VERTICAL DISTRIBUTION OF MEIOFAUNA IN SEDI-MENTS FROM CONTRASTING SITES IN THE ADRIATIC SEA: CLUES TO THE ROLE OF ABIOTIC VERSUS BIOTIC CONTROL

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ABSTRACT

The vertical profiles of dominant metazoan meiobenthic taxa were studied at four contrasting sites in the Adriatic Sea in relation to macrofaunal activity and oxygen penetration depth in the sediment. Vertical profiles of copepods were related to the oxygen penetration depth and showed peak densities in the upper cm or deeper in sediments with a broader oxic zone. In contrast, nematodes penetrated much deeper into the sediment and no direct relationship with oxygen penetration depth was evident. Nematode penetration depths were related to or reflected the differences in macrofaunal activity or mixing at the four sites which appears to be an important factor governing the subsurface activity of nematodes.

Keywords: Adriatic Sea, meiobenthos, nematodes, vertical distribution, oxygenation, bioturbation.

INTRODUCTION

Benthic meiofaunal communities are known to be influenced by a complex of interacting factors. The properties of the seabed are most commonly regarded as the principal factors controlling the composition and abundance of the benthos (Gray 1981), but it has also been recognized that food supply may be as important in determining benthic community structure (e.g. Josefson 1987; Grebmeier et al. 1988; Graf 1992; Rosenberg 1995). A positive correlation between abundance or biomass of benthic meiofauna and food availability has been demonstrated (e.g. Elmgren 1978; Thiel 1983; Pfannkuche 1985; Tietjen et al. 1989; Soetaert et al. 1991; Piepenburg et al. 1997; Cook et al. 2000), indicating the importance of food availability as a factor controlling meiofaunal populations. However in more productive areas and within a narrow water depth range, this relationship may not be straightforward (e.g. Widbom 1988; Soetaert et al. 1997), and the positive effect of an increased food supply may be counteracted by negative biological interactions with macrofauna, ciliates and protozoans (e.g. Widbom & Elmgren 1988; Moodley et al. 1998).

Previous studies of the meiobenthos in the Adriatic Sea were restricted to the northern basin (Vidakovic 1988; Tahey et al. 1996; Moodley et al. 1998). In this study, we examine the metazoan meiobenthos in sediments from the northern and middle basins using two proxies for food availability: the chlorophyll *a* and organic carbon content in the surface sediment.

Meiofauna commonly occur infaunally and, among the dominant metazoan taxa, nematodes are the principal group that penetrate deep into the sediment (e.g. Yingst 1985; Fleeger et al. 1995; Vanreusel et al. 1995). Factors promoting vertical gradients in the sediment are considered to be food availability (e.g. Thiel 1983; Lambshead et al. 1995; Soetaert et al. 1997; Steyaert et al. 1999); oxygenation of the sediment (Shirayama & Horikoshi 1982), a combination of these factors (Shiravama 1984; Vanreusel et al. 1995), or increased disturbance near the surface (Carman et al. 1987). Macrofaunal activity plays a prominent role in structuring or altering the properties of the sediment column but there are only a few studies of meiobenthos in combination with macrofauna or mixing data (Lambshead et al. 1995).

Station	BW	BW	BW	Oxygen	Median	% Silt	POC	Chlorophyll a	Macrofauna(0-20 cm)	
Number	Temp (°C)	Salinity	Oxygen (µM)	Penetration Depth (mm)	Grain Size (µm)	(< 63 µm)	$(g m^2)$	$(mg m^2)$	Abundance (n m ⁻²)	Biomass (gWW m ⁻²)
11	11.5	36.2	232.2	< 2	14.0	93	70.4 ± 4.6	172.2 ± 43.1	4093±141	498±122.7
2	11.0	37.4	221.8	< 2	10.2	99	47.8 ± 2.3	14.1 ± 2.0	7336±1517	115 ± 6.0
4	11.6	38.2	244.6	11.7	8.6	100	36.1 ± 1.6	1.7 ± 0.1	2469±106	14 ± 2.5
5	12.8	38.6	199.3	73.1	6.5	100	9.6 ± 0.2	nd	564±0	2 ± 0.4

Table 1. Bottom water (BW) and sediment parameters (0-1 cm), oxygen penetration depth and macrofauna data (average \pm SD). nd = not detected.

Table 2. Meiobenthos densities (average \pm SD) and average contribution of dominant taxa.

Station Number	Total 0-10 cm (n 10 cm ⁻²)	% Nematoda	% Copepoda + nauplii	Total 0-5 cm (n 10 cm ⁻²)	% Nematoda	% Copepoda + nauplii
11	4001 ± 481	97	1	3095 ± 356	96	2
2	1461 ± 143	88	6	1417 ± 157	88	6
4	821 ± 139	66	18	680 ± 97	62	22
5	146 ± 23	75	15	138 ± 24	77	14

In this study, we examine vertical distributional patterns of dominant metazoan meiobenthic taxa in relation to the thickness of the oxic layer and sediment mixing (as reflected by chlorophyll a and macrofauna profiles) at four contrasting sites (15 to 1039 meters water depth). Both macrofauna and chlorophyll aprofiles provide insight into mixing activity. Because chlorophyll a is highly degradable, deep penetration cannot be a consequence of mere sedimentation so that it reflects mixing or active subduction by organisms. Macrofauna biomass and abundance profiles provide direct information on subsurface activity.

RESULTS

Fine-grained sediment (muddy silt) was found at all stations (Table 1) with very limited variation downcore (data not shown). Bottom water temperature and salinity were comparable between stations (Table 1). The bottom waters were well oxygenated at all four sites but there were large differences in oxygen penetration depth. Penetration was shallowest at the coastal stations (< 2 mm) and deepest at the deep-sea site (73 mm, Table 1). No free sulphide was detected at any of the stations.

Both proxies for food availability revealed a clear gradient from shallow to deep water with minimum values at the deepest station (station 5, 1039 m water depth, Table 1).

Nematodes and copepods together made up 84 – 98 % to the metazoan meiofaunal community (Table 2). Other common, but not abundant, meiobenthic taxa include Polychaeta, Ostracoda, Tanaidacea, Kinorhyncha and Turbellaria. Both total meiofauna and nematode den-

 Table 3. List of the 10 dominant nematode genera found in the upper 5 cm and their relative abundance. * = Family.

Station 11	%	Station 2	%	Station 4	%	Station 5	%
Sabatieria	81.3	Parodontophora	12.6	Dorylaimopsis	11.2	Monhysteridae*	15.0
Terschellingia	8.0	Halalaimus	12.1	Daptonema	9.8	Chromadora	13.3
Parodontophora	2.8	Microlaimus	11.2	Monhysteridae*	9.2	Halalaimus	11.7
Daptonema	2.0	Sabatieria	10.5	Halalaimus	8.5	Sabatieria	11.7
Subsphaerolaimus	1.9	Sphaerolaimus	7.9	Sphaerolaimus	7.5	Hypodontolaimus	5.0
Aegialoallaimus	0.6	Daptonema	6.9	Sabatieria	6.6	Sphaerolaimus	5.0
Metalinhomoeus	0.5	Terschellingia	6.8	Pomponema	5.9	Oxystomina	3.3
Axonolaimus	0.5	Dorylaimopsis	4.6	Microlaimus	3.9	Ťricoma	3.3
Parasphaerolaimus	0.4	Campylaimus	4.5	Tricoma	3.2	Scaptrella	3.3
Metadesmolaimus	0.4	Halaphanolaimus	2.1	Acantholaimus	3.0	Microlaimus	3.3



Fig. 1. Vertical profiles of copepods at the four stations (average \pm SD). Dashed lines indicate the oxygen penetration depth.

sities were significantly different between stations (one-way ANOVA, p < 0.05) but copepod densities were not significantly different between stations. There was a positive correlation between nematode densities and chlorophyll *a* and POC content ($r^2 = 0.866$ and 0.967 respectively, p < 0.05). In contrast, copepod densities showed no significant correlation with either chlorophyll *a* or POC content.

The 10 most dominant nematode genera found in the upper 5 cm are listed in Table 3. Generic diversity was the lowest at the richest station (station 11) with a strong dominance of the genus *Sabatieria*, that constituted 81 % of the assemblage at station 11 but only 7-12 % at the other stations (Table 3).

The vertical distribution of the two dominant meiobenthic taxa (nematodes and copepods) showed two distinct patterns. Peak densities of copepods were found in the surface layers or deeper in the case of a deeper penetration of oxygen (Fig. 1, one-way ANOVA, p < 0.05). However, for nematodes no relationship with oxygen penetration depth was evident. In spite of different oxygen penetration depths, stations 11 and 4 had similar, significantly deeper weighted mean depths than both stations 2 and 5 (Fig. 2, ANOVA, p < 0.05). Deepest oxy-



Fig. 2. Vertical profiles of nematodes at the four stations (average \pm SD). Dashed lines indicate the oxygen penetration depth.

gen penetration depth (73 mm) was found at the deep-sea site (1039 m water depth, station 5) and the shallowest at the coastal sites (station 11 & 2). Station 4 (89 m water depth) had an intermediate oxygen penetration depth of 11 mm.

Station 11 had the largest biomass of macrofauna (Table 1) and although maximum numbers are found within the upper 5 cm, larger specimens inhabited deeper sediment layers (Fig. 3a & 3b). The striking subsurface biomass peaks at this station were due solely to the polychaete *Sternaspis* sp. that accounted for 91 \pm 6 % of the total biomass over a depth of 20 cm whilst accounting for only 26 ± 6 % of the total abundance of macrofauna. At the other shallow station (station 2) only 2 specimens of *Sternaspis* sp. were found (accounting for 0.04% of the total biomass) and none at stations 4 and 5. The abundance and biomass at station 4 was lower but more evenly distributed throughout the sediment column (Fig. 3a & 3b). In contrast, at stations 2 and 5 macrofauna biomass and abundance showed a sharp decline with depth (Fig. 3a & 3b) suggesting a relatively lower level of subsurface activity or mixing. The lowest macrofauna biomass was found at the deep-sea site (station 5), where also the lowest





average individual biomass was recorded (3 mg WW compared to 101, 16 and 6 mgWW for individuals at stations 11, 2 and 4 respectively).

There was a clear difference in chlorophyll a profiles between the two shallow stations, with a deeper penetration at station 11 compared to station 2 (Fig. 4). The relatively shallow penetration at station 4 is probably due to the low concentration of chlorophyll a in the surface sediment rather than limited subsurface activity as suggested by macrofauna profiles (Fig. 3a & 3b). No chlorophyll a was detected in sediments of station 5.

DISCUSSION

Although this study is limited to four sites, several trends are evident and in accordance with recent observations. The oxygen penetration depth in sediments is strongly related to the organic matter content, with limited penetration in organically rich coastal sediments such as the northwestern Adriatic Sea (Epping & Helder 1998; Moodley et al. 1998) and deeper penetration in deeper waters characterized by low organic matter input (e.g. Helder & Epping 1995; Soetaert et al. 1996). The absence of free sulphide in organic rich sediments of the northwestern Adriatic Sea has been related to the high reactive Fe content of the sediment (Barbanti et al. 1995; Moodley et al. 1998).

In accordance with earlier studies in the Adriatic Sea, nematodes and copepods were the dominant components of the metazoan meiobenthos (Vidakovic 1988; Tahey et al. 1996; Moodley et al. 1998). However, copepods did not occur in large numbers and their densities were not significantly different between



Fig. 3b. Vertical profiles of macrofauna biomass at the four stations (average \pm SD).

stations. Nematodes are generally the dominant component in fine-grained sediments and a positive correlation was found between food availability and nematode standing stock over 5 or 10 cm sediment depth. This confirms the role of food availability as a factor controlling nematode densities, at least over the broad range of food content examined in this study. However, station 11 which supported the highest densities also had the lowest generic diversity with a strong dominance of Sabatieria (81 %, Table 3). High dominance of this genus is often associated with oxygen depleted sediments (e.g. Jensen 1983; Vincx 1990; Olafsson 1992; Hendelberg & Jensen 1993) but station 2 with a similar oxygen penetration depth had a lower dominance of Sabatieria (11%) and Sabatieria constituted 12 % of the nematode assemblage at station 5 (Table 3). This together with

the high dominance of one polychaete (*Sternaspis*) at station 11 suggests that other, site-specific factors may be more important rather than oxygenation directly. However, trends may be different at the species level as *Sabatieria* spp. exhibits a wide range of ecological preferences (Stevaert et al. 1999).

Although vertical profiles of copepods seem to fit patterns of oxygen penetration depth, this clearly does not apply to nematodes. In spite of the deep penetration of oxygen at the deep-sea station (73 mm, station 5), nematodes were concentrated in the upper layers as was the case at station 2 with a much lower oxygen penetration depth (less than 2 mm). A significantly deeper penetration was found at the other two stations (station 11 & 4) characterized by relatively higher levels of mixing activity (as indicated by macrofauna and chlorophyll *a* pro-





Fig. 4. Chlorophyll a profiles in sediments of the different stations. No chlorophyll a was detected at station 5.

files, Figs. 3 & 4) but shallow oxygen penetration. This clearly demonstrates that the depth of the oxic zone does not necessarily delineate the habitat depth of nematodes and that nematode penetration into the sediment is closely related to mixing activities.

The ability of organisms to alter the properties of the sediment is mainly a function of their activity and size (Wheatcroft et al. 1990) and deep mixing of sediments is mainly due to macrofaunal activity (Aller 1984; Smith 1992; Soetaert et al. 1997; Boudreau 1998). Because of their relatively small size (on average, less than 1 mm long) one can assume the nematodes have only limited impact on the geochemistry of their environment (Soetaert et al. 1997). Consequently, vertical distribution of nematodes will be established as a response to the geochemical properties created by larger organisms (Soetaert et al. 1997), assuming that passive transport of nematodes does not play an important role. Two sediment features that can affect the distribution of higher organisms

are food abundance and oxygen availability; the concentration or presence of which in deeper sediment layers is strongly governed by mixing activities. In organic rich sediments with limited oxygen penetration (as registered by microelectrodes or chemical compounds), a substantial part of nematode assemblages are found below the oxic zone (e.g. Hengelberg & Jensen 1991; Soetaert et al. 1997; Steyaert et al. 1999, this study). However, infaunal habitats are, as a result of macrofaunal activity, not permanently anoxic; macrofauna activity evidently provides intermittent traces of oxygen (e.g. Boudreau & Marinelli 1994). With limited exceptions (Vopel et al. 1996), copepods are generally more sensitive to severe oxygen depletion or anoxia than nematodes (Elmgren 1975; Murrell & Fleeger 1989; Moodley et al. 1997; Modig & Olafsson 1998) which may explain the vertical profiles of copepods. In accordance with earlier observations, oxygenation seems to have a limited direct impact on nematode vertical distribution (Soetaert et al. 1997). Our ob-



Fig. 5. Location of sampling stations.

servations suggest that, provided that sediments are not sulphidic or continuously anoxic, oxygen may not the prime factor governing nematode distribution as it seems to affect copepods.

In a study along a depth gradient in the northeastern Atlantic, Soetaert et al. (1997) found a positive relationship between nematode vertical distribution and the vertical distribution of nitrogen (indicator of food content) in the sediment that was related to differential sediment mixing. Peak densities of nematodes were found in surface sediments at sites having limited mixing activity. Similarly, in our study, vertical profiles of nematodes were related to macrobenthic activity. However, our nematode profiles do not match chlorophyll *a* profiles; maximum values of chlorophyll a were found in the surface layers whereas nematodes had subsurface peaks. However, it should be noted that chlorophyll *a* represents only one pool of labile organic matter. Irrespective, nematodes evidently exploit subsurface conditions created by macrofauna activity.

MATERIALS AND METHODS

The metazoan meiobenthos (permanent + temporary) were studied at four sites in the

Adriatic Sea in April/May 1995 (Table 4). Station 11 was located close to the delta of the river Po, stations 2 and 4 were in the western and middle continental shelf and station 5 in the central basin at the base of the continental slope (Fig. 5). Sediment parameters were measured in sediment cores (perspex cores with 3.6 cm i.d.) that were taken from undisturbed boxcore samples. Cores were sliced on board within 2 hours. Grain size analysis were done with a Malvern Particle Analyser. Porosity was calculated from the water content of the sediment, assuming a density of 2.55 for sediment and 1.003 g cm⁻³ for seawater density. The organic carbon content was analysed with a Carlo Erba NA 1500 CN Analyser according to Nieuwenhuize et al. (1994). The chlorophyll a content was analysed by means of reverse-phase HPLC and the lower limit of detection was 0.015 µg

Table 4. List of stations, position and water depth.

Station Number	Posi	Water Depth (m)	
11	44°58′04″N	12°34′51″E	15
2	44°18′24″N	12°34′39″E	19
4	42°16′40″N	14°54′18″E	89
5	41°53′00″N	17°24′24″E	1039

chlorophyll *a* per g dry sediment. Values of both % POC and chlorophyll *a* were converted to content per area using the porosity and assuming a dry density of 2.55 g cm⁻³ for sediment.

The oxygen penetration depth was determined with glass microelectrodes with a built in reference electrode connected to a Keithley picoammeter. Profiling was done immediately using a micro-manipulator at *in situ* temperature. The lowest consistently repeated value at depth in a subcore was taken as zero oxygen. Sediment was also profiled for sulfide, using a metal microelectrode and a calomel reference electrode, connected to a Radiometer pH/millivolt meter.

For macrofauna, boxcores were subsampled with perspex cores (i.d. 19 cm). Duplicate sediment samples were sliced in 5 cm slices down to 20 cm and then washed over a 0.5 mm sieve. The fraction > 0.5 mm was preserved in borax buffered 4% formalin with rose Bengal. In the laboratory, organisms were sorted and counted under a stereo microscope and categorized in taxonomic groups. Biomass determinations were made by weighing formalin preserved samples (excluding shells of molluscs and tubes of polychaetes and amphipods), after blotting on absorbent paper.

For the meiofauna, duplicate samples (perspex cores with i.d. of 3.6 cm) were taken from boxcores. The sediment was sliced in 1 cm slices and stored in borax buffered 4% formalin with rose Bengal. In the laboratory, the fraction > 38 µm was analysed. Meiofauna was first separated from the sediment by centrifuging with Ludox HS 40. Stained specimens of both permanent and temporary meiofauna were identified to major taxon level and counted. The nematodes in the different layers within the upper 5 cm of single cores were identified to genus level. Fifty nematodes per sediment interval (or all specimens in samples that had low numbers) were picked at random and mounted in glycerine slides.

All data were *ln* transformed before statistical analysis by one-way analysis of variance (ANO-VA) or by linear regression analysis using the MGLH module of SYSTAT (Systat Inc.). In order to compare the vertical distribution, the weighted mean depth was calculated using non-transformed data (Fleeger et al. 1995). couragement and the Marine Biology Section, University of Gent for the facilities available and financial support. Special thanks to Jan Belgers and Eric Slim for field assistance. We are grateful to the Captain and crew of the R.V. Urania for their assistance in the collection of the samples. This work has been undertaken in the framework of the Mediterranean Targeted project (MTP)-Euromarge-AS project. We acknowledge the support from the European Commission's Marine Science and Technology (MAST) Programme and the Netherlands Organization for Scientific Research, Earth-and Life Sciences. This is publication 2709 of the NIOO-CE-MO.

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