176
QT 6.2	284,845	0	100	0,4084
QT 7	262,408	0	100	0,4736
QT 8	565,248	0	200	1,0295
QT 10.0	243,709	0	100	0,5215
QT 11	252,101	0	100	0,5715
QT 12	235,769	0	100	0,5142
QT 13	260,115	0	100	0,4996
QT 14	286,254	0	100	0,4898
QT 15	251,883	0	100	0,4974
QT 16	249,551	0	100	0,5122
QT 18.0	264,103	0	100	0,5751
QT 19	272,694	0	100	0,4642
QT 20	255,493	0	100	0,5622

Annex 5: Physical variables

Sample	Median particle size (µm)	Clay (%)	Sand (%)	Organic matter (%)
QT 21.0	246,346	0	100	0,5131
QT 22	270,658	0	100	0,4905
QT 23	278,979	0	100	0,5474
QT 24	277,694	0	100	0,5632
QT 25	270,995	0	100	0,7320
QT 26	275,498	0	100	0,6535
QT 27	286,873	0	100	0,5538
QT 28	282,203	0	100	0,4878
QT 29	284,243	0	100	0,5058
QT 30	283,8	0	100	0,5104
QT 31	314,711	0	100	0,5812
QT 32	295,293	0	100	0,5148
QT 33.0	294,817	0	100	0,5315
QT 34	283,53	0	100	0,4618
QT 35	285,693	0	100	0,5362
QT 36	270,324	0	100	0,5042
QT 37	291,775	0	100	0,4830
QT 38	290,063	0	100	0,4759
QT 39	258,86	4,86	95,14	0,4481
QT 40.0	282,403	0	100	0,4911
QT 41	285,097	0	100	0,4500
QT 42	302,284	0	100	0,5474
QT 43	288,934	0	100	0,5285
QT 44.0	292,669	0	100	0,4308
QT 46	296,755	0	100	0,4112
QT 47	272,967	0	100	0,4042
QT 48	274,637	7,93	92,07	0,4910
QCS 1	300,96	0	100	0,4785
QCS 2	318,4	0	100	0,4110
QCS 3.0	298,123	0	100	0,8074
QCS 4	315,406	0	100	0,4587
QCS 5	331,135	0	100	0,5082
QCS 6	519,987	0	200	0,9372
QCS 7	211,984	0	100	0,7396
QCS 7	295,94	0	100	0,5348
QCS 8	302,258	1,435	98,565	0,4715
QCS 9	301,745	0	100	0,4236
QCS 10	284,073	0	100	0,4699
QCS 11 .0	310,567	0	100	0,4287
QCS 12	301,822	0	100	0,4117
QCS 13 .0	276,168	1,311	98,689	0,4136
QCS 14	298,439	0	100	0,4307
QCS 15.0	301,763	0	100	0,4749
QCS 15.1	294,431	0	100	0,5895
QCS 16	345,725	0	100	0,9457
QCS 17	311,113	0	100	0,4586
QCS 18	325,551	0	100	0,5032
QCS 19	317,086	0	100	0,3385
QCS 20	322,99	1,62	98,38	0,4266
QCS 21	327,466	0	100	0,3733
QCS 22	305,646	0	100	0,3553
QCS 23.0	305,226	0	100	0,4632
QCS 24	311,84	0	100	0,4116
QCS 25	336,115	0	100	0,4431

Annex 5: Physical variables

Sample	Median particle size (μm)	Clay (%)	Sand (%)	Organic matter (%)
QAW 1.0	287,68	0	100	0,7909
QAW 2.0	276,745	0	100	0,4711
QAW 3.0	283,44	0	100	0,4391
QAW 4.0	278,165	0	100	0,5133
QAW 4.1	280,085	0	100	0,9185
QAW 5.0	270,419	0	100	0,5068
QAW 6.0	272,934	0	100	0,4926

